



Effective Health Care Program

Comparative Effectiveness Review
Number 50

Antinuclear Antibody, Rheumatoid Factor, and Cyclic-Citrullinated Peptide Tests for Evaluating Musculoskeletal Complaints in Children



Agency for Healthcare Research and Quality
Advancing Excellence in Health Care • www.ahrq.gov

Antinuclear Antibody, Rheumatoid Factor, and Cyclic-Citrullinated Peptide Tests for Evaluating Musculoskeletal Complaints in Children

Prepared for:

Agency for Healthcare Research and Quality
U.S. Department of Health and Human Services
540 Gaither Road
Rockville, MD 20850
www.ahrq.gov

Contract No. HHSA 290 2007 10021 I

Prepared by:

University of Alberta Evidence-based Practice Center
Edmonton, AB, Canada

Investigators:

Kai O. Wong, M.Sc.
Kenneth Bond, B.Ed., M.A.
Joanne Homik, M.D., M.Sc.
Janet E. Ellsworth, M.D.
Mohammad Karkhaneh, M.D.
Christine Ha, B.Sc.
Donna M. Dryden, Ph.D.

**AHRQ Publication No. 12-EHC015-EF
March 2012**

This report is based on research conducted by the University of Alberta Evidence-based Practice Center under contract to the Agency for Healthcare Research and Quality (AHRQ), Rockville, MD (Contract No. HHS 290 2007 10021 I). The findings and conclusions in this document are those of the author(s), who are responsible for its content, and do not necessarily represent the views of AHRQ. No statement in this report should be construed as an official position of AHRQ or of the U.S. Department of Health and Human Services.

The information in this report is intended to help health care decisionmakers—patients and clinicians, health system leaders, and policymakers, among others—make well-informed decisions and thereby improve the quality of health care services. This report is not intended to be a substitute for the application of clinical judgment. Anyone who makes decisions concerning the provision of clinical care should consider this report in the same way as any medical reference and in conjunction with all other pertinent information, i.e., in the context of available resources and circumstances presented by individual patients.

This report may be used, in whole or in part, as the basis for the development of clinical practice guidelines and other quality enhancement tools, or as a basis for reimbursement and coverage policies. AHRQ or U.S. Department of Health and Human Services endorsement of such derivative products or actions may not be stated or implied.

This document is in the public domain and may be used and reprinted without special permission. Citation of the source is appreciated.

Persons using assistive technology may not be able to fully access information in this report. For assistance contact: EffectiveHealthCare@ahrq.hhs.gov.

None of the investigators have any affiliations or financial involvement that conflicts with the material presented in this report.

Suggested Citation:

Wong KO, Bond K, Homik J, Ellsworth JE, Karkhaneh M, Ha C, Dryden DM. Antinuclear Antibody, Rheumatoid Factor, and Cyclic-Citrullinated Peptide Tests for Evaluating Musculoskeletal Complaints in Children. Comparative Effectiveness Review No. 50 (Prepared by the University of Alberta Evidence-based Practice Center under Contract No. HHS 290 2007 10021 I). AHRQ Publication No. 12-EHC015-EF. Rockville, MD: Agency for Healthcare Research and Quality. March 2012. Effectivehealthcare.ahrq.gov/reports/final.cfm.

Preface

The Agency for Healthcare Research and Quality (AHRQ) conducts the Effective Health Care Program as part of its mission to organize knowledge and make it available to inform decisions about health care. As part of the Medicare Prescription Drug, Improvement, and Modernization Act of 2003, Congress directed AHRQ to conduct and support research on the comparative outcomes, clinical effectiveness, and appropriateness of pharmaceuticals, devices, and health care services to meet the needs of Medicare, Medicaid, and the Children's Health Insurance Program (CHIP).

AHRQ has an established network of Evidence-based Practice Centers (EPCs) that produce Evidence Reports/Technology Assessments to assist public- and private-sector organizations in their efforts to improve the quality of health care. The EPCs now lend their expertise to the Effective Health Care Program by conducting comparative effectiveness reviews (CERs) of medications, devices, and other relevant interventions, including strategies for how these items and services can best be organized, managed, and delivered.

Systematic reviews are the building blocks underlying evidence-based practice; they focus attention on the strength and limits of evidence from research studies about the effectiveness and safety of a clinical intervention. In the context of developing recommendations for practice, systematic reviews are useful because they define the strengths and limits of the evidences, clarifying whether assertions about the value of the intervention are based on strong evidence from clinical studies. For more information about systematic reviews, see www.effectivehealthcare.ahrq.gov/reference/purpose.cfm.

