# DTP, DCTD TUMOR REPOSITORY

A CATALOG OF IN VITRO CELL LINES, TRANSPLANTABLE
ANIMAL AND HUMAN TUMORS AND YEAST

**Operated by Charles River Laboratories, Inc.
under contract to the Biological Testing Branch for
The Frederick National Laboratory for Cancer Research
Frederick, Maryland 21702-1201**

Sponsored by:

Biological Testing Branch
Developmental Therapeutics Program
Division of Cancer Treatment and Diagnosis
National Cancer Institute
National Institutes of Health

www.dtp.nci.nih.gov

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# DCTD Tumor Repository
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# INTRODUCTION

The Division of Cancer Treatment and Diagnosis (DCTD), National Cancer Institute, has maintained since the early 1960s a low temperature repository of transplantable *in vivo*-derived tumors and *in vitro*-established tumor cell lines from various species. Currently located at the National Cancer Institute at Frederick in Frederick, Maryland, the DCTD Tumor Repository* serves as a resource for viable, contaminant-free experimental tumor lines, many of which are not obtainable elsewhere. The Repository makes these materials available to qualified investigators as a service to the research community.

The Repository's tumor collection contains a wide variety of frozen types of human and animal origin. Virtually all of the human tumors are xenografts grown in athymic nude mice, although there are some that grow in conditioned rats or in hamster's cheek pouch. Several mouse leukemia lines in the collection are resistant to single drugs of varying modes of action. Multidrug-resistant lines are also available. In addition, the collection includes variant sublines of B16 melanomas that exhibit a different degree of metastasis to various organs.

The tumors in this catalog are categorized by species, namely, human, hamster, guinea pig, mouse, rabbit and rat. Within animal species, the list is in alphabetical order by tumor designation. Tumors with numeric designations are listed at the end. Human tumors are grouped by tumor type.

We request that the DCTD Tumor Repository, National Cancer Institute at Frederick, Frederick, Maryland, be cited in publications as the source of tumor materials. We also request that reprints of publications be furnished to the Repository.

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*Previous contract locations of the Tumor Repository include Microbiological Associates, Inc., Bethesda, MD; Arthur D. Little, Inc., Cambridge, MA; and Mason Research Institute, Worcester, MA.

## DTP, DCTD, NCI Repository Ordering Procedures - Tumor Fragments, Cell Lines and Yeast

Tumor materials are furnished to qualified investigators affiliated only with recognized research laboratories.
Items 1-3 below are required in order to receive material.

- **Send a Letter** of Request on official institutional letterhead.
- Provide a Purchase Order or **method of payment** (see purchase procedures).
- Send the **Material Transfer Agreement (MTA) with signatures affixed from both the requestor and authorizing official** (required by NCI). Please print or type this document. Request will be processed after receipt of all three items above. Legible mailed or e-mailed MTAs are accepted, please no faxed MTAs. Send paperwork to:

**DOMESTIC request**
Ms. Katherine Gill
BTB, DTP, DCTD, NCI
Frederick National Laboratory
for Cancer Research, Box B
Bldg. 1043, 7
Frederick, MD 21702
E: gillk@mail.nih.gov
P: 301-846-5483

**INTERNATIONAL request**
Ms. Christine Pacula-Cox
BTB, DTP, DCTD, NCI
Frederick National Laboratory
for Cancer Research, Box B
Bldg. 1043, 9
Frederick, MD 21702
E: paculac@mail.nih.gov
P: 301-846-1709

The Letter should briefly discuss either the requested material, research project or indicate method of payment. Multiple cell lines can be ordered using a single MTA. Clearly identify each item under #1 on the MTA or use an addendum page. Newly requested material and MTA renewals will require completion of a new MTA and are active for a period of three years.

Re-orders by the same investigator for previously received material should indicate the active MTA number in an email or letter of request, along with method of payment information.

The following fees include shipping in those cases where standard delivery methods can be used.
NOTE: *The rising cost of international transportation necessitates the addition of a shipping recovery fee of $150.00 to International recipients to help defray shipping costs. Exception: Canadian surcharge is $100.00.*

| | NCI/NIH Investigators & Fed. Government (at MD campuses only) | Academia & Non-profits both Domestic & International | Commercial entities both Domestic & International |
|---|---|---|---|
| **Cell or Tumor Lines** | | | |
| Per cryopreserved vial | N/A | $150.00 | $200.00 |
| NCI Anti-cancer cell line panel | N/A | $6,700.00 | $8,850.00 |

*The complete NCI panel is 60 cell lines. Commercial entities may receive the five NCI-H lines only through a licensing agreement therefore invoice cost for the panel will be adjusted to 55 lines.*

| **Yeast Strains:** | NCI/NIH (MD campuses only) & Fed. Gov't. | Academia & Non-profits Domestic/International | Commercial entities Domestic & International |
|---|---|---|---|
| One strain | N/A | $150.00 | $200.00 |
| Complete set (16 strains) | N/A | $1,800.00 | $2,400.00 |

International Recipients: The DCTD Tumor Repository obtains U.S. Exportation Declarations for each shipment. Recipients must obtain the applicable Import permits as required by the recipient's country**.** Email the Import document or permit number so we may apply for the Export document. Retaining a Broker to get your shipment through Customs is strongly suggested.

In cases where standard shipping methods are not applicable, the recipient will be contacted to cover all excess charges associated with shipping. Please contact NCI in advance to determine your eligibility for standard shipping rates. Material will be sent freight prepaid.

Other contact numbers:
Scientific & International request - Christine Pacula-Cox, NCI **P**: 301 846-1709 **E**: **paculac@mail.nih.gov**
Availability, W-9 & Invoicing - Jane Shelton, CRL **P**: 301 846-5748 **F**: 301-846-6941 :**sheltonmj@mail.nih.gov**
Shipping questions - Vicky Clark, CRL **P**: 301 846-7003 **F**: 301 846-6941 **E**: **clarkvj@mail.nih.gov**

# DTP, DCTD, NCI Tumor Repository Purchasing/Payment Procedures

**Cell Lines, Fragment Tissue, Yeast**

**Domestic Purchase Order To:**

**Charles River Labs.**
**c/o Kathy Gill**
Frederick National Laboratory
for Cancer Research, Box B
Bldg. 1043 Rm. 7
Frederick, MD 21702
E-mail: **gillk@mail.nih.gov**
Phone: 301-846-5483

**International Purchase Order To:**

**Charles River Labs.**
**c/o Christine Pacula-Cox**
Frederick National Laboratory
for Cancer Research, Box B
Bldg. 1043, Rm. 9
Frederick, MD 21702
E-mail: **paculac@mail.nih.gov**
Phone: 301-846-1709

**Payment goes to**:

Charles River Laboratories, Inc
**GPO Box 27812**
**New York, NY 10087-7812**

CRL Federal Tax ID # 76-0509980

**Charles River Labs is our contractor who will ship and invoice the research material on behalf of the NCI.**

CRL accepts the following credit cards for remittance: VISA, MASTERCARD and AMERICAN EXPRESS. Please provide the details when using a credit card. Purchase orders, purchase order numbers, checks and wire transfers are also accepted. **If using a P.O. please send a copy by e-mail.**

For invoice or W-9 questions please contact Ms. Jane Shelton at **Sheltonmj@mail.nih.gov**
Phone: 301-846-5748 or Fax: 301-846-6941

**Please note:** **All shipments from the DCTD Repository will be made by FedEx or World Courier without exception.**

**INTERNATIONAL:**

The rising cost of international transportation necessitates the addition of a shipping recovery fee of $150.00 to International recipients to help defray shipping costs. Exception: Canadian surcharge is $100.00. These fees apply in cases where standard delivery methods can be used.

**ELECTRONIC FUNDS WIRE TRANSFER INFORMATION EFT/ACH WIRES**

**JP Morgan**
**New York, NY**
**ABA Number: 021000021**
**Account of: Charles River Laboratories Lock Box Account**
**DDA (Checking) Account Number: 799-761499**
**Swift Number: CHASUS33**
**ACH HELP DESK: (800) 447-3593**
**WIRE HELP DESK: (866) 223-0359**
***** Please note: JPM uses the same ABA Number for both wires and ACH Payments***
**CTX Format required to utilize EFT/ACH**

**The full invoice amount is to be paid; any bank wire fees incurred are at the purchaser's expense.**

NCI MTA #1-

# National Cancer Institute

## MATERIAL TRANSFER AGREEMENT-A
## Cell lines maintained in the NCI-DCTD Repository

This Material Transfer Agreement ("MTA") has been adopted for use by the National Cancer Institute ("NCI") for transfers of cell lines from the Division of Cancer Treatment and Diagnosis ("DCTD") Tumor Repository ("Research Material"). The DCTD Tumor Repository has maintained, since the early 1960's, a low temperature repository of transplantable tumor and tumor cell lines from various species. The Repository serves as a resource for experimental tumor lines from various species, many of which are not obtainable elsewhere. The Repository makes these Materials available as a service to the Research Community.

Recipient: _____
Name of Recipient Investigator and Recipient Institution

1. NCI agrees to transfer to Recipient named above the following Research Material:

_____
(use an attachment page if necessary)

2. THIS RESEARCH MATERIAL MAY NOT BE USED IN HUMAN SUBJECTS. The Research Material will only be used for research purposes by Recipient's investigator in his/her laboratory, for the research project described below, under suitable containment conditions. This Research Material will not be used for commercial purposes such as production or sale. Recipient agrees to comply with all Federal rules and regulations applicable to the Research Project and the handling of the Research Material. These samples are being provided in a manner that does not allow for direct identifiable patient information to the Recipient, and therefore do not constitute Human Subject Research as defined in 45 CFR Part 46, “Protection of Human Subjects”.

3. This Research Material will be used by Recipient's investigator solely in connection with the following research project ("Research Project") described with specificity as follows (use an attachment page if necessary):

_____

_____

4. In all oral presentations or written publications concerning the Research Project, Recipient will acknowledge NCI's contribution of this Research Material unless requested otherwise. To the extent permitted by law, Recipient agrees to treat in confidence, for a period of three (3) years from the date of its disclosure, any of NCI's written information about this Research Material that is stamped **"CONFIDENTIAL,"** except for information that was previously known to Recipient or that is or becomes publicly available or which is disclosed to Recipient without a confidentiality obligation. Any oral disclosures from NCI to Recipient shall be identified as being **CONFIDENTIAL** by notice delivered to Recipient within ten (10) days after the date of the oral disclosure. Recipient may publish or otherwise publicly disclose the results of the Research Project, but if NCI has given **CONFIDENTIAL** information to Recipient such public disclosure may be made only after NCI has had thirty (30) days to review the proposed disclosure to determine if it includes any **CONFIDENTIAL** information, except when a shortened time period under court order or the Freedom of Information Act pertains.

5. This Research Material represents a significant investment on the part of NCI. Recipient's investigator therefore agrees to retain control over this Research Material and further agrees not to transfer the Research Material to other people not under her or his direct supervision without advance written approval of NCI. NCI reserves the right to distribute the Research Material to others and to use it for its own purposes. When the Research Project is completed or three (3) years have elapsed, whichever occurs first, the Research Material will be disposed of as directed by NCI.

NCI MTA #1-

6. This Research Material is provided as a service to the research community. IT IS BEING SUPPLIED TO RECIPIENT WITH NO WARRANTIES, EXPRESS OR IMPLIED, INCLUDING ANY WARRANTY OF MERCHANTABILITY OR FITNESS FOR A PARTICULAR PURPOSE. NCI makes no representations that the use of the Research Material will not infringe any patent or proprietary rights of third parties.

7. Recipient may retain title to the patent rights in inventions made by its employees in the course of the Research Project. Recipient agrees not to claim, infer, or imply Governmental endorsement of the Research Project, the institution or personnel conducting the Research Project or any resulting product(s). Unless prohibited by law from doing so, recipient agrees to hold the United States Government harmless and to indemnify the Government for all liabilities, demands, damages, expenses and losses arising out of Recipient's use for any purpose of the Research Material.

8. The undersigned Recipient expressly certifies and affirms that the contents of any statements made herein are truthful and accurate.

9. This MTA shall be construed in accordance with Federal law as applied by the Federal courts in the District of Columbia.

Date: _____ _____

Recipient **Investigator's Signature and Title**

Date:_____ _____

**Authorized Signature and Title**, for Recipient's Institution (**Note**: Authorized Signature has the authority to bind the Institution to the terms of this Material Transfer Agreement).

→ Recipient's Shipping Address: Recipients Billing Address (if different):

→Recipient's Phone, Fax, E-mail:

Authorized Signature for NCI: _____ Date: _____

**Melinda G. Hollingshead, D.V.M., Ph.D.,** Chief, BTB, DTP, DCTD, NCI

NCI's Mailing Address:

Email: **gillk@mail.nih.gov**

Ms. Katherine Gill
BTB/DTP/DCTD/NCI
Frederick National Laboratory
for Cancer Research, Box B
Bldg. 1043, Rm. 7
Frederick, MD 21702

Phone: 301-846-5483

## SHIPMENT OF TUMORS

The transplantable tumors are distributed as frozen vials of tumor tissues or cell suspension. Transplantable tumors as well as cell culture lines are shipped as frozen vials of tissues on dry ice. Each tumor shipment includes an information sheet showing, among other items, the proper tumor designation, cryopreserved date, *in vivo* host, etc.

Requested tumors are shipped two to three weeks after receipt of all completed paperwork. Domestic shipments leave the Repository no later than Wednesday in order to reach their destination on week days. Foreign shipments are sent according to final destination. Before shipment, the Repository notifies the Recipient of the waybill number and carrier information by e-mail or fax. Recipient must notify the Repository or make arrangements for receipt of the tumor lines, in the event that they will not be available to receive the shipment. An invoice for payment will follow and payment is due upon receipt.

## SUBMISSION OF TUMORS FOR CRYOPRESERVATION

Investigators who have unique and novel experimental tumor lines and wish to submit their tumors to the Repository for cryopreservation and storage should write a letter of intent to the Project Officer. Upon acceptance, the Project Officer will inform the investigator in writing, and will provide instructions on the procedure for shipping tumor materials to the Repository. Tumor tissues or cells (frozen or ambient) are preferred over tumor- bearing animals.

At the Repository, the tumor line(s) will be tested for viral and bacterial contamination. When proven "clean," the line(s) will be expanded, *in vivo* or *in vitro* as appropriate, for large batch cryopreservation. Viability and growth of the frozen tumors will be evaluated. The tumors will be included in the Repository's inventory, and upon joint approval of the submitting investigator and the Project Officer, they will be made available for distribution to the scientific community.

## MOUSE TUMORS FROM THE JACKSON LABORATORY

These tumors formerly were maintained and distributed by the Jackson Laboratory. The list of available tumors begins on page 39 of the DCTD Tumor Repository Catalog. They were cryopreserved at EG&G Mason Research Institute and are distributed only as vials of frozen tumor tissue. The required host animals for carrying the JAX tumors in serial transplantation may be obtained from:

Animal Resources
The Jackson Laboratory
600 Main Street
Bar Harbor, ME 04609
USA
T: 800.422.MICE
207.288.5845
F: 207.288.6150

## FREEZING PROCEDURE

Solid tumors are frozen as 2 x 2 x 2 mm fragments suspended in a freezing medium. Ascites or tissue culture cells are frozen as single cell suspension at a concentration of $10^6$-$10^7$ cells per ml. The freezing medium consists of appropriate tissue culture growth medium plus 10% DMSO and 10% fetal bovine serum.

Aseptically harvested ascites tumors are diluted in the freezing medium at a concentration of $10^6$-$10^7$ cells per ml. One ml suspension is pipetted into each 2 ml vial (Nunc cryotube). The vials are screw-capped tightly and labeled with Repository number. Tissue culture cells are prepared in a similar manner. For solid tumors, the aseptically excised tumor tissue is cut into 2 x 2 x 2 mm fragments after freeing it of necrotic materials. The fragments are placed in vials containing 1.5 ml of freezing medium.

The processed tumors are frozen initially in a controlled slow-rate freezing apparatus at the rate of 0.5°C per minute to -20°C and 1°C per minute to -80°C. The frozen vials are stored in specific locations in the liquid nitrogen freezers in the Repository after the controlled freezing cycle.

## RECOMMENDED THAWING PROCEDURE

Frozen tumor cells or tissues received from the Repository should be kept frozen at -70°C or lower until ready for use. For prolonged storage (more than two days), liquid nitrogen freezers are recommended.

The vials in which the cell lines are stored are reliable; however, they are very susceptible to contamination if thawed in a contaminated water bath. Thawing should be rapid, i.e., within 60-90 seconds. Place the vial in a warm water bath at 37-40°C and agitate vigorously to thaw. Immerse the vial in 70% ethanol before uncapping. Implant immediately after thawing. The concentration of DMSO is not toxic to tissues and implantation may be made directly from the vial.

For tissue culture, transfer the contents of the vial into a petri dish or a flask containing at least 10 volumes of the recommended culture medium and incubate. In order to remove the protective freezing additive (DMSO) from the culture medium, we suggest that the culture medium be changed 24 hours after seeding. If it is desired that DMSO be removed immediately, centrifuge the diluted suspension at approximately 125 x g for 10 minutes, discard the supernatant, and resuspend the cells in an appropriate volume of growth medium.

CAUTION: We strongly recommend wearing protective glasses or face shields when thawing tissues in glass vials.

TUMOR TRANSPLANTATION: It is recognized that transplantable tumor systems are experimental tools for investigators in scientific disciplines other than tumor biology or transplantation immunogenetics. Therefore, we encourage investigators with limited transplantation experience to contact the Tumor Repository for more detailed information on techniques.

## BACTERIAL AND VIRAL MONITORING OF CRYOPRESERVED TISSUES

In addition to testing all freeze-runs for bacterial contamination, cryopreserved tissues are tested for viral contamination by the MAP test. The viruses tested for are as follows: pneumonia virus of mice (PVM), reo virus-type 3 (Reo 3), Theiler's virus, murine encephalitis (GD VII), polyoma (Poly), minute virus of mice (MVM), mouse hepatitis virus (MHV), lymphocytic choriomeningitis (LCM), Sendia virus (Send), ectromelia, mouse pox (Ectro), lactic dehydrogenase virus (LDH).

