## United States Department of Agriculture Food Safety and Inspection Service, Office of Public Health Science

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Title: F	SIS Laboratory	Regulatory San	nple Pathogen Methods Table	and Defin	itions			
	Revision: 1/31/2012 Replaces: NA			Effective: 1/31/2012				
Analyte	Microbiology Laboratory Guidebook Chapter #	Screen Test	<b>Confirmatory Tests</b> (following culturing on tube and plating media; and for <i>E. coli</i> O157:H7 IMS bead capture)		Days to Reporting: Screen Negative	Days to Reporting: Potential + Result	Days to Reporting <b>Presumptive</b> + <b>Result</b>	Days to Reporting <b>Final</b> + <b>Result</b>
E. coli O157:H7	MLG 5A MLG 5	BAX <sup>®</sup> PCR Assay (alternatives: lateral flow devices – RapidChek <sup>®</sup> or Transia <sup>™</sup> )	Serological confirmation:   E. coli O157:H7 latex agglutination test kit (RIM <sup>®</sup> E. coli O157:H7 Latex Test Kit, REMEL)   Biochemical confirmation:   Vitek <sup>®</sup> GN/GNI/GNI Plus Cards (bioMerieux)   Shiga toxin/toxin genes confirmation:   Shiga Toxin test kit [Premier <sup>®</sup> EHEC, cat. # 608096 (Meridian Diagnostics, Inc)] or detection of toxin genes by PCR if needed		Day 2	Day 2 ( <u>Limited</u> distribution)	Day 3	Day 4-6
Non E. coli 0157 STEC	MLG 5B	Multiplex RT PCRs: eae, stx then wzx genes	Latex agglutination   Multiplex RT PCR typical colonies   eae, stx then wzx genes   Biochemical confirmation:   Vitek <sup>®</sup> GN/GNI/GNI Plus Cards (bioMerieux)		Day 2	Day 2 ( <u>Limited</u> distribution)	Day 3	Day 4-6
Listeria nonocytogenes	MLG 8 MLG 8A	BAX <sup>®</sup> PCR Assay	Tumbling Motility observation Biochemical confirmation:MICRO-ID® Listeria, ListeriaAPI®, Vitek 2CAMP/CAMP Factor Test β-lysin CAMP factor discs (Remel) with MICRO-ID®; Genetic Identification Testing if needed for speciation – GenProbe Accuprobe® Ribosomal RNA-based L. monocytogenes-specific test system		Day 3	NA	Day 4-5	Day 5-8

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Title: F	Title: FSIS Laboratory Regulatory Sample Pathogen Methods Table and Definitions							
Revision	Revision: 1/31/2012			Replaces: NA		Effective: 1/	Effective: 1/31/2012	
Analyte	Microbiology Laboratory Guidebook Chapter #	Screen Test	(following culturing on tube and plating media; and for <i>E. coli</i> O157:H7 IMS bead capture)		Days to Reporting: Screen Negative	Days to Reporting: Potential + Result	Days to Reporting <b>Presumptive</b> + <b>Result</b>	Days to Reporting <b>Final</b> + <b>Result</b>
Salmonella spp.	MLG 4 MLG 4C	BAX <sup>®</sup> PCR Assay	Serological confirmation: Somatic(O)Antigen Agglutinat polyvalent O antiserum); Flagellar (H) Antigen Agglut Salmonella Latex Test) Biochemical confirmation:	ination Tests (Oxoid	Day 2	NA	Day 5 NA for HACCP	Day 6-7 Depends on Vitek result available Day 6 PM; Day 7 AM
<i>Campylobacter</i> for Quantitative method	MLG 41	Direct Plating	Vitek <sup>®</sup> GNI/GNI Plus Cards (bi Typical colonies subject to same <u>Microscopic examination</u> Latex agglutination		Day 3	NA	NA	Day 3

\*Table doesn't include additional non regulatory testing (e.g.NVSL serotyping, PFGE subtyping, *Campylobacter* qualitative testing). On Day 1 sample arrives in the laboratory. Days listed do not include delays (e.g. restreak for purity, rare strains requiring additional testing in Outbreak Section Eastern Laboratory).

## **Definitions:**

Potential positive E. coli O157:H7 – Enrichment medium from one or more subsamples yields a positive when screen tested.

Presumptive positive E. coli O157:H7 – One or more typical colonies on plating agar agglutinate when tested with O157 antiserum.

Confirmed positive *E. coli* O157:H7 – One or more isolates from the sample is a biochemically identified *Escherichia coli* that is serologically or genetically determined to be "O157" that meets at least one of the following criteria:

- 1) Positive for Shiga toxin (ST) production
- 2) Positive for Shiga toxin gene(s) (stx)
- 3) Genetically determined to be "H7"

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Potential positive non *E. coli* O157 STEC – Enriched medium from a sample which yields a positive when screened on real-time PCR for each of the targeted genes (*eae*, stx1/2, and wzx) of one or more of six non-O157 serogroups (O26, O45, O103, O111, O121, O145).

Presumptive positive non E. coli O157 STEC – One or more typical colonies on modified Rainbow agar. Latex agglutination positive.

Confirmed positive non *E. coli* O157 STEC – Latex agglutination positive on Sheep Blood Agar, one or more isolates from the sample is confirmed positive on real-time PCR for the *eae*, *stx*, and *wzx* genes of one or more of six non-O157 serogroups, and biochemically identified as *Escherichia coli*.

Presumptive positive *L. monocytogenes* – A sample from which one or more typical colonies produces beta hemolysis on Horse Blood Agar.

Confirmed positive *L. monocytogenes* – A beta hemolytic isolate is Camp test positive, shows tumbling motility (optional) and is characterized biochemically as *L. monocytogenes*. Ribosomal RNA testing is occasionally required to resolve atypical strains.

Presumptive positive *Salmonella* spp. – A sample yields one or more isolates which show typical appearance on TSI and LIA slants and agglutinate salmonella somatic antisera.

Confirmed positive Salmonella spp. - Salmonella O group positive isolates are characterized biochemically as Salmonella spp.

Confirmed positive *Campylobacter* – Typical colony morphology, microscopic ID, latex agglutination positive for *C. jejuni, C. coli, and/or C. lari.*