AHRQ expects that CERs will be helpful to health plans, providers, purchasers, government programs, and the health care system as a whole. In addition, AHRQ is committed to presenting information in different formats so that consumers who make decisions about their own and their family's health can benefit from the evidence.

Transparency and stakeholder input are essential to the Effective Health Care Program. Please visit the Web site www.effectivehealthcare.ahrq.gov to see draft research questions and reports or to join an email list to learn about new program products and opportunities for input. Comparative Effectiveness Reviews will be updated regularly.

We welcome comments on this CER. They may be sent by mail to the Task Order Officer named below at: Agency for Healthcare Research and Quality, 540 Gaither Road, Rockville, MD 20850, or by email to epc@ahrq.hhs.gov.

Carolyn M. Clancy, M.D.
Director
Agency for Healthcare Research and Quality

Jean Slutsky, P.A., M.S.P.H.
Director, Center for Outcomes and Evidence
Agency for Healthcare Research and Quality

Stephanie Chang, M.D., M.P.H.
Director, EPC Program
Center for Outcomes and Evidence
Agency for Healthcare Research and Quality

Mary Nix, M.S.
EPC Program Task Order Officer
Agency for Healthcare Research and Quality

Acknowledgements

We are grateful to members of the Technical Expert Panel who provided direction for the scope and content of the review. We also thank all the individuals at AHRQ, the Scientific Resource Center, and the peer reviewers for their comments and suggestions.

We also thank the following University of Alberta Evidence-based Practice Center staff for their assistance: Amy Beath (designed and conducted literature searches), Ben Vandermeer (statistical consultation), Heather McPhee (screening and inclusion/exclusion), and Teodora Radisic (literature retrieval).

Technical Expert Panel

Timothy G. Beukelman, M.D.
University of Alabama at Birmingham
Birmingham, AL

Dennis Dietzen, Ph.D.
Washington University School of Medicine
St. Louis, MO

James N. Jarvis, M.D.
Oklahoma University Health Sciences Center
Oklahoma City, OK

Charles H. Spencer, M.D.
Ohio State University
Columbus, OH

Peer Reviewers

Donald Bloch, M.D.
Massachusetts General Hospital and
Harvard Medical School
Boston, MA

Peter Chira, M.D., M.S.
Stanford University School of Medicine
Stanford, CA

David F. Keren, M.D.
Warde Medical Laboratory
Ann Arbor, MI

Sampath Prahalad, M.B.B.S. (M.D.), M.S.
Emory University School of Medicine
Atlanta, GA

Sarah Ringold, M.D., M.S.
University of Washington School of
Medicine and Seattle Children's Hospital
Seattle, WA

Penny Whiting, Ph.D.
Faculty of Medicine and Dentistry
University of Bristol
Bristol, U.K.

Structured Abstract

Objectives. To assess the test performance of antinuclear antibody (ANA), rheumatoid factor (RF), and cyclic-citrullinated peptide (CCP) tests in children and adolescents with undiagnosed musculoskeletal (MSK) pain or joint swelling, compared with clinical diagnoses of pediatric systemic lupus erythematosus (pSLE) and juvenile idiopathic arthritis (JIA). To explore differences in test performance for accuracy modifiers including age, sex, race or ethnicity, comorbidities, and recent infections. To evaluate the impact of test results on clinical decisionmaking and clinically important outcomes such as referrals, ordering of additional tests, clinical management, and anxiety experienced by children and parents.

Data Sources. We conducted comprehensive searches in nine electronic databases. We also hand searched reference lists and conference proceedings. There were no restrictions on language, year of publication, and study design.

Review Methods. Study selection, quality assessment, data extraction, and grading the evidence were conducted independently by two reviewers. A combination of qualitative and quantitative approaches was used to synthesize the data. We calculated sensitivity (Sn) and specificity (Sp).

Results. The search identified 11,695 citations; 28 were included in the review. Only one cohort study examined the test performance of RF to diagnose JIA among children with undiagnosed MSK pain. It demonstrated an Sn of 5 percent and an Sp of 98 percent. Fifteen case-control studies did not specifically address the test performance of RF among children with MSK pain. The strength of evidence is low for both Sn and Sp. The 12 case-control studies that examined other test-disease combinations did not specifically address the prevalence of positive tests for ANA or CCP among children presenting with undiagnosed MSK pain. The strength of evidence is insufficient to determine the test performance of ANA or CCP to diagnose JIA or pSLE in children with undiagnosed MSK pain. No studies addressed children with joint swelling. There was no evidence addressing the prespecified accuracy modifiers or clinically important outcomes.