## *IN VITRO* ESTABLISHED CELL LINES:

A. ***Quality Control and Characterization*** - The quality control and characterization procedures for the incorporation of new cell lines into the Tumor Repository are as follows: Upon receipt, each cell line is immediately transferred to fresh antibiotic-free medium and cultured for one week, after which it is tested for mycoplasma (PPLO) contamination. Standard culture procedures under aerobic and anaerobic conditions, as well as the orcein staining procedure of Fogh, are used. The PPLO medium is extremely rich, and this procedure will also detect most bacterial and fungal contaminants.

B. ***Freezing and Storage*** - The cell cultures are frozen in ampules containing 1.0 ml of cell suspension at 2-6 x $10^6$ cells/ml in fresh culture medium containing 10% DMSO. Freezing is performed as on page 6. Twenty-four hours after freezing, a representative ampule is removed, thawed, and viable cell count is performed, using the trypan blue dye exclusion procedure. The culture is also tested for its ability to initiate a heavy viable culture. Cell preparations which show less than 50% viability or poor growth are discarded and a new lot is prepared.

C. ***Recommended Procedure for Thawing Frozen Cell Cultures*** - The vials in which the cell lines are stored are reliable; however, they are very susceptible to contamination if thawed in a contaminated water bath. The following procedures are provided to eliminate this problem.

Remove the ampule from the dry ice container and place it directly into a 37-40°C water bath (or vessel) of freshly drawn water containing an effective concentration of disinfectant. The thawing should be vigorous and rapid (within 40-60 seconds). As soon as the thawing is complete, remove the ampule from the water bath and immerse in 70% ethanol at room temperature. All of the operations from this point

should be carried out under strict aseptic conditions in a sterile room, cubicle, or hood.

Transfer the contents of the ampule (1 ml volume) into a 100 mm petri dish or 25 $cm^2$ flask containing 8-10 ml of the recommended culture medium, and incubate at the appropriate temperature and carbon dioxide level.

In order to remove the protective freezing additive (DMSO) from the culture medium, we suggest that the culture medium be changed 24 hours after thawing. If it is desired that the freezing additive be removed immediately or that a more concentrated cell suspension be obtained, centrifuge the above diluted suspension at approximately 125 x g for 10 minutes, discard the fluid, and resuspend the cells in an appropriate volume of growth medium.

# HUMAN TUMORS

| SPECIES: HUMAN | | | |
|---|---|---|---|
| **Tumor Designation** | **Histologic Type** | **General Information** | **Species and/or Strain of Transplantability** |
| | | **BREAST** | |
| COO-G | Mammary Carcinoma | Primary explant established *in vivo* in athymic nude mice by Dr. B. Giovanella, Stehlin Foundation. | Nude Athymic Mice |
| DU4475 | Mammary Carcinoma | Primary explant from cutaneous tumor nodule in region of mastectomy established *in vitro* by Dr. A.J. Langlois, Duke University Medical Center; then adapted to *in vivo* transplantation by Dr. A.E. Bogden. | Nude Athymic Mice |
| ELL-G | Mammary Carcinoma | Primary explant established *in vivo* in athymic nude mice by Dr. B. Giovanella, Stehlin Foundation. | Nude Athymic Mice |
| HIG-G | Mammary Carcinoma | Primary explant established *in vivo* in athymic nude mice by Dr. B. Giovanella, Stehlin Foundation. | Nude Athymic Mice |
| MCF/7 | Mammary Carcinoma | Primary explant from pleural effusate established *in vitro* by Dr. H.D. Soule, Michigan Cancer Foundation; then adapted to *in vivo* transplantation by Dr. A.E. Bogden. | Nude Athymic Mice |
| MDA-MB-436 | Mammary Carcinoma | Primary explant from pleural effusate established *in vitro* by Dr. Relda Cailleau, M.D. Anderson Hospital and Tumor Institute; then adapted to *in vivo* transplantation by Dr. A.E. Bogden. | Nude Athymic Mice |
| MX-1 | Mammary Carcinoma | Primary xenotransplant from an infiltrating duct carcinoma (CLO-G). Adapted to *in vivo* transplantation by Dr. B. Giovanella. | Nude Athymic Mice |
| MX-2 | Mammary Carcinoma | Adapted to *in vivo* from *in vitro* established pleural effusate. | Nude Athymic Mice |
| SW-613 | Mammary Carcinoma | Established from *in vitro* line. | Nude Athymic Mice |
| VAN-G | Mammary Carcinoma | Primary explant established *in vivo* in athymic nude mice by Dr. B. Giovanella, Stehlin Foundation. | Nude Athymic Mice |
| | | **LUNG** | |
| LX-1 | Lung, undifferentiated carcinoma | Xenotransplant from a metastasis to subcutaneous tissue (DOY-G). The primary lung tumor was an oat cell carcinoma. Adapted to *in vivo* transplantation by Dr. B. Giovanella. | Nude Athymic Mice |
| COS-G | Lung, papillary carcinoma | Xenotransplant adapted to *in vivo* transplantation by Dr. B. Giovanella. | Nude Athymic Mice |
| | | | |

| SPECIES: HUMAN | | | |
|---|---|---|---|
| **Tumor Designation** | **Histologic Type** | **General Information** | **Species and/or Strain of Transplantability** |
| | | **LUNG (continued)** | |
| H-MESO-1 | Lung, mesothelioma | Xenotransplant from a primary tumor received from Dr. R.M. Williams and Dr. A. Rossini. Adapted to *in vivo* transplantation by Dr. A.E. Bogden. | Nude Athymic Mice |
| H-MESO-1A | Lung, mesothelioma | H-MESO-1 converted to ascites form by Dr. A.E. Bogden. | Nude Athymic Mice |
| NCI-H23<br>H23 | Lung, nonsmall cell, adenocarcinoma | Obtained from Dr. Adi Gazdar. | Nude Athymic Mice |
| NCI-H460<br>H460 | Lung, nonsmall cell, epid. | Obtained from Dr. Adi Gazdar. | Nude Athymic Mice |
| MRI-H-165 | Lung, squamous cell carcinoma | Primary xenotransplant adapted to *in vivo* transplantation by Dr. A.E. Bogden. | Nude Athymic Mice |
| MRI-H-266 | Lung, poorly differentiated carcinoma | Xenotransplant from a metastasis, adapted to *in vivo* transplantation by Dr. A.E. Bogden. | Nude Athymic Mice |
| | | **COLON** | |
| CX-1 (HT-29) | Colon, adenocarcinoma | Primary, untreated adenocarcinoma (HT-29) adapted to *in vitro* cell culture by Dr. Jergen Fogh; then adapted to *in vivo* transplantation by Dr. A.E. Bogden. Secretes CEA. | Nude Athymic Mice |
| CX-2 | Colon, carcinoma | Primary xenotransplant (CA-1) adapted to *in vivo* transplantation by Dr. B. Giovanella. Received at MRI from Dr. A. Ovejera. | Nude Athymic Mice |
| CX-3 | Colon, carcinoma | | Nude Athymic Mice |
| CX-5 | Colon, adenocarcinoma | Xenotransplant from an untreated metastasis (SQU-G) adapted to *in vivo* transplantation by Dr. B. Giovanella. | Nude Athymic Mice |
| GOB-G | Colon, adenocarcinoma | Xenotransplant adapted to *in vivo* transplantation by Dr. B. Giovanella. | Nude Athymic Mice |
| HCC-2998 | Colorectal carcinoma | Obtained from Dr. I.J. Fidler. | Nude Athymic Mice |
| HCT-15 | Colon, carcinoma | Established from *in vitro* line. | Nude Athymic Mice |
| KLO-G | Colon, adenocarcinoma | Xenotransplant adapted to *in vivo* transplantation by Dr. B. Giovanella. | Nude Athymic Mice |
| KM20L2 | Colon, adenocarcinoma | Obtained from Dr. I.J. Fidler. | Nude Athymic Mice |
| MRI-H-194 | Colon, adenocarcinoma | Xenotransplant from a metastasis adapted to *in vivo* transplantation by Dr. A.E. Bogden. | Nude Athymic Mice |
| LOVO I | Colon, adenocarcinoma | Established from *in vitro* line. | Nude Athymic Mice |

| SPECIES: HUMAN | | | |
|---|---|---|---|
| **Tumor Designation** | **Histologic Type** | **General Information** | **Species and/or Strain of Transplantability** |
| **COLON (continued)** | | | |
| LOVO II | Colon, adenocarcinoma | Established from *in vitro* line. | Nude Athymic Mice |
| MRI-H-250 | Colon, carcinoma | Xenotransplant from a metastasis adapted to *in vivo* transplantation by Dr. A.E. Bogden. | Nude Athymic Mice |
| **MELANOMA** | | | |
| BOW-G | Melanosarcoma | Xenotransplant from metastasis adapted to *in vivo* transplantation by Dr. B. Giovanella. | Nude Athymic Mice |
| DEAC-7 | Melanoma | Received as cryopreserved ampules from Dr. S. Warren and Dr. W.B. Patterson. | Hamster Cheek Pouch |
| FO #1 | Melanoblastoma | Adapted to *in vivo* transplantation (FOS-G) by Dr. B. Giovanella. | Nude Athymic Mice |
| NIS-G | Melanosarcoma | Xenotransplant adapted to *in vivo* transplantation by Dr. B. Giovanella. | Nude Athymic Mice |
| TRI-G | Melanoma | Xenotransplant adapted to *in vivo* transplantation by Dr. B. Giovanella. | Nude Athymic Mice |
| WIL-G | Melanoma | Xenotransplant adapted to *in vivo* transplantation by Dr. B. Giovanella. | Nude Athymic Mice |
| MRI-H-121B | Melanoma, malignant | Primary xenotransplant adapted to *in vivo* transplantation by Dr. A.E. Bogden. | Nude Athymic Mice |
| MRI-H-187 | Melanoma, epithelioid, melanotic | Xenotransplant from metastasis adapted to *in vivo* transplantation by Dr. A.E. Bogden. | Nude Athymic Mice |
| MRI-H-221 | Melanoma, malignant | Xenotransplant from metastasis adapted to *In vivo* transplantation by Dr. A.E. Bogden. | Nude Athymic Mice |
| MRI-H-255 | Melanoma | Xenotransplant adapted to *in vivo* transplantation by Dr. A.E. Bogden. | Nude Athymic Mice |
| **CERVIX** | | | |
| MRI-H-130 | Cervix, squamous cell carcinoma | Primary xenotransplant adapted to *in vivo* transplantation by Dr. A.E. Bogden. | Nude Athymic Mice |
| MRI-H-177 | Cervix, squamous cell carcinoma | Xenotransplant from a metastasis adapted to *In vivo* transplantation by Dr. A.E. Bogden. | Nude Athymic Mice |
| MRI-H-186 | Cervix, invasive, large cell, nonkeratinizing, squamous cell carcinoma | Primary xenotransplant adapted to *in vivo* transplantation by Dr. A.E. Bogden. | Nude Athymic Mice |
| MRI-H-196 | Cervix, poorly differentiated squamous cell carcinoma | Primary xenotransplant adapted to *in vivo* transplantation by Dr. A.E. Bogden. | Nude Athymic Mice |

| SPECIES: HUMAN | | | |
|---|---|---|---|
| **Tumor Designation** | **Histologic Type** | **General Information** | **Species and/or Strain of Transplantability** |
| | | **CERVIX (continued)** | |
| MRI-H-215 | Cervix, invasive, large cell, nonkeratinizing, poorly differentiated, epidermoid carcinoma | Primary xenotransplant adapted to *in vivo* trans-plantation by Dr. A.E. Bogden. | Nude Athymic Mice |
| | | **KIDNEY** | |
| MRI-H-121 | Kidney, carcinoma | Xenotransplant from a metastasis adapted to *in vivo* transplant by Dr. A.E. Bogden. | Nude Athymic Mice |
| MRI-H-166 | Kidney, transitional cell carcinoma | Primary xenotransplant adapted to *in vivo* trans-plantation by Dr. A.E. Bogden. | Nude Athymic Mice |
| | | **ENDOMETRIUM** | |
| MRI-H-147 | Endometrium, carcinoma, Müllerian duct | Primary xenotransplant adapted *in vivo* trans-plantation by Dr. A.E. Bogden. | Nude Athymic Mice |
| MRI-H-171 | Endometrium, carcinoma | Primary xenotransplant adapted to *in vivo* trans-plantation by Dr. A.E. Bogden. | Nude Athymic Mice |
| MRI-H-220 | Endometrium, carcinoma | Primary xenotransplant adapted to *in vivo* trans-plantation by Dr. A.E. Bogden. | Nude Athymic Mice |
| MRI-H-253 | Endometrium, carcinoma | Primary xenotransplant adapted to *in vivo* trans-plantation by Dr. A.E. Bogden. | Nude Athymic Mice |
| | | **OVARY** | |
| MRI-H-207 | Ovary, undifferentiated carcinoma | Primary xenotransplant adapted to *in vivo* trans-plantation by Dr. A.E. Bogden. | Nude Athymic Mice |
| MRI-H-227 | Ovarian Adenocar-cinoma | Primary explant established *in vivo* by Dr. A.E. Bogden. Received from Dr. R. Hunter, University of Massachusetts Medical School. | Nude Athymic Mice |
| MRI-H-258 | Ovarian Adenocar-cinoma | Primary explant established *in vivo* by Dr. A.E. Bogden. Received from Dr. R. Hunter, University of Massachusetts Medical School. | Nude Athymic Mice |
| MRI-H-273 | Ovarian Carcinoma | Originated from metastasis. Established *in vivo* by Dr. A.E. Bogden. Received from New England Deaconess Hospital. | Nude Athymic Mice |
| MRI-H-1834 | Ovarian Carcinoma | Primary explant established *in vivo* by Dr. A.E. Bogden. Received from Dr. R. Hunter, University of Massachusetts Medical School. | Nude Athymic Mice |
| SWA-G | Ovarian Carcinoma | Xenotransplant adapted to *in vivo* transplantation by Dr. B. Giovanella. | Nude Athymic Mice |
| | | | |

| SPECIES: HUMAN | | | |
|---|---|---|---|
| **Tumor Designation** | **Histologic Type** | **General Information** | **Species and/or Strain of Transplantability** |
| | | **SARCOMA** | |
| HS-1 | Sarcoma | No historical information available. | Conditioned Rats |
| OGL-G | Sarcoma, spindle cell, periosteal osteogenic | Primary xenotransplant adapted to *in vivo* transplantation by Dr. B. Giovanella. | Nude Athymic Mice |
| DEL-G | Sarcoma | Primary xenotransplant adapted to *in vivo* transplantation by Dr. B. Giovanella. | Nude Athymic Mice |
| | | **EPIDERMOID** | |
| HEP-3 | Epidermoid carcinoma | No historical information available. | Conditioned Rats |
| DEAC-1 | Mucoepidermoid carcinoma | Received as cryopreserved ampules from Dr. S. Warren and Dr. W.B. Patterson. | Hamster Cheek Pouch |
| | | **CNS** | |
| SF 295 | Glioblastoma | Obtained from Dr. Rosenblum. | Nude Athymic Mice |
| | | **MISCELLANEOUS** | |
| CWR-22 | Prostate, adenocarcinoma | Received from Dr. T. Pretlow, *in vivo* cultivation requires testosterone supplementation. Not an *in vitro* cell line. | Nude Athymic Mice |
| DAU | Burkitt's lymphoma | Received from Dr. T. Griffin, adapted to *in vivo* transplantation by Dr. A.E. Bogden. | Nude Athymic Mice |
| MRI-H-254 | Stomach, adenocarcinoma | No historical information available. | Nude Athymic Mice |
| MRI-H-1579 | Prostate Adenocarcinoma | Primary explant established *in vivo* by Dr. A.E. Bogden. Received from Dr. Blute, St. Vincent Hospital, Worcester, MA. | Nude Athymic Mice |

**Note:** ***Human In Vitro Established Cell Lines are on pages 59-62.***

# HAMSTER TUMORS

| SPECIES: HAMSTER | | | | |
|---|---|---|---|---|
| **Tumor Designation** | **Histologic Type** | **Form** | **Strain of Origin/ Transplant** | **Comments** |
| Fibrosarcoma | Fibrosarcoma | Ascites | Not Specified | |
| H-12 | Mesothelioma | Solid | Golden Syrian | |
| H-75 | Mesothelioma | Solid | Golden Syrian | |
| Islet Cell | Pancreatic Adeno-carcinoma | Solid | Golden Syrian | |
| Lymphosarcoma | Lymphosarcoma | Ascites | Not Specified | |
| Melanoma | Melanotic Melanoma | Solid | Not Specified | |
| Pan #1 (Fortner) | Pancreatic Duct Adeno-carcinoma | Solid | Not Specified | |
| SB #1 (Fortner) | Small Bowel Adeno-carcinoma | Solid | Not Specified | |
| TG1-4 | Mesothelioma | Solid | Golden Syrian | |
| TS1-4 | Epidermoid Carcinoma | Solid | Golden Syrian | |
| 10-24 | Mesothelioma | Ascites | Golden Syrian | |
| 1382J | Liver Carcinoma | Solid | Golden Syrian | |
| 2309V | Pancreatic Islet β Cell Adenocarcinoma | Solid | Golden Syrian | |
| 4671 | Pancreatic Duct Adeno-carcinoma | Solid | Golden Syrian | Line B is insulin-secreting |
| 6973P | Leiomyosarcoma | Solid | Golden Syrian | |
| 8721R | Renal Carcinoma | Solid | Golden Syrian | |
| 8746Q | Uterine Adeno-carcinoma | Solid | Golden Syrian | |
| 9242ø | Parotid Acinar Cell Adenocarcinoma | Solid | Golden Syrian | |
| 10838 | Seminoma | Solid | Golden Syrian | |
| 11348P | Pulmonary Squamous Cell Carcinoma | Solid | Golden Syrian | |
| 11963V | Leiomyosarcoma | Solid | Golden Syrian | |
| 22047 | Adenocarcinoma | Solid | Golden Syrian | |

# GUINEA PIG TUMORS

| SPECIES: GUINEA PIG | | | | |
|---|---|---|---|---|
| **Tumor Designation** | **Histologic Type** | **Form** | **Strain of Origin/ Transplant** | **Comments** |
| Line 1 | Hepatocarcinoma | Ascites | Strain 2 | |
| Guinea Pig GP Line 10 | Hepatocarcinoma | Ascites | Strain 2 | |
| Nitrosamine-induced Hepatoma | Hepatoma | Solid | Strain 39 | |

# MOUSE TUMORS

Two addenda have been inserted in this section to facilitate identification or selection of mouse tumors by histologic type (Addendum A) and by mouse strain (Addendum B). A third (Addendum C) is a list of other models together with treatment information.

# *ADDENDUM A: MOUSE TUMORS LISTED BY HISTOLOGICAL TYPE*

Adrenal

AT
$LAF_1$

Anaplastic Carcinoma

dbrB (Jax)

Bladder

FCB

Colon

CA07/A
CA36/Ara C
CA51
Colon 26
Colon 38

Ependymoblastoma

Ependymoblastoma

Fibrosarcoma

36257 TTT
Hepatoma 129

38290 TTT
Hepatoma 134

46362 TTT

46363 TTT

FB SAR
SaD2 (Jax)

Glioma

Glioma 26
Glioma 261

Hemangioendothelioma

36230 TLT
42021 TCT
42022 TST
42052 TST
42076 TST
44316 TST
44347 TST

Hepatoma

BW 7756 (JAX)
H6 (JAX)
Hepatoma 129
Hepatoma 134

Inguinal

Krebs Ascites
Krebs-2

Leukemias

C58/J Spont.
C1498
Egg/Mouse
E Male/Gross
Gross
L1210
L4946
L5178Y
P288
P388
P815
P1534
P1798
RBL-n

Lung

ASB XIV
C4461
CAD2
LC-12
Lewis
M4898
Madison
Nettesheim

Lymphoma & Lymphosarcoma

BL12
EL-4 Male
L18464
LSTRA
Mecca
NK
6C3HED

Mammary

Adenocarcinoma 755
BW10232 (Jax)
CE1460 MACA
CH
C3HBA (Jax)
DBA/2 Spont. M114
Ehrlich Ascites
EMT-6
Gross
H2712 (Jax)
Klein
MA13C
MA16C
MC-n
MCS-1
MXT
Spont. DBA/2

ADDENDUM A (continued)

Melanoma

B16
Harding-Passey
HP (Jax)

Nervous System

C1300 (Jax)
Ependymoblastoma
Glioma

Osteogenic

HE10734

Pancreas

PAN 02
PAN 03

Pituitary

A+T
BW8685 (Jax)
BW8883 (Jax)
T+T #15
T+T #97

Plasmacytoma

ADJ-PC-n
LPC-1
MOPC-n
MPC-n
RPC-n
YPC-1
70429

Reticulum Cell

Friend Virus Leukemia
M5076
SJL/JW
91632

Sarcoma

Lewis
MA387
METH-A
MS-2
S37
S180
Sa-1 (Jax)

Squamous

LC-12

Testicular

M5480

Teratosarcoma

LS402AX

Thymus

Reif-Allen

*Refer to Inventory for details. A designation followed by n, *e.g.*, MOPC-n, indicates that there is a series of tumors with this main designation, *e.g.*, MOPC-4, MOPC-17, MOPC-21, etc.

ADDENDUM B (continued)

## *ADDENDUM B: MOUSE TUMORS LISTED BY HOST STRAIN**

INBRED HOSTS

A/HE

C4461
Hauschka Ascites
Klein (TA3)

A/J

C1300 (Jax)
H6 (Jax)
Sa 1 (Jax)

AKR

L4946
MA387
Mecca
Reif-Allen

BALB/c
ADJ-PC-nASB XIV
CA07/A
CA36/Ara C
CA51
Colon 26
EMT-6
Harding-Passey
HP (Jax)
LC-12
LPC-1
LSTRA
M4898
Madison Lung
MC-n
METH-A
Moloney Sarcoma
MOPC-n
MPC-n
MS-1
MS-2
P1798

BALB/c (cont'd)

RPC-n
S37
YPC-1
1247
36257 TTT
38290 TTT
42021 TCT
44316 TST
44316 LTST
44347 TST
46363 TTT

CE

CE1460 MACA

C3H

C3HBA (Jax)
FB SAR
Gross Leukemia
H2712 (Jax)
HE 10734
Hepatoma 129
Hepatoma 134
J30237
Krebs Ascites Carcinoma
Krebs 2 Carcinoma
MA13C
MA16C
Mecca
X5563
6C3HED
70429

C57BL/6

Adenocarcinoma 755
B16
BL12
BW 10232 (Jax)

C57BL/6 (cont'd)

C1498
Colon 38
EL-4
E Male Gross
Ependymoblastoma
FCB
Glioma 26
Glioma 261
L18464
Lewis Lung
Lewis Sarcoma T241
LS402AX
M5076
M5480
NK-Lymphoma
PAN 02
PAN 03
RBL-n
T+T #15
42022 TST
42052 TST
42076 TST
46362 TTT
91632

C57L/J

BW7756 (Jax)
BW8883 (Jax)

C57BR/cdj

BW8685 (Jax)

C58

C58J/Spont.

ADDENDUM B (continued)

DBA/1

CaD1 (Jax)
dbrB (Jax)
S37 (Jax)
S91 (Jax)
T1703 (Jax)

DBA/2

CAD2
DBA/2 Spont. M114
Egg/Mouse Leukemia
Friend Virus Leukemia

DBA/2 (cont'd)

Gross Mammary Adeno-carcinoma
L1210
L5178Y
Nettesheim
P288
P329
P388
P815
P1534
S180
SaD2 (Jax)

DBA/2 (cont'd)

Spont. DBA/2 Mammary
T1699 (Jax)

SJL/J

SJL/JW

129

LS402AX

## $F_1$ HYBRIDS AND NON-INBRED HOSTS

Swiss

Ehrlich Ascites
S180

$BDF_1$

MXT

Various resistant lines of L1210 and P388

$LAF_1$

AT
$LAF_1$
MST
T+T #97

$CAF_1$

Lymphoma-2

$CDF_1$

R-n
RC-2
102A

Various resistant lines of L1210 and P388

*Refer to Inventory for details. A designation followed by n, *e.g.*, MOPC-n, indicates that there is a series of tumors with this main designation, *e.g.*, MOPC-4, MOPC-17, MOPC-21, etc.

# *ADDENDUM C: OTHER MURINE MODELS*

## L1210 Lymphoid Leukemia

L1210/TSC (NSC-729)
L1210/MTX (NSC-740)
L1210/6MP (NSC-755)
L1210/L-PAM (NSC-8806)
L1210/NSC-19622
L1210/5FU (NSC-19893)
L1210/CTX (NSC-26271)
L1210/DF8 (NSC-29630)
L1210/HU (NSC-32065)
L1210/MeGAG (NSC-32946)
L1210/NSC-38280
L1210/DTIC (45388)
L1210/TIC (NSC-60339)
L1210/Ara-C (NSC-63878)
L1210/cis-DDP (NSC-119875)
L1210/Anhydro Ara C (NSC-145668)
L1210/Ftorafur (NSC-148958)
L1210/L-Alanosine (NSC-153353)
Note: Reo3$^+$
L1210/BCNU (NSC-409962)
L1210/C95 (NSC-740, 755, 26271)
L1210/FR3 DCM/R 100a
L1210/FR8/DCM
L1210/RT8 (Folate Reductase)
L1210/M-773
L1210/6MP/6TG

*Treatment information, where available, is given in the following pages. When resistant lines are shipped, treatment information, if any, is included.

## P388 Lymphocytic Leukemia

P388/MTX (NSC-740)
P388/Actinomycin D (NSC-3053)
P388/DON (NSC-7365)
P388/L-PAM (NSC-8806)
P388/5-FU (NSC-19893)
P388/CPA (NSC-26271)
P388/Ara C (NSC-63878)
P388/Daunomycin (NSC-82151)
P388/5-Azacytidine (NSC-102816)
P388/DDP (NSC-119875) Note: Reo3$^+$
P388/ADR (NSC-123127)
P388/L-Alanosine (NSC-153353)
P388/Acivicin (NSC-163501)
P388/Amsacrine (NSC-249992)
P388/Anthracenedione (NSC-287513)
Note: Reo3$^+$
P388/Mitoxantrone (NSC-299195 + 301739)
P388/Ara-A + 2'dcF (NSC-404241 + 218321)
P388/BCNU (NSC-409962)

## Other Resistant Leukemias

L5178/L-Ase R
P288/MTX (NSC-740)
P815/VLB (NSC-49842)

## Lewis Lung Carcinoma Only in Mice

LLC-Luc-GFP (LL-LUC-POL2)

DRUG-RESISTANT MURINE LEUKEMIAS - TREATMENT INFORMATION FOR LINES RECEIVED
FROM SOUTHERN RESEARCH INSTITUTE AND ARTHUR D. LITTLE, INC.

| Tumor Line | Host of Origin, Resistant Line | Passage Inoculum (i.p.) | Treatment Used with Serial Passage | | | | Optimal Treatment to Check Degree of Resistance | | | |
|---|---|---|---|---|---|---|---|---|---|---|
| | | | NSC # | mg/Kg | Route | Schedule | NSC # | mg/Kg | Route | Schedule |
| L1210/TSC (NSC-729) | DBA/2 or CDF1 | $10^5$ | 729 | 5.0 | i.p. | Days 1-6 | 729 | 6.0 | i.p. | Q3Hx8 Days 1,5,9 |
| L1210/6-MP (NSC-755) | DBA/2 | $10^6$ | NONE | | | | 755 | 50.0 | i.p. | QD1-9 days |
| L1210/L-PAM (NSC-8806) | BDF1 | $10^6$ | 8806 | 7.5 | i.p. | Day 2 only | 8806 | 15.0 | i.p. | Day 1 only |
| L1210/CPA (NSC-26271) | DBA/2 | $10^5$ | NONE | | | | 26271 | 265.0 | i.p. | Day 1 only |
| L1210/HU (NSC-32065) | DBA/2 or CDF1 | $10^5$ | 32065 | 130.0 | i.p. | Days 1-6 | 32065 | 60.0 | i.p. | Q3Hx8 Days 1,5,9 |
| L1210/ARA-C (NSC-63878) | DBA/2 or hybrid | $10^5$ | NONE | | | | 135962 | 125.0 | i.p. | Day 1 only |
| L1210/DDP (NSC-119875) | DBA/2 or CDF1 | $10^6$ | 119875 | 5.0 | i.p. | Day 4 only | 119875 | 8.0 | i.p. | Day 1 only |
| L1210/BCNU (NSC-409962) | BDF1 | $10^5$ | NONE | | | | 409962 | 30.0 | i.p. | Day 1 only |
| P388/MTX (NSC-740) | DBA/2 or CDF1 | $10^7$ | 740 | 0.75 | s.c. | Days 1-6 | 740 | 2.0 | i.p. | QD1-9 days |
| P388/ACT-D (NSC-3053) | DBA/2 or CDF1 | $10^7$ | 3053 | 0.2 | i.p. | Day 4 only | 3053 | 0.5 | i.p. | Day 1 only |
| P388/L-PAM (NSC-8806) | BDF1 | $10^6$ | 8806 | 7.5 | i.p. | Day 2 only | 8806 | 15.0 | i.p. | Day 1 only |
| P388/5-FU (NSC-19893) | BDF1 | $10^7$ | 19893 | 20.0 | s.c. | Days 1-6 | 19893 | 25.0 | i.p. | QD1-9 days |
| P388/CPA (NSC-26271) | BDF1 | $10^7$ | 26271 | 100.0 | s.c. | Day 4 only | 26271 | 265.0 | i.p. | Day 1 only |
| P388/VCR | BDF1 | $10^7$ | NONE | | | | 67574 | 1.5 | i.p. | Days 1,5,9 |

| Tumor Line | Host of Origin, Resistant Line | Passage Inoculum (i.p.) | Treatment Used with Serial Passage | | | | Optimal Treatment to Check Degree of Resistance | | | |
|---|---|---|---|---|---|---|---|---|---|---|
| | | | NSC # | mg/Kg | Route | Schedule | NSC # | mg/Kg | Route | Schedule |
| (NSC-67574) | | | | | | | | | | |
| P388/AZACYT (NSC-102816) | DBA/2 or CDF1 | $10^7$ | 102816 | 40.0 | i.p. | Day 4 only | 102816 | 3.5 | i.p. | QD1-9 days |
| P388/DDP (NSC-119875) | DBA/2 or CDF1 | $10^7$ | 119875 | 4.5 | i.p. | Days 1&5 | 119875 | 5.3 | i.p. | Days 1,5,9 |
| P388/ADR (NSC-123127) | BDF1 | $10^7$ | 123127 | 6.0 | i.p. | Day 2 only | 123127 | 12.5 | i.p. | Day 1 only |
| P388/ARA-A + 2'dcF (NSC-404241 + NSC-218321) | BDF1 | $10^6$ | 404241 + 218321 | 125.0 .02 | i.p. | Days 2-4 | 404241 + 218321 | 60.0 0.05 | i.p. | Q3Hx8, Days 1,5,9 |
| | | | | | | | | OR | | |
| | | | | | | | 404241 + 218321 | 150.0 0.25 | i.p. | QD1-9 days |
| P388/BCNU (NSC-409962) | CDF1 | $10^7$ | 409962 | 25.0 | i.p. | Day 2 only | 409962 | 30.0 | i.p. | Day 1 only |

[+]NSC-218321 thirty minutes before NSC-404241 each time.

| SPECIES: MOUSE | | | | |
|---|---|---|---|---|
| **Tumor Designation** | **Histologic Type** | **Form** | **Strain of Origin/ Transplant** | **Comments** |
| Adenocarcinoma 755 (CA755, Bagg-Jackson, Adeno-carcinoma) | Mammary Adenocarcinoma | Solid, Ascites or Brei | C57BL | |
| ADJ-PC-6 | Plasmacytoma | Solid or Ascites | BALB/c | |
| ASB XIV | Pulmonary Squamous Cell Carcinoma | Solid | BALB/c | MVM |
| AT (Clone $Y_1$) | Adrenal | Solid | $LAF_1$ | |
| AtT/20 | Anterior Pituitary | Solid | $LAF_1$ | |
| B16 | Melanoma | Solid | C57BL/6 | Also see Jax tumors |
| BL12 Sensitive | Lymphosarcoma | Solid | C57BL/Ka | |
| BL12/Hc Ra | Lymphosarcoma | Solid | C57BL/Ka | Resistant to cortisone |
| BW7756 | Hepatoma | Solid | C57L/J | See Jax tumors |
| BW8685 | Pituitary | Solid | C57BR/Cdj | See Jax tumors |
| BW8883 | Pituitary | Solid | C57L/J | See Jax tumors |
| BW10232 | Mammary Adenocarcinoma | Solid | C57BL/6J | See Jax tumors |
| C3HBA | Mammary Adenocarcinoma | Solid | C3H/An | See Jax tumors |
| C58/J Spontaneous | Leukemia | Spleen Brei | C58 | |
| C95 Spleen/R | | | | See L1210/C95 |
| C1498 | Myelogenous Leukemia | Solid | C57BL/6 | |
| C4461 | Lung Adenocarcinoma | Solid | A/He | |
| CA07/A | Colon Adenocarcinoma | Solid | BALB/c | |
| CA36/Ara C (NSC-63878) | Colon Adenocarcinoma | Solid | BALB/c | |
| CA51 | Colon Adenocarcinoma | Solid | BALB/c | |
| CaD1 | Mammary Adenocarcinoma | Solid | DBA/1J | See Jax tumors |
| CaD2 | Mammary Adenocarcinoma | Solid | DBA/2 | |

| SPECIES: MOUSE | | | | |
|---|---|---|---|---|
| **Tumor Designation** | **Histologic Type** | **Form** | **Strain of Origin/ Transplant** | **Comments** |
| CCO/1923 | Hemangiosarcoma | Solid | B6C3F1 | |
| CE1460 MACA | Mammary Adenocarcinoma | Solid | CE | |
| CH | Mammary Adenocarcinoma | Solid | Nude C3H | |
| Cloudman Melanoma | Melanoma | Solid | DBA/1J | See Jax S91 |
| Colon 26 | Carcinoma | Solid | BALB/c | |
| Colon 38 | Carcinoma | Solid | C57BL/6 | |
| Crocker Sarcoma | | | | See S180 |
| $D_1T_{10}$ | | | | See LSTRA/DTIC |
| DBA/2 Spontaneous Tumor M114 | Mammary Adenocarcinoma | Solid | DBA/2 | |
| dbrB | Anaplastic Carcinoma | Solid | DBA/1J | See Jax tumors |
| Egg/Mouse Leukemia | Lymphocytic Leukemia | Ascites | DBA/2, Truslow Egg | |
| Ehrlich Ascites | Mammary Adenocarcinoma | Solid or Ascites | Various | Several lines |
| Ehrlich Ascites/6-TG (NSC-752) | Mammary Adenocarcinoma | Ascites | Swiss | Resistant to 6-Thioguanine |
| Ehrlich Ascites, Tetraploid | Mammary Adenocarcinoma | Ascites | Swiss | |
| EL-4 Male | Lymphoma | Solid, Spleen C57BL/6 Fragments & Ascites | | |
| E Male Gross | Leukemia | Spleen Homogenate | C57BL/6 | |
| EMT-6 | Mammary Adenocarcinoma | Solid | BALB/c | |
| Ependymoblastoma (Zimmerman) | Ependymoblastoma | Solid | C57BL/6 | |
| FB SAR (A) | Fibrosarcoma | Solid | C3H | |
| FB SAR (B) | Fibrosarcoma | Solid | C3H | |
| FCB (C) | Transitional Cell Carcinoma of Bladder | Solid | C57BL/6 | |
| | | | | |