Conclusions. Most of the evidence from the 28 studies included in the review was not applicable to the population of interest as most studies examined children with known disease rather than with undiagnosed MSK pain. No studies provided a complete investigation on accuracy modifiers. No studies examined clinically important outcomes such as the impact of the test results on referrals, ordering of additional tests, patient management, and patient and parent anxiety levels. Because the Sn and Sp of these tests have yet to be verified, current evidence does not support their use as diagnostic tests for children with undiagnosed MSK pain. They have a potential application as an adjunct to a clinical assessment that suggests the presence of an inflammatory arthritis or connective tissue disease.

Contents

Executive Summary	ES-1
Introduction	1
Musculoskeletal Symptoms	1
Pediatric Systemic Lupus Erythematosus and Juvenile Idiopathic Arthritis	1
Antinuclear Antibody, Rheumatoid Factor, and Cyclic-Citrullinated Peptide Tests.....	2
Objectives of This Evidence Report	3
Key Questions	4
Analytic Framework	5
Methods	6
Topic Development and Refinement	6
Search Strategy	6
Study Selection	7
Data Extraction	9
Quality Assessment.....	9
Data Analysis and Synthesis.....	9
Rating the Body of Evidence	9
Applicability	10
Peer Review and Public Commentary	10
Results: Part One	11
Key Question 1.1. Incidence and Prevalence of Undiagnosed Musculoskeletal Complaints in Children	11
Key Question 1.2: The Positivity of Antinuclear Antibody, Rheumatoid Factor, and Cyclic-Citrullinated Peptide Tests in Healthy Children	11
Key Question 2.1. Noninflammatory Causes of Pediatric Musculoskeletal Pain.....	11
Key Question 2.2. Inflammatory Causes of Pediatric Musculoskeletal Pain	12
Key Question 2.3. The Resolution or Recurrence of Pediatric Musculoskeletal Pain	12
Results: Part Two	13
Literature Search	13
Characteristics of Included Studies.....	13
Methodological Quality of Included Studies	15
Key Question 3.1. Antinuclear Antibody Test for Pediatric Systemic Lupus Erythematosus.....	16
Study Characteristics	16
Quantitative Results	16
Key Question 3.2. Antinuclear Antibody Test for Juvenile Idiopathic Arthritis.....	18
Study Characteristics	18
Quantitative Results	18
Key Question 3.3. Rheumatoid Ractor Test for Pediatric Systemic Lupus Erythematosus	20
Study Characteristics	20

Quantitative Results	20
Key Question 3.4. Rheumatoid Factor Test for Juvenile Idiopathic Arthritis	20
Study Characteristics	20
Quantitative Results	20
Key Question 3.5. Cyclic-Citrullinated Peptide Test for Pediatric Systemic Lupus Erythematosus	24
Key Question 3.6. Cyclic-Citrullinated Peptide Test for Juvenile Idiopathic Arthritis	24
Study Characteristics	24
Quantitative Results	24
Key Question 4.1. Accuracy Modifiers of Antinuclear Antibody, Rheumatoid Factor, Cyclic-Citrullinated Peptide Test for Pediatric Systemic Lupus Erythematosus	26
Key Question 4.2. Accuracy Modifiers of Antinuclear Antibody, Rheumatoid Factor, Cyclic-Citrullinated Peptide Test for Juvenile Idiopathic Arthritis	26
Key Question 5. Clinical Impacts due to the Results of Antinuclear Antibody, Rheumatoid Factor, and Cyclic-Citrullinated Peptide Tests	26
Rating the Body of Evidence	27
Applicability	28
Summary and Discussion	29
Summary	29
Implications	29
Limitations	30
Conclusion	30
Future Research	30
References and Included Studies	33
Acronyms	34
Tables	
Table A. Summary of Evidence of the Diagnostic Characteristics of ANA, RF, and CCP Tests for pSLE and JIA in Children With Undiagnosed MSK Pain	ES-9
Table 1. Inclusion and Exclusion Criteria	8
Table 2. Description of Studies Evaluating an ANA Test for pSLE in Children With Undiagnosed MSK Pain	17
Table 3. Description of Studies Evaluating an ANA Test for JIA in Children With Undiagnosed MSK Pain	19
Table 4. Description of Studies Evaluating an RF (IgM) Test for JIA in Children With Undiagnosed MSK Pain	22
Table 5. Description of Studies Evaluating a CCP Test for Juvenile Idiopathic Arthritis in Children With Undiagnosed MSK Pain	25
Table 6. Strength of Evidence for ANA, RF, and CCP Tests for pSLE and JIA in Children With Undiagnosed MSK Pain	27
Table 7. Hypothetical Scenarios for PPV at Different Baseline Disease Prevalence	28
Table 8. Summary of Evidence of the Diagnostic Characteristics of ANA, RF, and CCP Tests for pSLE and JIA in Children With Undiagnosed MSK Pain	32

Figures

Figure 1. Analytic Framework for Antibody Testing for MSK Complaints in Pediatric Populations (≤ 18 years).....	6
Figure 2. Results of Literature Search, Retrieval, and Selection	14
Figure 3. Methodological Quality of Studies Evaluating ANA, RF, and CCP Tests for pSLE and JIA in Children With Undiagnosed MSK Pain	15
Figure 4. Sensitivity and Specificity of an ANA Test for pSLE in Children With Undiagnosed MSK Pain	16
Figure 5. Sensitivity and Specificity of an ANA Test for JIA in Children With Undiagnosed MSK Pain	18
Figure 6. Sensitivity and Specificity of an RF (IgM) Test for JIA in Children With Undiagnosed MSK Pain	21
Figure 7. Sensitivity and Specificity of a CCP Test for JIA in Children With Undiagnosed MSK Pain	24

Appendixes

Appendix A. American College of Rheumatology Criteria for Classification of Systemic Lupus Erythematosus	
Appendix B. Literature Search String	
Appendix C. Forms	
Appendix D. List of Excluded Studies	
Appendix E. Methodological Quality of Included Studies	
Appendix F. Evidence Tables	
Appendix G. Subgroup Analyses by Onset Type of Juvenile Idiopathic Arthritis	

Executive Summary

Background

Musculoskeletal (MSK) pain is common in children and adolescents, with an estimated prevalence ranging from 2 to 50 percent.¹ MSK pain can affect physical, psychological, and social function and often prompts consultation with a physician.² However, MSK pain is often nonspecific, which can make it difficult to arrive at an accurate diagnosis.^{3,4}

MSK pain may be due to rheumatic or nonrheumatic causes. Nonrheumatic causes are more common, generally benign, and most often attributable to trauma, overuse, and normal bone growth.⁵ Rheumatic causes, such as inflammatory arthritis, are infrequent, generally chronic, and require accurate, timely diagnosis and effective intervention to prevent progression and long-term damage.⁶ Common rheumatic causes of childhood MSK pain include pediatric systemic lupus erythematosus (pSLE) and juvenile idiopathic arthritis (JIA).

A complete history and physical examination is generally considered to be the best way to make a diagnosis of inflammatory arthritis.^{3,5} Physicians may request serological tests such as antinuclear antibody (ANA), rheumatoid factor (RF), and cyclic-citrullinated peptide (CCP) when children and adolescents are suspected of having inflammatory arthritis, despite the fact that the diagnostic performance, usefulness, and proper interpretation of these tests are uncertain in pediatric populations.

This comparative effectiveness review summarizes the evidence on the test performance of ANA, RF, or CCP tests for pSLE and JIA in children with undiagnosed MSK pain. The report is intended for a broad audience including primary care physicians who may consider ordering these tests in a child with MSK pain, health payers who provide coverage for these tests, and parents or caregivers who want to know whether these tests can determine if their child does or does not have a particular disease.

Key Questions

In order to better understand how the ANA, RF, and CCP tests perform in the clinical setting of a child with undiagnosed MSK pain, it is important to know the prevalence of MSK complaints (including MSK pain and joint swelling) in children who do not have JIA and pSLE. It is also important to be aware of the rate of false positives for these tests (i.e., the proportion of otherwise healthy children who have a positive ANA, RF, or CCP test). Appropriate interpretation of test performance also requires an understanding of the disease progression and changes in signs and symptoms in children with MSK pain who may or may not also have JIA or pSLE.