| SPECIES: MOUSE | | | | |
|---|---|---|---|---|
| **Tumor Designation** | **Histologic Type** | **Form** | **Strain of Origin/ Transplant** | **Comments** |
| Friend Virus Leukemia (FV01) | Reticulum Cell Sarcoma | Solid or Spleen Homogenate | DBA/2 | |
| Furth Tumor | | | | See Carcinoma 1025 |
| Glioma 26 | Glioma | Solid | C57BL/6 | |
| Glioma 261 | Glioma | Solid | C57BL/6 | |
| Gross Leukemia | Leukemia | Solid | C3H | |
| Gross Mammary Adenocarcinoma | Mammary Adenocarcinoma | Solid | DBA/2 | |
| Hageman Mastocytoma | | | | See P815 |
| Harding-Passey | Melanoma | Solid | BALB/c | Melanotic and amelanotic types |
| Hauschka Ascites Tumor | Unknown | Ascites | A/He | |
| HE10734 | Osteogenic Sarcoma | Solid | C3H | |
| HE10734/FR | Osteogenic Sarcoma | Solid | C3H | |
| Hepatoma 129 (HE129) | Hepatoma | Solid | C3H or Hybrid | |
| Hepatoma 134 (HE134, Shear Hepatoma 134) | Hepatoma | Ascites | C3H | |
| HP | Amelanotic Melanoma | Solid | BALB/cJ | See Jax tumors |
| H6 | Hepatoma | Solid | A/J | See Jax tumors |
| H2712 | Mammary Adenocarcinoma | Solid | C3H/HeJ | See Jax tumors |
| J-30237 | Unknown | Ascites | C3H | |
| Klein Tumor (TA3) | Mammary Adenocarcinoma | Ascites | A/He or $CAF_1$ | Several lines |
| Krebs Ascites Carcinoma | Carcinoma of Inguinal Region | Ascites | C3H or CDBA | |
| Krebs 2 Carcinoma | Carcinoma of Inguinal Region | Ascites | C3H | |
| L1210 | Lymphoid Leukemia | Ascites or Spleen Homog-enate | DBA/2 or CDBA | |
| L1210/TSC (NSC-729) | Lymphoid Leukemia | Ascites | DBA/2 | Resistant to Thio-semicarbazone |

| SPECIES: MOUSE | | | | |
|---|---|---|---|---|
| **Tumor Designation** | **Histologic Type** | **Form** | **Strain of Origin/ Transplant** | **Comments** |
| L1210/MTX (NSC-740) | Lymphoid Leukemia | Ascites or Spleen Homog-enate | DBA/2 or CDBA | Treated |
| L1210/6MP (NSC-755) | Lymphoid Leukemia | Ascites | DBA or Hybrid | Several lines |
| L1210/L-PAM (NSC-8806) | Lymphoid Leukemia | Ascites | DBA/2 or $BDF_1$ | Treated |
| L1210/NSC-19622 | Lymphoid Leukemia | Ascites | DBA/2 | |
| L1210/5FU (NSC-19893) | Lymphoid Leukemia | Ascites | $BDF_1$ | |
| L1210/CTX (NSC-26271) | Lymphoid Leukemia | Ascites or Spleen Homog-enate | DBA/2 or CDBA | |
| L1210/DF8 (NSC-29630) | Lymphoid Leukemia | Ascites or Spleen Homog-enate | DBA/2 or CDBA | Resistant to dichloro-methotrexate |
| L1210/HU (NSC-32065) | Lymphoid Leukemia | Ascites | DBA/2 | |
| L1210/MeGAG (NSC-32946) | Lymphoid Leukemia | Ascites or Spleen Homog-enate | $CDF_1$ | |
| L1210/NSC-38280 | Lymphoid Leukemia | Ascites | $CDF_1$ | |
| L1210/DTIC (NSC-45388) | Lymphoid Leukemia | Ascites or Solid | DBA/2 or CDBA | Untreated and treated lines |
| L1210/TIC (NSC-60339) | Lymphoid Leukemia | Spleen Homogenate | CDBA | Untreated and treated lines |
| L1210/Ara-C (NSC-63878) | Lymphoid Leukemia | Ascites or Spleen Homogenate | DBA/2 or Hybrid | Untreated and treated lines |
| L1210/cis-DDP (NSC-119875) | Lymphoid Leukemia | Ascites | DBA/2 | Treated |
| L1210/Anhydro-Ara C (NSC-145668) | Lymphoid Leukemia | Ascites | DBA/2 or Hybrid | Untreated and treated lines |
| L1210/Ftorafur (NSC-148958) | Lymphoid Leukemia | Ascites | DBA/2 or $BDF_1$ | Untreated and treated lines |
| L1210/BCNU (NSC-409962) | Lymphoid Leukemia | Ascites | DBA/2 or Hybrid | Untreated and treated lines |
| L1210/C95 | Lymphoid Leukemia | Ascites or Spleen Brei | CDBA | CTX, MTX, MP resistant |

| SPECIES: MOUSE | | | | |
|---|---|---|---|---|
| **Tumor Designation** | **Histologic Type** | **Form** | **Strain of Origin/ Transplant** | **Comments** |
| L1210 FR3 DCM/R 100a | Lymphoid Leukemia | Spleen Brei | CDBA | |
| L1210/FR8/DCM | Lymphoid Leukemia | Spleen Brei | $CDF_1$ | |
| L1210/FR8 (Folate Reductase) | Lymphoid Leukemia | Spleen Brei | $CDF_1$ | |
| L1210/M-773 | Lymphoid Leukemia | Ascites | DBA/2 | Treated |
| L1210/6MP/6TG | Lymphoid Leukemia | Ascites | $CDF_1$ | Untreated and treated lines |
| L1210 variants | | | | See $PR_1C_1T5$/ NSC-45388, $PR_1SE_1T5$ and $PR_1SE_1T5$/ NSC-45388 |
| L4946 | Lymphocytic Leukemia | Solid | AKR | |
| L5178Y | Leukemia | Ascites | DBA/2 or Hybrid | Several lines |
| L18464 | Lymphoma | Solid | C57BL/6 | |
| $LAF_1$ | Adrenal Cortical Adenocarcinoma | Solid | $LAF_1$/J | |
| LC-12 | Pulmonary Squamous Cell Carcinoma | Solid | BALB/c | |
| LLC-Luc-GFP (LL-LUC-POL2) | Lung Squamous Cell Carcinoma | Solid | C57BL/6 | Grown in mice only, never in tissue culture |
| Lewis Lung | Carcinoma | Solid | C57BL | |
| Lewis Lung/PALA (NSC-224131) | Carcinoma | Solid | C57BL/6 | |
| Lewis Sarcoma T241 | Pleiomorphic Cell Sarcoma | Solid | C57BL | |
| LPC-1 | Plasmacytoma | Solid or Ascites | BALB/c | |
| LS402AX | Teratosarcoma | Solid | C57BL/6 and 129 | |
| LSTRA | Lymphosarcoma | Ascites | BALB/c | Several lines |
| LSTRA/DTIC (NSC-45388) | Lymphosarcoma | Ascites | BALB/c | Untreated and treated lines |
| M4898 | Lung Adenocarcinoma | Solid | BALB/c | |
| M5076 | Reticulum Cell Sarcoma | Solid or Ascites | C57BL/6 | |
| | | | | |

| SPECIES: MOUSE | | | | |
|---|---|---|---|---|
| **Tumor Designation** | **Histologic Type** | **Form** | **Strain of Origin/ Transplant** | **Comments** |
| M5076/L-PAM (NSC-8806) | Reticulum Cell Sarcoma | Solid | C57BL/6 | Treated |
| M5076/HMM (NSC-13875) | Reticulum Cell Sarcoma | Solid | C57BL/6 | Treated |
| M5076/cis-DDP (NSC-119875) | Reticulum Cell Sarcoma | Solid | C57BL/6 | Treated |
| M5480 | Testicular Carcinoma (Seminoma) | Solid | C57BL/6 | |
| MA13C | Mammary Adenocarcinoma | Solid | C3H | |
| MA16C | Mammary Adenocarcinoma | Solid | C3H | |
| MA387 | Fusiform Cell Sarcoma | Solid | AKR | |
| Madison Lung (TA109) | Carcinoma | Solid | BALB/c | |
| MC-5 | Mammary Adenocarcinoma | Spleen | BALB/c | |
| MC-6 Female | Mammary Adenocarcinoma | Ascites | BALB/c | |
| MC-11 | Mammary Adenocarcinoma | Spleen Homogenate | BALB/c | |
| MCS-1 | Mammary Adenocarcinoma | Solid or Spleen Homogenate | BALB/c | |
| Mecca (ME61, MLS) | Lymphosarcoma | Solid or Ascites | C3H or AKR | |
| METH-A | Sarcoma | Ascites | BALB/c | |
| MLS | | | | See Mecca |
| Moloney Sarcoma (SV-122-TR4) | Sarcoma | Solid | BALB/c | |
| MOPC-4 | Plasmacytoma | Solid or Ascites | BALB/c | |
| MOPC-17 | Plasmacytoma | Solid | BALB/c | |
| MOPC-21 | Plasmacytoma | Solid or Ascites | BALB/c | |
| MOPC-28 | Plasmacytoma | Solid | BALB/c | |
| MOPC-30 | Plasmacytoma | Solid | BALB/c | |
| MOPC-31 | Plasmacytoma | Solid | BALB/c | |
| MOPC-41 | Plasmacytoma | Solid | BALB/c | |

| SPECIES: MOUSE | | | | |
|---|---|---|---|---|
| **Tumor Designation** | **Histologic Type** | **Form** | **Strain of Origin/ Transplant** | **Comments** |
| MOPC-46 | Plasmacytoma | Solid | BALB/c | |
| MOPC-47 | Plasmacytoma | Solid | BALB/c | |
| MOPC-48 | Plasmacytoma | Solid | BALB/c | |
| MOPC-49 | Plasmacytoma | Solid | BALB/c | |
| MOPC-51 | Plasmacytoma | Solid | BALB/c | |
| MOPC-61 | Plasmacytoma | Solid | BALB/c | |
| MOPC-63 | Plasmacytoma | Solid | BALB/c | |
| MOPC-67 | Plasmacytoma | Solid | BALB/c | |
| MOPC-69 | Plasmacytoma | Solid | BALB/c | |
| MOPC-70 | Plasmacytoma | Solid | BALB/c | |
| MOPC-78 | Plasmacytoma | Solid | BALB/c | |
| MOPC-88 | Plasmacytoma | Solid | BALB/c | |
| MOPC-91 | Plasmacytoma | Spleen Homogenate | BALB/c | |
| MOPC-96 | Plasmacytoma | Solid | BALB/c | |
| MOPC-99 | Plasmacytoma | Solid | BALB/c | |
| MOPC-104 | Plasmacytoma | Solid | BALB/c | |
| MOPC-112 | Plasmacytoma | Solid | BALB/c | |
| MOPC-113 | Plasmacytoma | Solid | BALB/c | |
| MOPC-114 | Plasmacytoma | Solid | BALB/c | |
| MOPC-116 | Plasmacytoma | Solid | BALB/c | |
| MOPC-118 | Plasmacytoma | Solid | BALB/c | |
| MOPC-121 | Plasmacytoma | Solid | BALB/c | |
| MOPC-123 | Plasmacytoma | Solid | BALB/c | |
| MOPC-129 | Plasmacytoma | Solid | BALB/c | |
| MOPC-132 | Plasmacytoma | Solid | BALB/c | |
| MOPC-140 | Plasmacytoma | Solid | BALB/c | |
| MOPC-141 | Plasmacytoma | Solid | BALB/c | |
| | | | | |

| SPECIES: MOUSE | | | | |
|---|---|---|---|---|
| **Tumor Designation** | **Histologic Type** | **Form** | **Strain of Origin/ Transplant** | **Comments** |
| MOPC-157 | Plasmacytoma | Solid | BALB/c | |
| MOPC-172 | Plasmacytoma | Solid | BALB/c | |
| MOPC-173 | Plasmacytoma | Solid | BALB/c | |
| MOPC-209 | Plasmacytoma | Solid | BALB/c | |
| MPC-1 | Plasmacytoma | Solid or Ascites | BALB/c | |
| MPC-2 | Plasmacytoma | Solid or Ascites | BALB/c | |
| MPC-15 | Plasmacytoma | Solid | BALB/c | |
| MPC-25 | Plasmacytoma | Solid | BALB/c | |
| MPC-26 | Plasmacytoma | Solid | BALB/c | |
| MPC-31 | Plasmacytoma | Solid | BALB/c | |
| MPC-36 | Plasmacytoma | Solid | BALB/c | |
| MPC-37 | Plasmacytoma | Solid | BALB/c | |
| MPC-40 | Plasmacytoma | Solid | BALB/c | |
| MPC-42 | Plasmacytoma | Solid | BALB/c | |
| MPC-44 | Plasmacytoma | Solid or Ascites | BALB/c | |
| MPC-48 | Plasmacytoma | Solid | BALB/c | |
| MPC-49 | Plasmacytoma | Solid | BALB/c | |
| MPC-59 | Plasmacytoma | Solid | BALB/c | |
| MPC-60 | Plasmacytoma | Solid | BALB/c | |
| MPC-63 | Plasmacytoma | Solid | BALB/c | |
| MPC-64 | Plasmacytoma | Solid | BALB/c | |
| MPC-67 | Plasmacytoma | Solid | BALB/c | |
| MPC-73 | Plasmacytoma | Solid | BALB/c | |
| MPC-H | Plasmacytoma | Solid | BALB/c | |
| MST | Mast Cell | Solid | $LAF_1$ | |
| MS-2 | Sarcoma | Solid | BALB/c | |
| MXT | Mammary Ductal Papillary Carcinoma | Solid | $BDF_1$ | Estrogen Responsive |
| NK-Lymphoma | Lymphoma | Ascites | C57BL/6 | |

| SPECIES: MOUSE | | | | |
|---|---|---|---|---|
| **Tumor Designation** | **Histologic Type** | **Form** | **Strain of Origin/ Transplant** | **Comments** |
| Nettesheim Lung (LePage Clone KLN205) | Squamous Cell Lung Carcinoma | Solid | DBA/2 or Hybrid | |
| P288 | Lymphocytic Leukemia | Solid or Ascites | DBA/2 or CDBA | |
| P288/MTX (NSC-740) | Lymphocytic Leukemia | Ascites | DBA/2 or $BDF_1$ | |
| P388 | Lymphocytic Leukemia | Ascites | DBA/2 or CDBA | |
| P388/MTX (NSC-740) | Lymphocytic Leukemia | Ascites | DBA/2 | Treated |
| P388/ActinomycinD (NSC-3053) | Lymphocytic Leukemia | Ascites | DBA/2 | |
| P388/DON (NSC-7365) | Lymphocytic Leukemia | Ascites | DBA/2 | Treated |
| P388/L-PAM (NSC-8806) | Lymphocytic Leukemia | Ascites | $BDF_1$ | Treated |
| P388/5FU (NSC-19893) | Lymphocytic Leukemia | Ascites | $BDF_1$ | Treated |
| P388/CPA (NSC-26271) | Lymphocytic Leukemia | Ascites | $BDF_1$ | Treated |
| P388/Ara-C (NSC-63878) | Lymphocytic Leukemia | Ascites | $BDF_1$ | |
| P388/VCR (NSC-67574) | Lymphocytic Leukemia | Ascites | $BDF_1$ | Untreated and treated lines |
| P388/Daunomycin (NSC-82151) | Lymphocytic Leukemia | Ascites | DBA/2 | |
| P388/5-Azacytidine (NSC-102816) | Lymphocytic Leukemia | Ascites | DBA/2 | Treated |
| P388/ADR (NSC-123127) | Lymphocytic Leukemia | Ascites | $BDF_1$ | Treated |
| P388/L-Alanosine (NSC-153353) | Lymphocytic Leukemia | Ascites | DBA/2 | |
| P388/Acivicin (NSC-163501) | Lymphocytic Leukemia | Ascites | DBA/2 | |
| P388/Amsacrine (NSC-249992) | Lymphocytic Leukemia | Ascites | DBA/2 | |
| P388/Mitoxantrone (NSC-301739) | Lymphocytic Leukemia | Ascites | DBA/2 | |
| P388/Ara-A + 2'dcF (NSC-404241 + NSC-218321) | Lymphocytic Leukemia | Ascites | $BDF_1$ | Treated |

| SPECIES: MOUSE | | | | |
|---|---|---|---|---|
| **Tumor Designation** | **Histologic Type** | **Form** | **Strain of Origin/ Transplant** | **Comments** |
| P388/BCNU (NSC-409962) | Lymphocytic Leukemia | Ascites | $BDF_1$ | Treated |
| P815 (Hageman Mastocytoma) | Mast Cell Leukemia | Ascites | DBA/2 or Hybrid | |
| P815/VLB (NSC-49842) | Mast Cell Leukemia | Ascites | DBA/2 or $BDF_1$ | |
| P1534 | Lymphocytic Leukemia | Spleen Homogenate or Ascites | DBA/2 | Several lines |
| P1798 | Lymphosarcoma | Solid or Ascites | BALB/c | |
| P1798/CR-JS | Lymphoma | Solid | BALB/c | Glucocorticoid resistant, treated |
| P1798/CS-JS | Lymphoma | Solid | BALB/c | Glucocorticoid sensitive |
| PAN 02 | Pancreas | Solid | C57BL/6 | |
| PAN 03 | Pancreas | Solid | C57BL/6 | |
| $PR_1C_1T5$/NSC-45388 | Lymphoid Leukemia | Ascites | $CDF_1$ | L1210 variant; treated |
| $PR_1SE_1T5$ | Lymphoid Leukemia | Ascites | $CDF_1$ | L1210 variant |
| $PR_1SE_1T5$/NSC-45388 | Lymphoid Leukemia | Ascites Homo-genate | $CDF_1$ | L1210 variant; treated |
| R-26 | Unknown | Ascites | $CDF_1$ | |
| R-46 | Unknown | Ascites | $CDF_1$ | |
| R-53 | Unknown | Ascites | $CDF_1$ | |
| R-74 | Unknown | Ascites | $CDF_1$ | |
| RBL-5 (Rauscher Virus Induced Transplantable Tumor-5) | Leukemia | Ascites | C57BL/6 | |
| RC-2 | Renal Adenocarcinoma | Ascites | $CDF_1$ | |
| Reif-Allen Tumor | Thymoma | Ascites | AKR | |
| RPC-5 | Plasmacytoma | Solid | BALB/c | |
| RPC-9 | Plasmacytoma | Solid or Ascites | BALB/c | |
| RPC-20 | Plasmacytoma | Solid or Ascites | BALB/c | |
| S37 | Pleomorphic Cell Sarcoma | Ascites | BALB/c, Nonspecific | See Jax tumors |