In addition to providing this background information, the objectives of this report were to assess the test performance of ANA, RF, and CCP tests in children and adolescents with undiagnosed MSK pain and/or joint swelling compared with clinical diagnoses of pSLE and JIA; to explore the difference in test performance for accuracy modifiers including age, sex, race or ethnicity, comorbidities, and recent infections; and to evaluate the impact of test results on clinical decisionmaking and clinically important outcomes such as referrals, ordering of additional tests, clinical management, and anxiety experienced by children and parents. We addressed the following Key Questions (KQs):

KQ 1. Prevalence and Incidence

KQ 1.1. In children and adolescents aged 18 years or less, what is the incidence and prevalence of undiagnosed MSK complaints?

KQ 1.2. In healthy children and adolescents aged 18 years or less, what is the incidence of positive test results in ANA, RF, and CCP?

KQ 2. Natural History

KQ 2.1. What proportion of children and adolescents aged 18 years or less with undiagnosed MSK pain is due to noninflammatory causes?

KQ 2.2. What proportion of children and adolescents aged 18 years or less with undiagnosed MSK pain is due to inflammatory causes?

KQ 2.3. What proportion of children and adolescents aged 18 years or less experiences symptom resolution or recurrence?

KQ 3. Diagnostic Performance

KQ 3.1. In children and adolescents aged 18 years or less with undiagnosed MSK pain and/or joint swelling, what is the test performance (sensitivity [Sn], specificity [Sp], and positive and negative predictive values [PPV, NPV]) of ANA for pSLE compared with a clinical diagnosis?

KQ 3.2. In children and adolescents aged 18 years or less with undiagnosed MSK pain and/or joint swelling, what is the test performance (Sn, Sp, PPV, NPV) of ANA for JIA compared with a clinical diagnosis?

KQ 3.3. In children and adolescents aged 18 years or less with undiagnosed MSK pain and/or joint swelling, what is the test performance (Sn, Sp, PPV, NPV) of RF for pSLE compared with a clinical diagnosis?

KQ 3.4. In children and adolescents aged 18 years or less with undiagnosed MSK pain and/or joint swelling, what is the test performance (Sn, Sp, PPV, NPV) of RF for JIA compared with a clinical diagnosis?

KQ 3.5. In children and adolescents aged 18 years or less with undiagnosed MSK pain and/or joint swelling, what is the test performance (Sn, Sp, PPV, NPV) of CCP for pSLE compared with a clinical diagnosis?

KQ 3.6. In children and adolescents aged 18 years or less with undiagnosed MSK pain and/or joint swelling, what is the test performance (Sn, Sp, PPV, NPV) of CCP for JIA compared with a clinical diagnosis?

KQ 4. Accuracy Modifiers

KQ 4.1. In children and adolescents aged 18 years or less with undiagnosed MSK pain and/or joint swelling, do age, sex, race/ethnicity, comorbidities, and recent infections modify the diagnostic performance of ANA, RF, and CCP for pSLE compared with a clinical diagnosis?

KQ 4.2. In children and adolescents aged 18 years or less with undiagnosed MSK pain and/or joint swelling, do age, sex, race/ethnicity, comorbidities, and recent infections modify the diagnostic performance of ANA, RF, and CCP for JIA compared with a clinical diagnosis?

KQ 5. Clinical Impacts of Test Results

In children and adolescents aged 18 years or less with undiagnosed MSK pain and/or joint swelling, do ANA, RF, and CCP test results affect referral decisions, additional tests ordered, clinical management, and patient and parent anxiety due to the clinical uncertainty and additional tests?

Methods

KQs 1 and 2, serving as background information, were addressed in a narrative approach by locating and summarizing the related prevalence, incidence, and natural history information from the main search (described below) and additional searches using MEDLINE[®] and Google Scholar. For KQs 3 to 5, we followed standard methods for conducting comparative effectiveness reviews; these methods were outlined in a prospectively developed protocol.

We searched electronic databases including MEDLINE[®], Cochrane Central Register of Controlled Trials, Cochrane Database of Systematic Reviews (CDSR), Embase, Cumulative Index to Nursing and Allied Health Literature (CINAHL[®]), Science Citation Index Expanded[®] and Social Sciences Citation Index[®] (both via Web of Science[®]), Academic Search Complete, Proquest Dissertations & Theses, and OCLC PapersFirst. In addition, we searched conference proceedings from key scientific meetings, grey literature, and reference lists of included studies. We applied a diagnostic search filter and a child filter, when applicable. We conducted the original searches from 1960 to January 2010, and updated them in December 2010 and September 2011.