| SPECIES: MOUSE | | | | |
|---|---|---|---|---|
| **Tumor Designation** | **Histologic Type** | **Form** | **Strain of Origin/ Transplant** | **Comments** |
| S180 (Crocker, S III) | Pleomorphic Cell Sarcoma | Solid or Ascites | BALB/c, Swiss or hybrids | See Jax tumors |
| Sa 1 | Spindle Cell Sarcoma | Solid | A/J | See Jax tumors |
| Sa D2 | Fibrosarcoma | Solid | DBA/2J | See Jax tumors |
| Shear Hepatoma 134 | | | | See Hepatoma 134 |
| SJL/JW | Reticulum Cell Sarcoma | Spleen Homogenate | SJL/JW | |
| Spontaneous Adrenal | Adrenal | Solid | CE/J | |
| Spontaneous DBA/2 Mammary | Mammary Adenocarcinoma | Solid | DBA/2 | |
| Spontaneous Mammary | Mammary Adenocarcinoma | Solid | DBA/2 | |
| SV-122-TR4 | | | | See Moloney Sarcoma |
| TA3 | | | | See Klein tumor |
| TA109 | | | | See Madison Lung |
| T1699 | Mammary Adenocarcinoma | Solid | DBA/2J | See Jax tumors |
| T1703 | Mammary Adenocarcinoma | Solid | DBA/1J | See Jax tumors |
| X5563 | Unknown | Solid | C3H/He | |
| YPC-1 | Plasmacytoma | Ascites | BALB/c | |
| Zimmerman Ependymoblastoma | | | | See Ependymoblastoma |
| 6C3HED (Gardner) | Lymphosarcoma | Ascites | C3H | Several lines. See Jax tumors |
| 6C3HED/AR Res. | Lymphosarcoma | Spleen Homogenate | C3H | |
| 102A | Unknown | Ascites | $CDF_1$ | |
| 1247 | Mammary Adenocarcinoma | Solid | BALB/c | |
| 36230 TLT | Hemangio-endothelioma | Solid | C57BL/6J | |
| 36257 TTT | Fibrosarcoma | Solid | BALB/cAnN | |
| | | | | |

| SPECIES: MOUSE | | | | |
|---|---|---|---|---|
| **Tumor Designation** | **Histologic Type** | **Form** | **Strain of Origin/ Transplant** | **Comments** |
| 38290 TTT | Fibrosarcoma | Solid | BALB/cAnN | |
| 42021 TCT | Hemangio-endothelioma | Solid | BALB/cAnN | |
| 42022 TST | Hemangio-endothelioma | Solid | BALB/cAnN | |
| 42052 TST | Hemangio-endothelioma | Solid | C57BL/6J | |
| 42076 TST | Hemangio-endothelioma | Solid | C57BL/6J | |
| 44316 TST | Hemangio-endothelioma | Solid | BALB/cAnN | |
| 44316 LTST | Hemangio-endothelioma | Solid | BALB/cAnN | |
| 44347 TST | Hemangio-endothelioma | Solid | BALB/cAnN | |
| 46362 TTT | Fibrosarcoma | Solid | C57BL/6J | |
| 46363 TTT | Fibrosarcoma | Solid | BALB/cAnN | |
| 70429 | Plasmacytoma | Ascites | C3H | |
| 70429/Azaserine (NSC-3425) | Plasmacytoma | Ascites | C3HF/LW | |
| 91632 | Reticulum Cell Sarcoma | Solid | C57BL/Kaplan | |

# MOUSE TUMORS FROM THE JACKSON LABORATORY

**MOUSE TUMORS FROM THE JACKSON LABORATORY IN CRYOPRESERVATION IN THE DCTD TUMOR REPOSITORY**

| Tumor Designation | Tumor Type | Host Strain | Transplantation Frequency (days) | Host Survival (days) | Lag Time* (days) | Strain of Origin | Sex of Origin | MRI Bank # | MAP Test + |
|---|---|---|---|---|---|---|---|---|---|
| dbrB | Anaplastic Carcinoma | DBA/1J | 7 | 7-9 | 5-7 | DBA | | J-730 | LDH+ |
| SaD2 | Fibrosarcoma | DBA/2J | 10 | 19-12 | 8-15 | DBA/2J | | J-765 | LDH+ |
| H6 | Hepatoma | A/J | 10-14 | 14-44 | 7-9 | A/J | | J-750 | LDH+ |
| BW7756 | Hepatoma | C57L/J | 14 | 13-31 | 12-16 | C57L/J | | J-748 | LDH+ |
| 6C3HED (GL-1) | Lymphosarcoma | C3H/HeJ | 7 | 7-9 | 16 | C3h | -- | J-744 | LDH+ |
| BW10232 | Mammary Adenocarcinoma | C57BL/6J | 10 | 23-30 | 11 | C57BL/6J | | J-762 | LDH+ |
| C3HBA | Mammary Adenocarcinoma | C3H/HeJ | 10 | 39-77 | 11-16 | C3H/An | | H-758 | LDH+ |
| H2712 | Mammary Adenocarcinoma | C3H/HeJ | 7 | 14-27 | 11-19 | C3H/HeHu | | J-731 | LDH+ |
| CaD1 | Mammary Adenocarcinoma | DBA/1J | 10 | 25-43 | 9-17 | DBA/1J | | J-742 | LDH+ |
| T1703 | Mammary Adenocarcinoma | DBA/1J | 10 | 47-74 | 9-12 | DBA/1Hu | | J-737 | LDH+ |
| T1699 | Mammary Adenocarcinoma | DBA/2J | 10 | 19-39 | 8-10 | DBA/2J | | J-736 | LDH+ |
| HP | Melanoma (amelanotic) | BALB/cJ | 14 | 28-77 | 19-23 | nonlabred | -- | J-746 | LDH+ |
| B16 | Melanoma (amelanotic) | C57BL/6J | 10 | 24-44 | 15-21 | C57BL/6J | -- | J-753 | LDH+ |
| S91 | Melanoma (melanotic) | DBA/1J | 17-21 | 49-98 | 16-18 | DBA (Snell) | -- | J-749 | LDH+ |
| C1498 | Myeloid Leukemia | C57BL/6J | 7 | 9-16 | 7-9 | C57BL | -- | J-738 | LDH+ |
| BW8883 | Pituitary | C57L/J | 60 | 182-273 | 65-73 | C57L/J | -- | J-756 | LDH+ |

| Tumor Designation | Tumor Type | Host Strain | Transplantation Frequency (days) | Host Survival (days) | Lag Time* (days) | Strain of Origin | Sex of Origin | MRI Bank # | MAP Test [+] |
|---|---|---|---|---|---|---|---|---|---|
| BW8685 | Pituitary | C57BR/dcJ | 90-120 | 210-238 | 395 | C57BR/cdj | | J-794 | LDH+ |
| S180 | Pleomorphic Sarcoma | BALB/cJ | 10 | 21-31 | 9-11 | “white” mouse | | J-757 | LDH+ |
| S37 | Pleomorphic Sarcoma | DBA/1J | 7 | 21-28 | 6-13 | “stock” mouse | | J-759 | LDH+ |
| C1300 | Round cell (Neuroblastoma?) | A/J | 10 | 19-32 | 14-21 | A albino | -- | J-734 | LDH+ MHV+ |
| Sal | Spindle-cell Sarcoma | A/J | 7 | 9-15 | 7-9 | A albino | -- | J-733 | LDH+ |

*Length of lag phase before measurable tumor growth (5 mm average diameter) is evident in the first passage post thaw.
[+]MAP Test - Murine Antigen Profile for 12 common viruses: PVH, Rco 3, Sendal, GDVII, K, Polyoma, MVH, MAB, MHV, LCM, Ectromelia, LDH. Only positive results are listed.

Adapted from Jax Notes, No. 424, December 1975.

# RABBIT TUMORS

| SPECIES: RABBIT | | | | |
|---|---|---|---|---|
| **Tumor Designation** | **Histologic Type** | **Form** | **Strain of Origin/ Transplant** | **Comments** |
| Brown-Pearce | Carcinoma (Epithelioma) | Solid | New Zealand White or Dutch | |
| VX-2 (V2) | Skin Carcinoma | Solid | New Zealand White or Dutch | |

# RAT TUMORS

| SPECIES: RAT | | | | |
|---|---|---|---|---|
| **Tumor Designation** | **Histologic Type** | **Form** | **Strain of Origin/ Transplant** | **Comments** |
| AA Ascites | Spontaneous Ascites | Ascites | Wistar | |
| ATC 64 | Thyroid Carcinoma | Solid | Fischer 344 | |
| 9A Ascites | Spontaneous Ascites | Ascites | Inbred PA | |
| A546 (DMBZ Attenuated) | Unknown | Solid | Fischer 344 | |
| A920 (Tetramin Attenuated Resistant) | Unknown | Solid | Fischer 344 | |
| A1011 | Unknown | Solid | Fischer 344 | |
| A1131-AR | Unknown | Solid | Fischer 344 | |
| A1138-AL | Unknown | Solid | Fischer 344 | |
| A1140-CL-10 | Unknown | Solid | Fischer 344 | |
| BT/M520 | Fibrosarcoma | Solid | Marshall 520 | |
| Carcinosarcoma | Carcinosarcoma (skin) (18-day embryos) | Solid | Sprague-Dawley Derived Holtzman | |
| CA 20948 | Pancreatic Acinar Carcinoma | Solid | Wistar | |
| CCO 1865 | Mesothelioma | Solid | Fischer 344 | |
| CSE | Fibrosarcoma | Solid | Fischer 344 | H-1 |
| DD81-23 | T cell lymphoma | Mince | Fischer 344 | |
| DL Wells-17 | T cell lymphoma | Mince | Fischer 344 | |
| DMBA | Mammary Adenocarcinoma | Solid | Fischer 344 | Several lines |
| DSL62-38 | Pancreatic Tumor | Solid | Wistar Lewis | |
| Dunning Leukemia | Atypical Monocytic Leukemia | Solid or Ascites | Fischer 344 | H-1<br>H-1, KRV |
| Dunning Leukemia/ NSC-755 (6-MP) | Atypical Monocytic Leukemia | Solid | Fischer 344 | |
| Dunning Leukemia/ NSC-3088 (chlor-ambucil) | Atypical Monocytic Leukemia | Solid | Fischer 344 | |
| Dunning Leukemia/ NSC-10107 (nitromin) | Atypical Monocytic Leukemia | Solid | Fischer 344 | |
| Dunning Leukemia/ NSC-13875 (HMM) | Atypical Monocytic Leukemia | Solid | Fischer 344 | |

| SPECIES: RAT | | | | |
|---|---|---|---|---|
| **Tumor Designation** | **Histologic Type** | **Form** | **Strain of Origin/ Transplant** | **Comments** |
| Dunning Leukemia/ NSC-17261 (benzo-quinone) | Atypical Monocytic Leukemia | Solid | Fischer 344 | |
| Dunning Leukemia/ NSC-23892 (dimethyl-benzimidazole) | Atypical Monocytic Leukemia | Solid | Fischer 344 | |
| Dunning Leukemia/ NSC-26980 (mitomycin C) | Atypical Monocytic Leukemia | Solid | Fischer 344 | |
| Dunning Leukemia/ NSC-29422 (thio-guanosine) | Atypical Monocytic Leukemia | Solid | Fischer 344 | |
| Dunning Leukemia/ NSC-45059 (o-acetyl-tetramin) | Atypical Monocytic Leukemia | Solid | Fischer 344 | |
| Dunning Leukemia/ NSC-51845 (cyclo-hexylamine) | Atypical Monocytic Leukemia | Solid | Fischer 344 | |
| Flexner-Jobling | Seminal Vesicle Adenocarcinoma | Solid | Nonspecific, Sprague-Dawley or Wistar | |
| Fran Tumor | Ovarian Carcinoma | Ascites | Sprague-Dawley | |
| GBT/W | Glial Tumor | Solid | Wistar | |
| HB Lynch-Fibroma 522 | Fibroma | Solid | Fischer 344 | |
| Hepatoma NK | | | | See Novikoff Hepatoma |
| HMC | Histiocytoma | Solid | Fischer 344 | |
| H-372 | Leydig | Solid | Fischer 344 | |
| H-540 | Leydig | Solid | Fischer 344 | |
| Iglesias | Ovarian Carcinoma | Solid | ACI | |
| IRS 9802 | Spindle Cell Sarcoma | Solid | Fischer 344 | |
| LC-18 | Hepatoma | Solid | Fischer 344 | |
| Lymphoma 8 | Lymphoma | Solid, Spleen Homogenate, Whole Blood | Lewis | |
| L.T.W. (Furth) | Leydig | Solid | Wistar | |
| MAMF2-TC | Fibrosarcoma | Solid | Fischer 344 | |
| MET 149-2 | Adenocarcinoma | Solid | Fischer 344 | |

| SPECIES: RAT | | | | |
|---|---|---|---|---|
| **Tumor Designation** | **Histologic Type** | **Form** | **Strain of Origin/ Transplant** | **Comments** |
| Moore Sarcoma #1 | Sarcoma | Solid | Wistar | |
| Morris Hepatomas | Hepatoma | Solid | Buffalo | |
| 16 Morris | Hepatoma | Solid | Buffalo | |
| 20 Morris | Hepatoma | Solid | Buffalo | |
| 44 Morris | Hepatoma | Solid | Buffalo | |
| 3924A | Hepatoma | Solid | ACI | |
| 5123 Morris | Hepatoma | Solid | Buffalo | |
| 7777 Morris | Hepatoma | Solid | Buffalo | |
| 7800 Morris | Hepatoma | Solid | Buffalo | |
| 8999 Morris | Hepatoma | Solid | Buffalo | |
| 9618A Morris | Hepatoma | Solid | Buffalo | |
| MNU-Buffalo | Mammary Carcinoma | Solid | Buffalo | Several lines |
| MtT | Anterior Pituitary | Solid | Fischer, Wistar | Several lines |
| MT/W9 | Mammary Adenocarcinoma | Solid | Wistar | Several lines |
| MT/W449 | Mammary Adenocarcinoma | Solid | Wistar | Several lines |
| Murphy-Sturm Lymphosarcoma (MSL) | Lymphosarcoma | Solid | CRL, Wistar, Fischer 344, Sprague-Dawley | |
| NBW-37 | T cell lymphoma | Mince | Fischer 344 | |
| Novikoff Hepatoma (Hepatoma NK) | Hepatoma | Solid or Ascites | Random bred albino | (Sprague-Dawley weanlings) |
| NS104 | Rhabdomyosarcoma | Solid | Fischer 344 | |
| OR-16-3 | Thymus Tumor | Solid | Fischer 344 | |
| R35 | Mammary Adenocarcinoma | Solid | Holtzman | |
| R3259 | Giant Cell Sarcoma | Solid | Fischer 344 | |
| Rice D6 | Leydig | Solid | Fischer 344 | |
| Rice 500 | Leydig | Solid | Fischer 344 | |
| Riejoel | Thyroid Adenocarcinoma | Solid | Fischer 344 | |
| | | | | |

| SPECIES: RAT | | | | |
|---|---|---|---|---|
| **Tumor Designation** | **Histologic Type** | **Form** | **Strain of Origin/ Transplant** | **Comments** |
| RNC 259 | Pheochromocytoma | Solid | NEDH | |
| RNC 288 | Insylinoma | Solid | NEDH | |
| RNK-16 | LGL Leukemia | Solid or Spleen Homogenate | Fischer 344 | |
| R3149 | Leukemia | Solid | Fischer 344 | |
| R3230AC | Mammary Adenocarcinoma | Solid | Fischer 344 | |
| R3327 | Prostate | Solid | Copenhagen 2331 | |
| R3327 (Pap) | Prostate | Solid | Copenhagen 2331 | |
| 3M2N | Mammary Squamous Cell Carcinoma | Solid | Fischer 344 | |
| Shay Leukemia | Myelogenous Leukemia | Solid | Sprague-Dawley | |
| SMT-2A | Mammary Carcinoma | Solid | Fischer 344 | |
| Swarm | Chondrosarcoma | Solid | Sprague-Dawley | |
| TR.CLXXXVIII | Melanoma | Solid | ACI | |
| TR.DCCXLIII | Pituitary | Solid | ACI | |
| Walker 256 | Carcinosarcoma | Solid or Ascites | Sprague-Dawley | |
| Yoshida Hepatoma | Hepatoma | Ascites | Sprague-Dawley | |
| Yoshida Sarcoma | Sarcoma | Solid or Ascites | Holtzman, S-D | |
| 68-2 | Alveolar/Bronchiolar Carcinoma | Solid | Fischer 344 | |
| 13762 | Mammary | Solid or Ascites | Fischer 344 | Several lines |
| 13762-FS | Fibrosarcoma | Solid | Fischer 344 | |
| 23 Methapyrilene | Hepatocellular Carcinoma | Solid | Fischer 344 | |
| 29 Methapyrilene | Hepatocellular Carcinoma | Solid | Fischer 344 | |
| 33 Methapyrilene | Hepatocellular Carcinoma | Solid | Fischer 344 | |
| 2982 | Olfactory Neuroblastoma | Solid | Fischer 344 | |
| 11095 | Prostate | Solid | Fischer 344 | |

# RAT TUMORS FROM DR. ROBERT NOBLE (ENDOCRINE-RESPONSIVE)

SPECIES: RAT (tumors received from Dr. Robert Noble)

Information concerning the endocrinologic characteristics of the various tumor systems indicated in the "comments" is that supplied by Dr. Noble.

| **SPECIES: RAT** (tumors received from Dr. Robert Noble) | | | | |
|---|---|---|---|---|
| **Tumor Designation** | **Histologic Type** | **Form** | **Strain of Origin/ Transplant** | **Comments** |
| 1 Cvx-34A(1) | Cervical Carcinoma | Solid | NB | |
| 1 Cvx-44Z | Cervical Carcinoma | Solid | NB | Estrogen dependent |
| 1 Lym-206 | Lymphoma | Solid | NB | Hormone stimulated |
| 1 Lym-209(A) | Lymphoma | Solid | NB | Hormone stimulated, VLB sensitive |
| 1 Lym-214 | Lymphosarcoma | Solid | NB | Hormone stimulated |
| 1 Og-3 | Osteogenic Sarcoma | Solid | NB | |
| 1 Pan-14Ax(1) | Adenocarcinoma | Solid | NB | Hormone stimulated |
| 1 Sal-23(2) | Salivary Gland, Secretory Tumor | Solid | NB | Estrogen dependent |
| 1 Tes-13E | Leydig Cell Carcinoma | Solid | NB | Estrogen dependent |
| 1 TES-15E | Leydig Cell Carcinoma | Solid | NB | Estrogen dependent |
| Kid-27B | Kidney Adenocarcinoma | Solid | NB | |
| 2 Lym-11(a) | Metaplastic, Adeno-carcinoma Fibroblast Overgrowth | Solid | NB | Estrogen dependent |
| 2-Pan-6A | Pituitary Adenoma | Solid | NB | Hormone stimulated |
| 2-Pr-7A | Prostatic Adenocarcinoma | Solid | NB | Estrogen dependent |
| 2 Pr-9F | Prostatic Adenocarcinoma | Solid | NB | Estrogen dependent |
| 2 Pr-12 | Prostatic Adenocarcinoma | Solid | NB | Estrogen dependent |
| 2 Pr-112Bx(1) | Prostatic Carcinoma, Scirrhous | Solid | NB | |
| 2 Pr-114B | Prostatic Adenocarcinoma | Solid | NB | Estrogen dependent |
| 2 Pr-121D(1) | Prostatic Carcinoma, Secretory | Solid | NB | Androgen dependent |
| 2 Pr-121D(1)/R | Prostatic Carcinoma | Solid | NB | Resistant to testosterone |
| 2 Sk-103 | Melanoma | Solid | NB | |
| 2 Ut-10(5) | Fibroma | Solid | NB | Estrogen dependent |
| 3 Kid-13 | Kidney Adenocarcinoma | Solid | NB | |
| 3 Lym-19 | Lymphosarcoma | Solid | NB | |
| 4 Pan-6 | Adenocarcinoma | Solid | NB | |

| **SPECIES: RAT** (tumors received from Dr. Robert Noble) | | | | |
|---|---|---|---|---|
| **Tumor Designation** | **Histologic Type** | **Form** | **Strain of Origin/ Transplant** | **Comments** |
| 4 Sk-3A(3)Z | Squamous Cell Carcinoma | Solid | NB | |
| 4 Ut | Hemangiosarcoma | Solid | NB | |
| 4 Ut-6(2) | Fibrosarcoma | Solid | NB | Estrogen dependent or hormone stimulated |
| 5 Pan-7 | Undifferentiated Pancreatic Carcinoma | Solid | NB | |
| 5 Sal | Undifferentiated Carcinoma | Solid | NB | |
| 5 Sk-3 | Melanoma | Solid | NB | |
| 5 Ut-2 | Uterine Adenocarcinoma | Solid | NB | Estrogen dependent (?) |
| 6 Pan-4 | Undifferentiated Pancreatic Carcinoma | Solid | NB | |
| 7 Ut-13 | Endometrial Adeno-carcinoma | Solid | NB | |
| 8 Lym-9(1) | Lymphosarcoma | Solid | NB | VLB resistant |
| 8 Lym-108(1) | Lymphatic Leukemia | Solid | NB | VLB resistant |
| 9 Lym-23 | Lymphosarcoma | Solid | NB | |
| 10 Lym-4 | Negative Spleen | Solid | NB | |
| 11 Lym-9 | Lymphosarcoma | Solid | NB | |
| 13 Pr-5 | Prostatic Carcinoma, Undifferentiated | Solid | NB | Estrogen pellet implant required |
| 14 Lym-5 | Lymphosarcoma stimulated | Solid | NB | Hormone |
| 14 Pr-5 | Prostatic Carcinoma | Solid | NB | |
| 15 Pr-2 | Prostatic Adenocarcinoma | Solid | NB | |
| 16 Pr-3 | Prostatic Adenocarcinoma | Solid | NB | |
| 17 Lym-4 | Lymphosarcoma implant required | Solid | NB | Estrogen pellet |
| 17 Lym-5 | Leukemia | Solid | NB | |
| 18 Lym-6 | Lymphosarcoma | Solid | NB | Estrogen dependent |
| 19 Lym-3 | Lymphosarcoma | Solid | NB | Estrogen dependent |
| 19 Pr-19 | Prostatic Fibroadenoma | Solid | NB | |
| 20 Pr-1 | Prostatic Fibroadenoma | Solid | NB | |
| 20 Lym-3 | Lymphosarcoma | Solid | NB | Estrogen pellet implant required |

| **SPECIES: RAT** (tumors received from Dr. Robert Noble) | | | | |
|---|---|---|---|---|
| **Tumor Designation** | **Histologic Type** | **Form** | **Strain of Origin/ Transplant** | **Comments** |
| 21 Pr-9 | Prostatic Carcinoma | Solid | NB | |
| 22 Pr-8 | Prostatic Adenocarcinoma | Solid | NB | |
| 23 Pr-7 | Prostatic Carcinoma | Solid | NB | |
| 24 Pr-2 | Prostatic Carcinoma | Solid | NB | |
| 25 Pr-3 | Prostatic Adenocarcinoma | Solid | NB | |

# *IN VITRO* ESTABLISHED CELL LINES

## *HUMAN* IN VITRO *CELL LINES*

| Designation | Tissue of Origin | Histologic Type | Growth Medium | Remarks |
|---|---|---|---|---|
| ACHN | Kidney | Renal Cell Carcinoma | RPMI 1640 | From Dr. Schmid |
| ASPS-1 | Lymph Node Metastasis | Alveolar Soft Part Sarcoma | DMEM | From Dr. Shoemaker |
| A2780 | Ovary | Adenocarcinoma | RPMI 1640 | From Dr. Hamilton |
| A498 | Kidney | Renal Cell Carcinoma | RPMI 1640 | ATCC |
| A549 | Lung | Non-small Cell | RPMI 1640 | ATCC |
| A704 | Kidney | Renal Cell Carcinoma | RPMI 1640 | ATCC |
| BT-549 | Breast | Adenocarcinoma | RPMI 1640 | ATCC |
| CAKI-1 | Kidney | Renal Cell Carcinoma | RPMI 1640 | From Dr. Loveless |
| CCRF-CEM | Lymph | Leukemia | RPMI 1640 | ATCC |
| CCRF-SB | Lymph | Leukemia | RPMI 1640 | ATCC |
| CHA-59 | Bone | Osteosarcoma | RPMI 1640 | From Drs. Shoemaker and McLachlan |
| COLO 205 | Colon | Adenocarcinoma | RPMI 1640 | ATCC |
| DMS-114 | Lung | Small Cell | RPMI 1640 | From Dr. Pettengill |
| DU-145 | Prostate | Carcinoma | RPMI 1640 | ATCC |
| EKVX | Lung | Adenocarcinoma | RPMI 1640 | From Dr. Fodstad |
| HCC-2998 | Colon | Adenocarcinoma | RPMI 1640 | From Dr. Fidler |
| HCT-15 | Colon | Carcinoma | RPMI 1640 | ATCC |
| HCT-116 | Colon | Adenocarcinoma | RPMI 1640 | ATCC |
| HELA | Cervix | Carcinoma | | |
| HOP-18 | Lung | Large Cell Carcinoma | RPMI 1640 | From Drs. Liu/Casero |
| HOP-62 | Lung | Adenocarcinoma | RPMI 1640 | From Drs. Liu/Casero |
| HOP-92 | Lung | Large Cell | RPMI 1640 | From Dr. Liu |
| HL-3 | | Lymphoma | | |
| HL-4 | | Lymphoma | | |
| HL-60 | Ascites | Pro-myelocytic Leukemia | RPMI 1640 | From E. Jensen |
| H-MESO-1 | | Mesothelioma | RPMI 1640 | |
| HS 578T | Breast | Adenocarcinoma | RPMI 1640 | ATCC |
| HS 913T | Lung | Mixed Cell | RPMI 1640 | ATCC |
| HT-29 | Colon | Adenocarcinoma | RPMI 1640 | ATCC |

## *HUMAN* IN VITRO *CELL LINES*

| Designation | Tissue of Origin | Histologic Type | Growth Medium | Remarks |
|---|---|---|---|---|
| IGR-OV1 | Ovary | Adenocarcinoma | RPMI 1640 | From Dr. Benard |
| KB-ADL #12 | Oral Cavity | Epidermoid | EMEM | |
| KM-12 | Colon | Adenocarcinoma | RPMI 1640 | From Dr. Fidler |
| KM 20L2 | Colon | Adenocarcinoma | RPMI 1640 | From Dr. Fidler |
| K-562 | Lymph | Leukemia | RPMI 1640 | ATCC |
| LOVO | Colon | Adenocarcinoma | RPMI 1640 | ATCC |
| LOX IMVI | Lymph Node Metastasis | Amelanotic Melanoma | RPMI 1640 | From Dr. Fodstad |
| LXFL 529** | Lung | Large Cell Carcinoma | RPMI 1640 | From Dr. Fiebig** |
| MALME-3M | Lung Metastasis | Melanoma | RPMI 1640 | ATCC |
| MCF7 | Breast | Adenocarcinoma | RPMI 1640 | From Dr. Cowan |
| MDA-MB-231 | Breast | Adenocarcinoma | RPMI 1640 | From Dr. Moore |
| MDA-MB-435 | Melanoma | Adenocarcinoma | RPMI 1640 | From Dr. Steeg |
| MDA-MB-468 | Breast | Adenocarcinoma | | |
| MOLT-4 | Lymph | Leukemia | RPMI 1640 | ATCC |
| MX-1 | Breast | Carcinoma | RPMI 1640 | From Dr. Giovannelli |
| M14 | | Amelanotic Melanoma | RPMI 1640 | From Dr. Kern |
| M19-MEL | | Amelanotic Melanoma | RPMI 1640 | From Dr. Kern |
| NC-37 | Lymphoblast | Normal | | |
| NCI-H23 | Lung | Adenocarcinoma | RPMI 1640 | From Dr. Gazdar |
| NCI-H69 | Lung | Small Cell Carcinoma | RPMI 1640 | From Dr. Gazdar |
| NCI-H82 | Lung | Small Cell Carcinoma | RPMI 1640 | From Dr. Gazdar |
| NCI-H125 | Lung | Adenosquamous Carcinoma | RPMI 1640 | From Dr. Gazdar |
| NCI-H226 | Lung | Squamous Cell | RPMI 1640 | From Dr. Gazdar |
| NCI-H292 | Lung | Adenosquamous Carcinoma | RPMI 1640 | From Dr. Gazdar |
| NCI-H322M | Lung | Adenocarcinoma | RPMI 1640 | From Dr. Gazdar |
| NCI-H358M | Lung | Bronchioalveolar Carcinoma | RPMI 1640 | From Dr. Gazdar |
| NCI-H460 | Lung | Large Cell | RPMI 1640 | From Dr. Gazdar |
| NCI-H522 | Lung | Adenocarcinoma | RPMI 1640 | From Dr. Gazdar |
| NCI/ADR-RES | Ovary | Adenocarcinoma | RPMI 1640 | From Dr. Cowan Please see JNCI correspondence on page 64 |

## *HUMAN* IN VITRO *CELL LINES*

| Designation | Tissue of Origin | Histologic Type | Growth Medium | Remarks |
|---|---|---|---|---|
| OV | Ovary | | | |
| OVCAR-3 | Ovary | Adenocarcinoma | RPMI 1640 | From Drs. Ozols and Hamilton |
| OVCAR-4 | Ovary | Adenocarcinoma | RPMI 1640 | From Drs. Ozols and Hamilton |
| OVCAR-5 | Ovary | Adenocarcinoma | RPMI 1640 | From Drs. Ozols and Hamilton |
| OVCAR-8 | Ovary | Adenocarcinoma | RPMI 1640 | From Drs. Ozols and Hamilton |
| PC-3 | Prostate | Carcinoma | RPMI 1640 | From Dr. Kaighn |
| PC-3/M | Prostate | Carcinoma | RPMI 1640 | From Dr. Kaighn |
| RPMI-7951 | | Melanoma | RPMI 1640 | ATCC |
| RPMI-8226 | Lymph | Leukemia | RPMI 1640 | ATCC |
| RXF 393 | Kidney | Renal Cell Carcinoma | RPMI 1640 | From Dr. Fiebig |
| RXF 631 | Kidney | Renal Cell Carcinoma | RPMI 1640 | From Dr. Fiebig |
| SF-268 | CNS | Glioblastoma | RPMI 1640 | From Dr. Rosenblum |
| SF-295 | CNS | Glioblastoma | RPMI 1640 | From Dr. Rosenblum |
| SF-539 | CNS | Glioblastoma | RPMI 1640 | From Dr. Rosenblum |
| SHP-77 | Lung | Small Cell Carcinoma | RPMI 1640 | From Dr. Fisher |
| SK-OV-3 | Ovary | Adenocarcinoma | RPMI 1640 | ATCC |
| SK-MEL-2 | | Melanoma | RPMI 1640 | ATCC |
| SK-MEL-5 | | Melanoma | RPMI 1640 | ATCC |
| SK-MEL-28 | | Melanoma | RPMI 1640 | ATCC |
| SK-MES-1 | Lung | Squamous Cell Carcinoma | RPMI 1640 | ATCC |
| SN12A1 | Kidney | Renal Cell Carcinoma | RPMI 1640 | From Dr. Fidler |
| SN12C | Kidney | Renal Cell Carcinoma | RPMI 1640 | From Dr. Fidler |
| SN12K1 | Kidney | Renal Cell Carcinoma | RPMI 1640 | From Dr. Fidler |
| SN12L1 | Kidney | Renal Cell Carcinoma | RPMI 1640 | From Dr. Fidler |
| SN12S1 | Kidney | Renal Cell Carcinoma | RPMI 1640 | From Dr. Fidler |
| SNB-7 | CNS | Glioblastoma | RPMI 1640 | From Dr. Kornblith |
| SNB-19 | CNS | Glioblastoma-<br>(Same as U251) | RPMI 1640 | From Dr. Kornblith |
| SNB-75 | CNS | Glioblastoma | RPMI 1640 | From Dr. Kornblith |
| SNB-78 | CNS | Astrocytoma | RPMI 1640 | From Dr. Kornblith |

## *HUMAN* IN VITRO *CELL LINES*

| Designation | Tissue of Origin | Histologic Type | Growth Medium | Remarks |
|---|---|---|---|---|
| SR | Pleural effusion | Lymphoma | RPMI 1640 | From Dr. Urba |
| SW-620 | Colon | | RPMI 1640 | ATCC |
| T-47D | Breast | | RPMI 1640 | Not distributed to commercial firms or for commercial purposes |
| TK-10 | Kidney | Renal Cell Carcinoma | RPMI 1640 | From Dr. Clayman |
| UACC-62 | | Melanoma | RPMI 1640 | From Dr. Leibowitz |
| UACC-257 | | Melanoma | RPMI 1640 | From Dr. Leibowitz |
| UCSD 242L | | Melanoma | RPMI 1640 | From Dr. Taetle |
| UCSD 354L | | Melanoma | RPMI 1640 | From Dr. Taetle |
| UO-31 | Kidney | Renal Cell Carcinoma | RPMI 1640 | From Dr. Linehan |
| U-251 | CNS | Glioblastoma- (Same as SNB-19) | RPMI 1640 | From Dr. Bigner |
| WIDR | Colon | Adenocarcinoma | RPMI 1640 | ATCC |
| XF 498 | CNS | Glioblastoma | RPMI 1640 | From Dr. Fiebig |
| 786-0 | Kidney | Renal Cell Carcinoma | RPMI 1640 | From Dr. Williams |