Two reviewers independently screened the search results (titles and abstracts) to determine if an article met broad inclusion criteria. The full text of potentially relevant articles was assessed independently by two reviewers using detailed standardized criteria. Two reviewers independently assessed the methodological quality of individual studies using the QUADAS tool.⁷ Data were extracted by one reviewer and verified by another using a standardized data extraction form. For each of these steps, disagreements were resolved through discussion or third-party adjudication, as needed.

We examined the diagnostic test characteristics, including Sn, Sp, PPV, and NPV, for each study and presented forest plots to summarize the results for each test–disease pairing. Accuracy modifiers including age, sex, race or ethnicity, recent infection, and comorbidity were analyzed if studies provided sufficient data to calculate Sn and Sp. We examined any qualitative or

quantitative information on clinically important outcomes including referral, additional tests ordered, change in clinical management, and patient or parent anxiety due to the test results.

Two reviewers independently assessed the strength of evidence for KQs 3 to 5 using the Agency for Healthcare Research and Quality (AHRQ) system for grading evidence (AHRQ Guidance for the Evaluation of Medical Tests [draft]).⁸ We assessed the strength of evidence for Sn and Sp. Assessments were based on the quantity and quality of individual studies, the directness of evidence, and the consistency and precision of the results. For each outcome, the strength of evidence was graded as high, moderate, low, or insufficient.

Results

KQ 1.1. The Prevalence of Undiagnosed MSK Complaints in Children and Adolescents

The prevalence of MSK pain ranged between 2 and 52 percent⁹⁻¹¹ and increased steadily with age throughout childhood and adolescence.^{12,13} No studies reported the prevalence of joint swelling in children.

KQ 1.2. The Prevalence of Test Positivity in Healthy Children and Adolescents

The prevalence of positive ANA in healthy children ranged from 0 to 18 percent.¹⁴⁻²² The prevalence of positive RF in healthy children was estimated at 3 percent.²³ The prevalence of CCP positivity in healthy children was reported in two studies and ranged from 0 to 0.6 percent.^{24,25}

KQ 2. The Etiology and Resolution of Pediatric MSK Pain

Noninflammatory etiologies accounted for the MSK pain in almost all (97 percent) children seen in a primary care setting.¹² Physical trauma was the most common noninflammatory cause and accounted for 44 percent of children with MSK pain. In contrast, only 3.3 percent of children had their MSK pain attributed to inflammatory causes including toxic synovitis (2.5 percent) and inflammatory arthritides (0.8 percent). The recurrence rates of pediatric MSK pain were generally high and varied considerably by site of the pain.

KQ 3. Test Performance of ANA, RF, and CCP

One cohort study and 27 case-control studies addressed KQ 3 (diagnostic performance). In studies using the case-control design, children with known disease (i.e., JIA or pSLE) were compared with children who were healthy (i.e., the control group). This does not represent the target population of children with undiagnosed MSK pain, and therefore, these studies are at high risk of spectrum bias. None of the case-control studies provided information about the presence of MSK pain in either the cases or controls. None of the studies specifically addressed children with joint swelling.

KQ 3.1. ANA Test for pSLE in Children With MSK Pain

Two case-control studies^{26,27} including 201 children (67 pSLE, 134 controls) examined the prevalence of a positive ANA test in children with pSLE and control groups including healthy children and children scheduled for elective orthopedic surgery. The Sn's were 91 and 100 percent, and Sp's were 84 and 85 percent (Table A).

KQ 3.2. ANA Test for JIA in Children With MSK Pain

Eight case-control studies^{26,28-34} including 1,382 children (1,067 JIA, 315 controls) examined the prevalence of a positive ANA test in children with JIA and controls including healthy children, children with nonrheumatic conditions, and children with other rheumatic diseases.. The Sn ranged from 1 to 62 percent, and Sp ranged from 73 to 100 percent (Table A).

KQ 3.3. RF Test for pSLE in Children With MSK Pain

One case-control study³⁵ with 46 children (14pSLE, 32 controls) examined the prevalence of a positive IgM-RF test for pSLE. The control group comprised healthy children and children with other rheumatic conditions or ulcerative colitis. The Sn was 29 percent, and Sp was 88 percent (Table A).

KQ 3.4. RF Test for JIA in Children With MSK Pain

One retrospective cohort study³⁶ examined the records of pediatric patients who had an RF test and were seen at a children's hospital. Among the 437 patient records, 105 had a diagnosis of JIA. The remaining 332 patients had a mix of MSK complaints (n = 201) or symptoms suggestive of an underlying autoimmune disease (n = 131). The Sn was 5 percent, and Sp was 98 percent (Table A).

Fifteen case-control studies^{28,30,33,35-47} including 1,647 children (986 JIA, 661 controls) examined the prevalence of a positive IgM-RF test in children with JIA and controls. The control groups included healthy children, children with nonrheumatic conditions, and children with other rheumatic conditions. The Sn ranged from 0 to 35 percent, and Sp ranged from 94 to 100 percent (Table A).

KQ 3.5. CCP Test for pSLE in Children With MSK Pain

No studies provided information to address this question.

KQ 3.6. CCP Test for JIA in Children With MSK Pain

Seven case-control studies^{24,25,30,48-51} including 1,643 participants (729 JIA, 914 controls) examined the prevalence of a positive CCP test in children with JIA and controls including healthy children, children with nonrheumatic conditions, and children with other autoimmune diseases. Sn ranged from 2 to 42 percent, and Sp ranged from 93 to 100 percent (Table A).

KQ 4. Accuracy Modifiers of ANA, RF, and CCP Tests

No studies provided data on accuracy modifiers (age, sex, race or ethnicity, comorbidities, recent infections) for any of the tests.

KQ 5. Clinical Impacts of ANA, RF, and CCP Tests

No studies provided information to address this question.

Summary

Studies that have investigated the prevalence of MSK pain in children report a wide range of prevalence from 2 to 52 percent. Noninflammatory causes of MSK pain account for the majority of diagnoses (97 percent). Among the healthy children, the median ANA positivity is 3 percent, median RF positivity is 0 percent, and CCP positivity is less than 1 percent.

Only one retrospective cohort study examined the diagnostic test characteristics of RF to diagnose JIA among children with undiagnosed MSK pain compared with a clinical diagnosis. It demonstrated a Sn of 5 percent and a Sp of 98 percent. Fifteen case-control studies did not specifically address the test performance of RF among children with undiagnosed MSK pain. The strength of evidence is low for both Sn and Sp (Table A). Further evidence is likely to change our confidence in the estimates of performance and is likely to change the estimates.

The 12 case-control studies looking at other test-disease combinations did not specifically address the prevalence of positive tests for ANA or CCP among children presenting with undiagnosed MSK pain. The strength of evidence is insufficient to determine the test performance of ANA or CCP to diagnose JIA or pSLE in children with undiagnosed MSK pain. No studies specifically addressed children with joint swelling.

A general pattern of high Sp and low Sn was observed for almost all the test-disease combinations; however, the design of case-control studies may lead to bias.⁵²⁻⁵⁴ The selective inclusion of cases with established disease (i.e., JIA or pSLE) is likely to lead to an overestimation of Sn. The inclusion of healthy controls is expected to decrease the likelihood of false positive test results and lead to an overestimation of Sp.

Implications

There is insufficient evidence to determine the test performance of ANA or CCP in children with undiagnosed MSK pain. The strength of evidence is low for the utility of RF in the diagnosis of JIA in children with undiagnosed MSK pain. A result of high Sp and low Sn was observed for almost all the test-disease combinations. The generally low Sn suggests that it is inappropriate to use these tests in isolation (i.e., without clinical assessment) to make a diagnosis of JIA and pSLE. In spite of the high Sp, the low prevalence of JIA and pSLE in the target population (i.e., children with undiagnosed MSK pain) makes the tests of limited diagnostic value. The presence of other clinical characteristics (e.g., morning stiffness, joint swelling, malar rash, cytopenia) may increase the pretest probability of the disease in question. While both the Sn and Sp for ANA for pSLE were high, this test in isolation has limited diagnostic value for children with undiagnosed MSK given the very low prevalence of pSLE, and up to 18 percent prevalence of a false positive ANA in the general population.

Limitations

The generally insufficient strength of evidence is primarily attributable to the high risk of spectrum bias of the case-control studies, a result of the distinct disease and control groups not being representative of the target population of children with undiagnosed MSK pain. For studies examining ANA for pSLE, incorporation bias is a concern because ANA is considered one of the classification criteria for SLE.⁵⁵

There is no evidence with which to assess the impact of potential accuracy modifiers, and there is no evidence with which to assess the clinical utility of the tests including the impact of the test results on referrals, ordering of additional tests, patient management, and patient and parent anxiety levels.