## NIH LICENSED CELL LINES

| Designation | Tissue of Origin | Histologic Type | Growth Medium | Remarks |
|---|---|---|---|---|
| NCI-293TT | Human Embryonic Kidney | Kidney | DMEM 10% FBS | Schiller/Pang |
| NCI-H1299 | Human Lung NSCLC | Adenocarcinoma | RPI 1640 | Gazdar/Minna |
| NCI-H226 | Human Lung | Squamous Cell | RPMI 1640 | Gazdar |
| NCI-H23 | Human Lung | Adenocarcinoma | RPMI 1640 | Gazdar/Minna |
| NCI-H2887 | Human Lung NSCLC | Adenocarcinoma | RPMI 1640 | Gazdar/Minna |
| NCI-H3122 | Human Lung NSCLC | Adenocarcinoma | RPMI 1640 | Gazdar/Minna |
| NCI-H322M | Human Lung | Bronchi Alveolar Carcinoma | RPMI 1640 | Gazdar |
| NCI-H3255 | Human Lung NSCLC | Adenocarcinoma | ACL-4 + 10%FBS + 1% Glutamine | Gazdar/Minna |
| NCI-H460 | Human Lung | Large Cell Carcinoma | RPMI 1640 | Gazdar |
| NCI-H522 | Human Lung | Adenocarcinoma | RPMI 1640 | Gazdar |
| NCI-H838 | Human Lung NSCLC | Adenocarcinoma | RPMI 1640 | Gazdar/Minna |

| NCI ANTI-CANCER CELL LINE PANEL | | | | | | |
|---|---|---|---|---|---|---|
| **Cell Line** | **Sex** | **Age** | **Histology** | **Comment** | **Treat-ment** | **Source** |
| **COLON** | | | | | | |
| COLO 205 | M | 70 | Adenocarcinoma | Can Res 38: 1345-1355, 1978 | | |
| HCC-2998 | | | Carcinoma | | N | |
| HCT-15 | | | Adenocarcinoma | Can Res 39: 1020-1025, 1979 | | |
| HCT-116 | | | Carcinoma | Can Res 41: 1761-1756, 1981 | | |
| HT29 | F | 44 | Adenocarcinoma, GR III | Human Tumor Cells In Vitro: 115-159, 1975 | | Primary |
| KM12 | | | Adenocarcinoma | Can Res 48: 1943-1948, 1988 | N | |
| SW-620 | M | 51 | Adenocarcinoma | Can Res 36: 4562-4569, 1976 | | Metastasis |
| **CNS** | | | | | | |
| SF-268 | F | 24 | Anaplastic Astrocytoma | Acta Neuropathol 75: 92-103, 1987 | | |
| SF-295 | F | 67 | Glioblastoma-Multiforme | Acta Neuropathol 75: 92-103, 1987 | | |
| SF-539 | | | | J Neuropathol Exp Neurol 40: 201- 229, 1981 | | |
| SNB-19 | M | 47 | Glioblastoma (same as U251) | Cancer 47: 255, 1981 | N | |
| SNB-75 | F | | Astrocytoma | | N | |
| U251 | M | 75 | Glioblastoma (same as SNB-19) | J Neuropathol Exp Neurol 40: 410-427, 1981 | | |
| **LEUKEMIA** | | | | | | |
| CCRF-CEM | M | 4 | Acute Lymphoblastic Leukemia | Can Res 18: 522-529, 1965 | | |
| HL-60(TB) | F | 36 | Promyelocytic Leukemia | Nature 270: 347-349, 1977 | | PBL |
| K-562 | F | 53 | Chronic Myelogenous Leukemia | Blood 45: 321-334, 1975 | | Pleural Effusion |
| MOLT-4 | M | 19 | Acute Lymphoblastic Leukemia | JNCI 49: 891-895, 1972 | | PB |
| RPMI-8226 | M | 61 | Myeloma | Proc Soc Exp Biol Med 125: 1246-1250, 1967 | | PB |
| SR | M | 11 | Large Cell, Immunoblastic | | Y | |
| **LUNG** | | | | | | |
| A549/ATCC | M | 58 | Adenocarcinoma | JNCI 51: 1417-1423, 1973 | | Primary |
| EKVX | M | | Adenocarcinoma | | | |
| HOP-62 | F | 60 | Adenocarcinoma | | N | |
| HOP-92 | M | 62 | Large Cell, Undifferentiated | | N | |
| NCI-H23 | | | Adenocarcinoma | Can Res 45: 2913-2923, 1985 | N | |
| NCI-H226 | | | Squamous | Can Res 45: 2913-2923, 1985 | | |

| NCI ANTI-CANCER CELL LINE PANEL | | | | | | |
|---|---|---|---|---|---|---|
| **Cell Line** | **Sex** | **Age** | **Histology** | **Comment** | **Treat-ment** | **Source** |
| **LUNG (continued)** | | | | | | |
| NCI-H322M | | | Small Cell Bronchioalveolar Carcinoma | | N | |
| NCI-H460 | M | | Large Cell Carcinoma | Science 246: 491-494, 1989 | N | Pleural Effusion |
| NCI-H522 | | | Adenocarcinoma | Can Res 45: 2913-2923, 1985 | | |
| **MAMMARY** | | | | | | |
| MCF7 | F | 69 | Adenocarcinoma | JNCI 51: 1409-1417, 1973 | Y | |
| HS 578T | F | 74 | Carcinosarcoma | JNCI 58: 1795-1806, 1977 | | Primary |
| MDA-MB-231 | F | 51 | Adenocarcinoma | JNCI 53: 661-674, 1974 | Y | |
| MDA-MB-468 | F | 51 | Adenocarcinoma | Cancer Res 40: 3118-3129, 1980 | | |
| BT-549 | F | 72 | Papillary Infiltrating Ductal Carcinoma | No Publication | | Metastasis |
| T-47D***** | F | 54 | Infiltrating Ductal Carcinoma | Eur J Cancer 15: 659-670, 1979 | | *not for com-mercial use |
| **MELANOMA** | | | | | | |
| LOX IMVI | | | Malignant Amelanotic Melanoma | Int J Cancer 41: 442-449, 1988 | | |
| M14 | | | | | | |
| MALME-3M | M | 43 | Malignant Melanoma | Human Tumor Cells In Vitro, 115-159, 1975 | | Metastasis |
| MDA-MB-435 | F | 31 | Adenocarcinoma | Can Res 40: 3118-3129, 1980 | N | |
| SK-MEL-2 | M | 60 | Malignant Melanoma | Human Tumor Cells In Vitro, 115-159, 1975 | | Metastasis |
| SK-MEL-5 | | | Malignant Melanoma | PNAS 73: 3278-3282, 1976 | | Metastasis |
| SK-MEL-28 | | | Malignant Melanoma | PNAS 73: 3278-3282, 1976 | | |
| UACC-62 | | | | | | |
| UACC-257 | | | | | | |
| **OVARIAN** | | | | | | |
| IGR-OV1 | F | 47 | Cystoadenocarcinoma | Can Res 45: 4970-4979, 1985 | N | |
| NCI/ADR-RES | F | | Adenocarcinoma | JNCI 90(11): 6/3/1998 | | |
| OVCAR-3 | F | 60 | Adenocarcinoma | Can Res 43: 5379-5389, 1983 | Y | Ascites |
| OVCAR-4 | F | 42 | Adenocarcinoma | Sem Oncol 11: 285-298, 1984 | Y | |
| OVCAR-5 | F | 67 | Adenocarcinoma | Sem Oncol 11: 285-298, 1984 | N | |
| OVCAR-8 | F | 64 | Adenocarcinoma | Sem Oncol 11: 285-298, 1984 | Y | |
| SK-OV-3 | F | 64 | Adenocarcinoma | Human Tumor Cells In Vitro, pp 115-159, | Y | Ascites |

| NCI ANTI-CANCER CELL LINE PANEL | | | | | | |
|---|---|---|---|---|---|---|
| **Cell Line** | **Sex** | **Age** | **Histology** | **Comment** | **Treat-ment** | **Source** |
| | | | | 1975 | | |
| **PROSTATE** | | | | | | |
| DU-145 | M | 69 | Carcinoma | Int J Cancer 21: 274-281, 1978 | Y | |
| PC-3 | M | 62 | Adenocarcinoma | Invest Urol 17: 16-23, 1979 | Y | Metastasis |
| **RENAL** | | | | | | |
| 786-O | M | 58 | Adenocarcinoma | In Vitro 12: 623-627, 1976 | N | |
| A498 | F | 52 | Adenocarcinoma | JNCI 51: 1417-1423, 1973 | | |
| ACHN | M | 22 | Renal Cell Carcinoma | Can Res 42: 4948-4953, 1982 | Y | Pleural Effusion |
| CAKI-1 | M | 49 | Clear Cell Carcinoma | Human Tumor Cells In Vitro, pp. 115-159, 1975 | Y | Metastasis |
| RXF 393 | M | 54 | Poorly Differentiated Hypernephroma | Contrib Oncol 42, 1992 | N | |
| SN12C | M | 43 | Carcinoma | Can Res 46: 4109-4115, 1986 | | |
| TK-10 | M | 43 | Spindle Cell Carcinoma | Can Res 47: 3856-3862, 1987 | N | |
| UO-31 | | | Carcinoma | | N | |

# CORRESPONDENCE

## Cell Line Designation Change: Multidrug-Resistant Cell Line in the NCI Anticancer Screen

Since 1990, the Developmental Therapeutics Program (DTP) of the National Cancer Institute (NCI) has screened over 60,000 compounds and a larger number of natural product extracts for their capacity to inhibit the growth of 60 different human tumor cell lines *(1)*. These cell lines have been maintained in cryopreservation and in culture, and they have been subjected to strict quality controls, including adventitious agent testing, human isoenzyme analysis, karyology, morphological and immunocytochemical characterization *(2)*, and DNA fingerprinting. One of these cell lines, previously designated as MCF-7/ADR-RES, has been included in the *in vitro* cell line screening panel because of its stable multidrug-resistant (MDR) phenotype *(3)* characterized by high levels of MDR-1 and P-glycoprotein expression *(4,5)*. Recently, we submitted cell lines from the screening panel for DNA fingerprinting analysis by three different laboratories. Included in the tested cell lines were MCF-7 and MCF-7/ADR-RES. Utilizing restriction fragment length polymorphism (RFLP) testing, CellMark Diagnostics (Germantown, MD) concluded that their DNA fingerprinting data were consistent with each of the cell lines (MCF-7 and MCF-7/ADR-RES) having different donors. The other laboratories–American Type Culture Collection (Rockville, MD) used both RFLP and amplification fragment length polymorphism (AmpFLP) methods, and Children's Hospital of Michigan Cell Culture Laboratory (Detroit, MI) used the AmpFLP method–reached the same conclusions. Based on the reports from these DNA fingerprinting analyses, we have concluded that the preponderance of the information available suggests that the MCF-7/ADR-RES multidrug-resistant cell line that is included in the DTP screening program is not related to the MCF-7 cell line that is a part of the screening panel. **Thus, we have changed the nomenclature of the MCF-7/ADR-RES multidrug-resistant cell line.** The new designation of this cell line is **NCI/ADR-RES**. This nomenclature change will soon appear in all DTP databases, including the Worldwide Web. The DTP Web site address is: http://dtp.nci.nih.gov

Irrespective of its origin, this cell line has served as a valuable sentinel for compounds interacting with the multidrug-resistant mechanism *(5)*.

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**References**

(1) Monks A, Scudiero D, Skehan P, Shoemaker R, Paull K, Vistica D, et al. Feasibility of a high-flux anticancer drug screen using a diverse panel of cultured human tumor cell lines. J Natl Cancer Inst 1991; 83:757-66.
(2) Stinson SF, Alley MC, Kopp WC, Fiebig HH, Mullendore LA, Pittman AF, et al. Morphological and immunocyto-chemical characteristics of human tumor cell lines for use in a disease-orientated anticancer drug screen. Anticancer Res 1992; 12:1035-53.
(3) Wu L, Smythe AM, Stinson SF, Mullendore LA, Monks A, Scudiero DA, et al. Multidrug-resistant phenotype of disease-orientated panels of human tumor cell lines used for anticancer drug screening. Cancer Res 1992;52:3029-34.
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(5) Alvarez M, Paull K, Monks A, Hose C, Lee JS, Weinstein J, et al. Generation of a drug resistance profile by quantitation of mdr-1/P-glycoprotein in the cell lines of the National Cancer Institute Anticancer Drug Screen. J Clin Invest 1995; 95:2205-14.

## Notes

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## *NONHUMAN* IN VITRO *CELL LINES*

| Designation | Species | Histologic Type | Tissue of Origin | Growth Medium | Remarks |
|---|---|---|---|---|---|
| CHO 1C T6 | Hamster | Normal | Ovary | F12 | |
| $B16F_1$ | Mouse | Melanoma | Ear (B16) | EMEM | From Fidler |
| $B16F_{10}$ | Mouse | Melanoma | Lung met. | EMEM | From Fidler; high lung met. |
| $B16F^{Lr6}$ | Mouse | Melanoma | Lung met. | EMEM | From Fidler; low lung met. |
| B16BL-6 | Mouse | Melanoma | Bladder met. | EMEM | From Fidler; intermediate lung met. |
| BALB/c 3T3 | Mouse | Normal | Embryo | Dulbecco | |
| C3HIOT 1/2 | Mouse | | | | No info |
| Colon 26 | Mouse | Carcinoma | Colon | RPMI 1640 | |
| EL-4 | Mouse | Lymphoma | | | |
| FBL-3 | Mouse | Leukemia | | | |
| Lewis Lung | Mouse | Carcinoma | Lung | EMEM | |
| L1210/MRI | Mouse | Leukemia | Ascites | Fischer's | |
| L5178Y(R)/MRI | Mouse | Leukemia | Ascites | Fischer's | |
| MPC-11 | Mouse | Myeloma | | | |
| M5076 | Mouse | Reticulum cell sarcoma | | RPMI 1640 | |
| P3X63 | Mouse | | | | No info |
| P388 | Mouse | Leukemia | Ascites | RPMI 1640 | |
| P388/ADR | Mouse | Leukemia | Ascites | RPMI 1640 | |
| PAN 02 | Mouse | Adenocarcinoma | Pancreas | RPMI 1640 | |
| YAC | Mouse | Lymphoma | | EMEM | |
| K-1735 | Mouse | Melanoma | | EMEM | |
| UV-2237 | Mouse | Fibrosarcoma | | EMEM | |
| MADB 106 | Rat | | | | |
| MADB 200 | Rat | | | | |

# YEAST STRAINS

| YEAST STRAINS USED FOR NCI COMPOUND SCREENING | | |
|---|---|---|
| **SPY #** | **Relevant Mutation(s)** | **Complete Genotype** |
| 50636 | *rad52* | *MATα rad52ΔURA3 erg6ΔLEU2 pdr1ΔLEU2 pdr3ΔhisG::URA3::hisG ade2 ade3 leu2 trp1 ura3 cyh2* |
| 50644 | none (wild-type control) | *MATα erg6ΔLEU2 pdr1ΔLEU2 pdr3ΔhisG::URA3::hisG ade2 ade3 leu2 ura3 cyh2* |
| 50648 | *rad50* | *MATα rad50Δkan$^r$ ade2 ade3 leu2 ura3 trp1 cyh2* |
| 50649 | *mlh1 rad18* | *MATα mlh1ΔTRP1 rad18ΔLEU2 erg6ΔLEU2 pdr1ΔLEU2 pdr3ΔhisG::URA3::hisG ade2 ade3 leu2 ura3 cyh2* (*trp1*?) |
| 50650 | *mgt1* | *MAT mtg1Δkan$^r$ erg6ΔLEU2 pdr1ΔLEU2 pdr3ΔhisG::URA3::hisG ade2 ade3 leu2 trp1 ura3 cyh2* |
| 50652 | *rad50* | *MATα rad50Δkan$^r$ erg6ΔLEU2 pdr1ΔLEU2 pdr3ΔhisG::URA3::hisG ade2 ade3 leu2 ura3 cyh2* |
| 50654 | *mec2-1* | *MATα mec2-1 erg6ΔLEU2 pdr1ΔLEU2 pdr3ΔhisG::URA3::hisG ade2 ade3 leu2 ura3 cyh2* |
| 50740 | *rad14* | *MATα rad14Δkan$^r$ erg6ΔLEU2 pdr1ΔLEU2 pdr3ΔhisG::URA3::hisG ade2 ade3 leu2 ura3 cyh2* |
| 50745 | *sgs1 mgt1* | *MATα sgs1ΔLEU2 mgt1kan$^r$ erg6ΔLEU2 pdr1ΔLEU2 pdr3ΔhisG ade2 ade3 leu2 ura3 cyh2* |
| 50768 | *GPD*p-*CLN2* | *MATα URA3-GPD*p-*CLN2 erg6ΔTRP1 pdr1ΔLEU2 pdr3ΔhisG ade2 ade3 leu2 trp1 ura3 cyh2* |
| 50771 | *GPD*p-*CLN2 rad14* | *MATα URA3-GPD*p-*CLN2 rad14Δkan$^r$ erg6ΔTRP1 pdr1ΔLEU2 pdr3ΔhisG ade2 ade3 leu2 ura3 cyh2 trp1* |
| 50779 | *bub3* | *MATα bub3ΔURA3 erg6ΔTRP1 pdr1ΔLEU2 pdr3ΔhisG ade2 ade3 leu2 ura3 cyh2 trp1* |
| 50780 | none (wild-type control) | *MATα erg6ΔTRP1 pdr1ΔLEU2 pdr3ΔhisG ade2 ade3 leu2 trp1 ura3 cyh2* |
| 50834 | *mlh1* | *MATα mlh1ΔTRP1 erg6ΔTRP1 pdr1ΔLEU2 pdr3ΔhisG ade2 ade3 leu2 ura3 cyh2* |
| 50835 | *sgs1* | *MATα sgs1ΔLEU2 erg6ΔTRP1 pdr1ΔLEU2 pdr3ΔhisG ade2 ade3 leu2 ura3 cyh2* |
| 50891 | *rad18* | *MATα rad18ΔURA3 erg6ΔTRP1 pdr1ΔLEU2 pdr3ΔhisG ade2 ade3 leu2 trp1 ura3 cyh2* |

Notes:

Store at -70°C to -80°C. To establish working stock: scrape frozen culture with a wooden applicator stick and apply sample to agar-containing media (vials should remain frozen).

All strains are derived from L. Hartwell laboratory strains in the A364a genetic background.

The *erg6 pdr1 pdr3* mutations in all strains serve to make yeast more sensitive to a variety of compounds.

The allele present at the *TRP1* locus is unknown for SPY50649 (strain is phenotypically Trp$^+$ by virtue of *TRP1* at the *MLH1* locus).

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|---|---|---|---|
| A1011 | Unknown | Rat | 49 |
| A1131-AR | Unknown | Rat | 49 |
| A1138-AL | Unknown | Rat | 49 |
| A1140-CL-10 | Unknown | Rat | 49 |
| A2780 | Adenocarcinoma | Human *In Vitro* Cell Lines | 58 |
| A498 | Renal Cell Carcinoma | Human *In Vitro* Cell Lines | 58 |
| A546 (DMBZ) | | Rat | 49 |
| A549 | Nonsmall Cell | Human *In Vitro* Cell Lines | 58 |
| A704 | Renal Cell Carcinoma | Human *In Vitro* Cell Lines | 58 |
| A920 (Tetramin) | | Rat | 49 |
| AA Ascites | Spontaneous Ascites | Rat | 49 |
| ACHN | Renal Cell Carcinoma | Human *In Vitro* Cell Lines | 58 |
| Adenocarcinoma 755 (CA755, Bagg-Jackson, Adenocarcinoma) | Mammary Adenocarcinoma | Mouse | 31 |
| ADJ-PC-6 | Plasmacytoma | Mouse | 31 |
| ASB XIV | Pulmonary Squamous Cell Carcinoma | Mouse | 31 |
| ASPS-1 | Alveolar Soft Part Sarcoma | Human | 58 |
| AT | Adrenal | Mouse | 31 |
| ATC 64 | Thyroid Carcinoma | Rat | 49 |
| AtT/20 | Anterior Pituitary | Mouse | 31 |
| Attenuated Resistant | Unknown | Rat | 49 |
| B16 | Melanoma (amelanotic) | Jackson Lab Mouse | 44 |
| B16BL-6 | Melanoma | NonHuman *In Vitro* Cell Lines | 66 |
| $B16F^{1}$ | Melanoma | NonHuman *In Vitro* Cell Lines | 66 |
| $B16F^{10}$ | Melanoma | NonHuman *In Vitro* Cell Lines | 66 |
| $B16F^{Lr6}$ | Melanoma | NonHuman *In Vitro* Cell Lines | 66 |
| BALB/c 3T3 | Normal | NonHuman *In Vitro* Cell Lines | 66 |
| BL12 Sensitive | Lymphosarcoma | Mouse | 31 |
| BL12/Hc Ra | Lymphosarcoma | Mouse | 31 |
| BOW-G | Melanosarcoma | Human | 16 |
| Brown-Pearce | Carcinoma (Epithelioma) | Rabbit | 47 |
| BT/M520 | Fibrosarcoma | Rat | 49 |
| BT-549 | Adenocarcinoma | Human *In Vitro* Cell Lines | 58 |
| BW10232 | Mammary Adenocarcinoma | Mouse | 31 |
| BW10232 | Mammary Adenocarcinoma | Jackson Lab Mouse | 44 |
| BW7756 | Hepatoma | Mouse | 31 |
| BW7756 | Hepatoma | Jackson Lab Mouse | 44 |
| BW8685 | Pituitary | Mouse | 31 |
| BW8685 | Pituitary | Jackson Lab Mouse | 45 |
| BW8883 | Pituitary | Mouse | 31 |
| BW8883 | Pituitary | Jackson Lab Mouse | 44 |
| C1300 | Round Cell (Neuroblastoma?) | Jackson Lab Mouse | 45 |

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| C1498 | Myeloid Leukemia | Jackson Lab Mouse | 44 |
| C3HBA | Mammary Adenocarcinoma | Mouse | 31 |
| C3HBA | Mammary Adenocarcinoma | Jackson Lab Mouse | 44 |
| C3HIOT ½ | | NonHuman *In Vitro* Cell Lines | 66 |
| C4461 | Lung Adenocarcinoma | Mouse | 31 |
| C58/J Spontaneous | Leukemia | Mouse | 31 |
| C95 Spleen/R | | Mouse | 31 |
| CA 20948 | Pancreatic Acinar Carcinoma | Rat | 49 |
| CA07/A | Colon Adenocarcinoma | Mouse | 31 |
| CA36/Ara C (NSC-63878) | Colon Adenocarcinoma | Mouse | 31 |
| CA51 | Colon Adenocarcinoma | Mouse | 31 |
| CaD1 | Mammary Adenocarcinoma | Mouse | 31 |
| CaD1 | Mammary Adenocarcinoma | Jackson Lab Mouse | 44 |
| CaD2 | Mammary Adenocarcinoma | Mouse | 31 |
| CAKI-1 | Renal Cell Carcinoma | Human *In Vitro* Cell Lines | 58 |
| Carcinosarcoma | Carcinosarcoma (skin) (18-day embryos) | Rat | 49 |
| CCO 1865 | Mesothelioma | Rat | 49 |
| CCO/1923 | Hemangiosarcoma | Mouse | 32 |
| CCRF-CEM | Leukemia | Human *In Vitro* Cell Lines | 58 |
| CCRF-SB | Leukemia | Human *In Vitro* Cell Lines | 58 |
| CE1460 MACA | Mammary Adenocarcinoma | Mouse | 32 |
| CH | Mammary Adenocarcinoma | Mouse | 32 |
| CHA-59 | Bone, Osteosarcoma | Human | 58 |
| CHO 1C T6 | Normal | NonHuman *In Vitro* Cell Lines | 66 |
| Cloudman Melanoma | Melanoma | Mouse | 32 |
| COLO 205 | Adenocarcinoma | Human *In Vitro* Cell Lines | 58 |
| Colon 26 (C26) | Carcinoma | Mouse | 32 |
| Colon 26 | Carcinoma | NonHuman *In Vitro* Cell Lines | 66 |
| Colon 38 | Carcinoma | Mouse | 32 |
| COO-G | Mammary Carcinoma | Human | 14 |
| COS-G | Lung, Papillary Carcinoma | Human | 14 |
| Crocker Sarcoma | | Mouse | 32 |
| CSE | Fibrosarcoma | Rat | 49 |
| CWR-22 | Prostate, Adenocarcinoma | Human | 18 |
| CX-1 (HT-29) | Colon, Adenocarcinoma | Human | 15 |
| CX-2 | Colon, Carcinoma | Human | 15 |
| CX-3 | Colon, Carcinoma | Human | 15 |
| CX-5 | Colon, Adenocarcinoma | Human | 15 |
| D1T10 | | Mouse | 32 |
| DAU | Burkitt's Lymphoma | Human | 18 |
| DBA/2 Spontaneous Tumor M114 | Mammary Adenocarcinoma | Mouse | 32 |

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| dbrB | Anaplastic Carcinoma | Jackson Lab Mouse | 44 |
| DDB81-23 | T cell Lymphoma | Rat | 49 |
| DEAC-1 | Mucoepidermoid Carcinoma | Human | 18 |
| DEAC-7 | Melanoma | Human | 16 |
| DEL-G | Sarcoma | Human | 18 |
| DL Wells-17 | T cell Lymphoma | Rat | 49 |
| DMBA | Mammary Adenocarcinoma | Rat | 49 |
| DMS-114 | Small Cell | Human *In Vitro* Cell Lines | 58 |
| DSL62-38 | Pancreatic Tumor | Rat | 49 |
| DU-145 | Adenocarcinoma | Human *In Vitro* Cell Lines | 58 |
| DU4475 | Mammary Carcinoma | Human | 14 |
| Dunning Leukemia | Atypical Monocytic Leukemia | Rat | 49 |
| Dunning Leukemia/ NSC-10107 (nitromin) | Atypical Monocytic Leukemia | Rat | 49 |
| Dunning Leukemia/ NSC-13875 (HMM) | Atypical Monocytic Leukemia | Rat | 49 |
| Dunning Leukemia/ NSC-17261 (benzoquinone) | Atypical Monocytic Leukemia | Rat | 50 |
| Dunning Leukemia/ NSC-23892 (dimethylbenzimidazole) | Atypical Monocytic Leukemia | Rat | 50 |
| Dunning Leukemia/ NSC-26980 (mitomycin C) | Atypical Monocytic Leukemia | Rat | 50 |
| Dunning Leukemia/ NSC-29422 (thioguanosine) | Atypical Monocytic Leukemia | Rat | 50 |
| Dunning Leukemia/ NSC-3088 (chlorambucil) | Atypical Monocytic Leukemia | Rat | 49 |
| Dunning Leukemia/ NSC-45059 (o-acetyltetramin) | Atypical Monocytic Leukemia | Rat | 50 |
| Dunning Leukemia/ NSC-51845 (cyclohexylamine) | Atypical Monocytic Leukemia | Rat | 50 |
| Dunning Leukemia/ 755 (6-MP) | Atypical Monocytic Leukemia | Rat | 49 |
| E Male Gross | Leukemia | Mouse | 32 |
| Egg/Mouse Leukemia | Lymphocytic Leukemia | Mouse | 32 |
| Ehrlich Ascites | Mammary Adenocarcinoma | Mouse | 32 |
| Ehrlich Ascites, Tetraploid | Mammary Adenocarcinoma | Mouse | 32 |
| Ehrlich Ascites/6-TG (NSC-752) | Mammary Adenocarcinoma | Mouse | 32 |
| EKVX | Adenocarcinoma | Human *In Vitro* Cell Lines | 58 |
| EL-4 | Lymphoma | NonHuman *In Vitro* Cell Lines | 66 |
| EL-4 Male | Lymphoma | Mouse | 32 |
| ELL-G | Mammary Carcinoma | Human | 14 |
| EMT-6 | Mammary Adenocarcinoma | Mouse | 32 |
| Ependymoblastoma (Zimmerman) | Ependymoblastoma | Mouse | 32 |
| FB SAR (A) | Fibrosarcoma | Mouse | 32 |

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|---|---|---|---|
| FB SAR (B) | Fibrosarcoma | Mouse | 32 |
| FBL-3 | Leukemia | NonHuman *In Vitro* Cell Lines | 66 |
| FCB (C) | Transitional Cell Carcinoma of the Bladder | Mouse | 32 |
| Fibrosarcoma | Fibrosarcoma | Hamster | 20 |
| Flexner-Jobling | Seminal Vesicle Adenocarcinoma | Rat | 50 |
| FO #1 | Melanoblastoma | Human | 16 |
| Fran Tumor | Ovarian Carcinoma | Rat | 50 |
| Friend Virus Leukemia (FV01) | Reticulum Cell Sarcoma | Mouse | 33 |
| Furth Tumor | | Mouse | 33 |
| FV01 | | Mouse | 33 |
| Gardner Lymphosarcoma | | Mouse | |
| GBT/W | Glial Tumor | Rat | 50 |
| GL-1 | | Jackson Lab Mouse | 44 |
| Glioma 26 | Glioma | Mouse | 33 |
| Glioma 261 | Glioma | Mouse | 33 |
| GOB-G | Colon, Adenocarcinoma | Human | 15 |
| Gross Leukemia | Leukemia | Mouse | 33 |
| Gross Mammary Adenocarcinoma | Mammary Adenocarcinoma | Mouse | 33 |
| Guinea Pig Line 1 GP1 | Hepatocarcinoma | Guinea Pig | 22 |
| Guinea Pig Line 1 GP10 | Hepatocarcinoma | Guinea Pig | 22 |
| H-12 | Mesothelioma | Hamster | 20 |
| H23 (NCI-H23) | Lung, Nonsmall Cell, Adenocarcinoma | Human | 15 |
| H2712 | Mammary Adenocarcinoma | Mouse | 33 |
| H2712 | Mammary Adenocarcinoma | Jackson Lab Mouse | 44 |
| H-372 | Leydig | Rat | 50 |
| H460 (NCI-H460) | Lung, Nonsmall Cell, Epid. | Human | 15 |
| H-540 | Leydig | Rat | 50 |
| H6 | Hepatoma | Mouse | 33 |
| H6 | Hepatoma | Jackson Lab Mouse | 44 |
| H-75 | Mesothelioma | Hamster | 20 |
| Hageman | | Mouse | 33 |
| Harding-Passey | Melanoma | Mouse | 33 |
| Hauschka Ascites Tumor | Unknown | Mouse | 33 |
| HB Lynch-Fibroma 522 | Fibroma | Rat | 50 |
| HCC-2998 | Colorectal Carcinoma | Human | 15 |
| HCC-2998 | Adenocarcinoma | Human *In Vitro* Cell Lines | 58 |
| HCT-116 | Adenocarcinoma | Human *In Vitro* Cell Lines | 58 |
| HCT-15 | Colon, Carcinoma | Human | 15 |
| HCT-15 | Carcinoma | Human *In Vitro* Cell Lines | 58 |
| HE10734 | Osteogenic Sarcoma | House | 33 |
| HE10734/FR | Osteogenic Sarcoma | House | 33 |
| HELA | Carcinoma | Human *In Vitro* Cell Lines | 58 |

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| HEP-3 | Epidermoid Carcinoma | Human | 18 |
| Hepatoma 129 (HE129) | Hepatoma | Mouse | 33 |
| Hepatoma 134 (HE134, Shear Hepatoma 134) | Hepatoma | Mouse | 33 |
| Hepatoma NK | | Rat | 50 |
| HIG-G | Mammary Carcinoma | Human | 14 |
| HL-3 | Lymphoma | Human *In Vitro* Cell Lines | 58 |
| HL-4 | Lymphoma | Human *In Vitro* Cell Lines | 58 |
| HL-60 | Pro-myelocytic Leukemia | Human *In Vitro* Cell Lines | 58 |
| HMC | Histiocytoma | Rat | 50 |
| H-MESO-1 | Lung, Mesothelioma | Human | 15 |
| H-MESO-1 | Mesothelioma | Human *In Vitro* Cell Lines | 58 |
| H-MESO-1A | Lung, Mesothelioma | Human | 15 |
| HOP-18 | Large Cell Carcinoma | Human *In Vitro* Cell Lines | 58 |
| HOP-62 | Adenocarcinoma | Human *In Vitro* Cell Lines | 58 |
| HOP-92 | Large Cell | Human *In Vitro* Cell Lines | 58 |
| HP | Amelanotic Melanoma | Mouse | 33 |
| HP | Melanoma (Amelanotic) | Jackson Lab Mouse | 44 |
| HS 578T | Adenocarcioma | Human *In Vitro* Cell Lines | 58 |
| HS 913T | Mixed Cell | Human *In Vitro* Cell Lines | 58 |
| HS-1 | Sarcoma | Human | 18 |
| HT-29 | Adenocarcinoma | Human *In Vitro* Cell Lines | 58 |
| IGR-OV1 | Adenocarcinoma | Human *In Vitro* Cell Lines | 59 |
| IRS 9802 | Spindle Cell Sarcoma | Rat | 50 |
| Iglesias | Ovarian Carcinoma | Rat | 50 |
| Islet Cell | Pancreatic Adenocarcinoma | Hamster | 20 |
| J-30237 | Unknown | Mouse | 33 |
| K-1735 | Melanoma | NonHuman *In Vitro* Cell Lines | 66 |
| K-562 | Leukemia | Human *In Vitro* Cell Lines | 59 |
| KB-ADL #12 | Epidermoid | Human *In Vitro* Cell Lines | 59 |
| Kid-27B | Kidney Adenocarcinoma | Rat | 54 |
| Klein Tumor (TA3) | Mammary Adenocarcinoma | Mouse | 33 |
| KLO-G | Colon, Adenocarcinoma | Human | 15 |
| KM 20L2 (derivative of HT-29) | Colon, Adenocarcinoma | Human | 15 |
| KM 20L2 (derivative of HT-29) | Adenocarcinoma | Human *In Vitro* Cell Lines | 59 |
| KM-12 | Adenocarcinoma | Human *In Vitro* Cell Lines | 59 |
| Krebs 2 Carcinoma | Carcinoma of Inguinal Region | Mouse | 33 |
| Krebs Ascites | | Mouse | 33 |
| L.T.W. (Furth) | Leydig | Rat | 50 |
| L1210 | Lymphoid Leukemia | Mouse | 33 |
| L1210 FR3 DCM/R 100a | Lymphoid Leukemia | Mouse | 35 |
| L1210 Variants | | Mouse | 35 |

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| L1210/5FU (NSC-19893) | Lymphoid | Mouse | 34 |
| L1210/6MP (NSC-755) | Lymphoid Leukemia | Mouse | 34 |
| L1210/6MP/6TG | Lymphoid Leukemia | Mouse | 35 |
| L1210/Anhydro-Ara C (NSC-145668) | Lymphoid Leukemia | Mouse | 34 |
| L1210/Ara-C (NSC-63878) | Lymphoid Leukemia | Mouse | 34 |
| L1210/BCNU (NSC-409962) | Lymphoid Leukemia | Mouse | 34 |
| L1210/C95 | Lymphoid Leukemia | Mouse | 34 |
| L1210/cis-DDP (NSC-119875) | Lymphoid Leukemia | Mouse | 34 |
| L1210/CTX (NSC-26271) | Lymphoid Leukemia | Mouse | 34 |
| L1210/DF8 (NSC-29630) | Lymphoid Leukemia | Mouse | 34 |
| L1210/DTIC (NSC-45388) | Lymphoid Leukemia | Mouse | 34 |
| L1210/FR8 (Folate Reductase) | Lymphoid Leukemia | Mouse | 35 |
| L1210/FR8/DCM | Lymphoid Leukemia | Mouse | 35 |
| L1210/Ftorafur (NSC-148958) | Lymphoid Leukemia | Mouse | 34 |
| L1210/HU (NSC-32065) | Lymphoid Leukemia | Mouse | 34 |
| L1210/L-PAM (NSC-8806) | Lymphoid Leukemia | Mouse | 34 |
| L1210/M-773 | Lymphoid Leukemia | Mouse | 35 |
| L1210/MeGAG (NSC-32946) | Lymphoid Leukemia | Mouse | 34 |
| L1210/MRI (NSC-153353) | Leukemia | NonHuman *In Vitro* Cell Lines | 66 |
| L1210/MTX (NSC-740) | Lymphoid Leukemia | Mouse | 34 |
| L1210/NSC-19622 | Lymphoid Leukemia | Mouse | 34 |
| L1210/NSC-38280 | Lymphoid Leukemia | Mouse | 34 |
| L1210/TIC (NSC-60339) | Lymphoid Leukemia | Mouse | 34 |
| L1210/TSC (NSC-729) | Lymphoid Leukemia | Mouse | 33 |
| L18464 | Lymphoma | Mouse | 35 |
| L4946 | Lymphocytic Leukemia | Mouse | 35 |
| L5178Y | Leukemia | Mouse | 35 |
| L5178Y (R)/MRI | Leukemia | NonHuman *In Vitro* Cell Lines | 66 |
| $LAF_1$ | Adrenal Cortical Adenocarcinoma | Mouse | 35 |
| LC-12 | Pulmonary Squamous Cell Carcinoma | Mouse | 35 |
| LC-18 | Hepatocarcinoma | Rat | 50 |
| Lewis Lung | Carcinoma | Mouse | 35 |
| Lewis Lung | Carcinoma | NonHuman *In Vitro* Cell Lines | 66 |
| Lewis Lung/PALA (NSC-224131) | Carcinoma | Mouse | 35 |
| Lewis Sarcoma T241 | Pleiomorphic | Mouse | 35 |
| Line 1 | Hepatocarcinoma | Guinea Pig | 22 |
| Line 10 | Hepatocarcinoma | Guinea Pig | 22 |
| LLC-Luc-GFP (LL-LUC-POL2) | Lung Squamous Cell Carcinoma | Mouse | 35 |
| LOVO | Adenocarcinoma | Human *In Vitro* Cell Lines | 59 |
| LOVO I | Colon, Adenocarcinoma | Human | 15 |
| LOVO II | Colon, Adenocarcinoma | Human | 16 |

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