In addition to the issues identified above, there are general limitations for systematic reviews such as publication bias. We addressed this issue by conducting a comprehensive search of the published literature for potentially relevant studies. Search strategies included combinations of subject headings and free text words. Even though we applied a diagnostic search filter to the search strategies of the electronic databases, our searches identified over 11,000 records. Furthermore, these searches were supplemented by hand searching for grey literature (i.e., unpublished or difficult to find studies). There is also a possibility of study selection bias. However, we employed at least two independent reviewers to identify potentially relevant studies, and feel confident that the studies that were excluded from this report were done so for consistent and appropriate reasons.

Conclusion

Most of the evidence from the 28 studies included in this review was not applicable to the population of interest as studies examined children with known disease rather than with undiagnosed MSK pain. No studies specifically addressed children with joint swelling. No study provided a complete investigation on accuracy modifiers. No studies examined clinically important outcomes such as the impact of the test results on referrals, ordering of additional tests, patient management, and patient and parent anxiety levels.

Because the Sn and Sp of these tests have yet to be verified, current evidence does not support their use as diagnostic tests for children with undiagnosed MSK pain. They have a potential application as an adjunct to a clinical assessment that suggests the presence of an inflammatory arthritis or connective tissue disease.

Future Research

The following general recommendations for future research are based on the preceding discussion of the evidence.

- In order to better understand the natural history of undiagnosed MSK pain in children and the probability of a diagnosis of JIA or pSLE in this population, prospective cohort studies of children and adolescents with MSK pain are needed. Given the low prevalence of JIA or pSLE, a sufficiently large number of participants is required.
- For the research to be generalizable, researchers need to use consistent test methodology and cutoffs as well as consistent and well-accepted clinical criteria for the diagnoses of JIA and pSLE.

- Potential accuracy modifiers of test performance need to be examined, including age, sex, race, history of recent infections, presence of clinical characteristics other than MSK pain (e.g., morning stiffness, joint swelling, uveitis, malar rash, cytopenias).
- The clinical impact of these tests (e.g., referral decisions, additional tests ordered, clinical management, quality of life, psychological distress of child and/or parents) should be assessed in cohort studies.
- Efforts are needed to improve the overall quality of reporting of primary studies of diagnostic test accuracy. The STARD checklist includes 25 items that address the level of detail that should be specified within such studies including descriptions of participants, tests methods, statistical methods, and results.⁵⁶ This could be considered as a guide for authors reporting studies that evaluate diagnostic tests.

Table A. Summary of evidence of the diagnostic characteristics of ANA, RF, and CCP tests for pSLE and JIA in children with undiagnosed MSK pain

Key Questions	N Studies, Sample Size	Sensitivity Range (median)*	Specificity Range (median)	PPV Range (median)	NPV Range (median)	Strength of Evidence
KQ 3: Test performance						
3.1 ANA – pSLE	2 case-control, 201	91–100%	84–85%	71–84%	96–100%	Insufficient
3.2 ANA – JIA	8 case-control, 1,382	1–62% (54)	73–100% (95)	88–100% (96)	15–70% (30)	Insufficient
3.3 RF (IgM) – pSLE	1 case-control, 46	29%	88%	50%	74%	Insufficient
3.4 RF (IgM) – JIA	1 cohort study, 437	5%	98%	45%	77%	Low
	15 case-control, 1,647	0–35% (11)	94–100% (100)	0–100% (100)	20–71% (48)	Insufficient
3.5 CCP – pSLE	No studies					Insufficient
3.6 CCP – JIA	7 case-control, 1,643	2–42% (6)	93–100% (100)	20–100% (100)	11–71% (28)	Insufficient
KQ 4: Accuracy modifiers	No studies	NA	NA	NA	NA	Insufficient
KQ 5: Clinical impacts	No studies	NA	NA	NA	NA	Insufficient

*Median not presented if ≤ 4 studies.

ANA = antinuclear antibody; CCP = cyclic-citrullinated peptide; IgM = immunoglobulin M; JIA = juvenile idiopathic arthritis; KQ = Key Question; MSK = musculoskeletal; N = number; NA = not applicable; NPV = negative predictive value; PPV = positive predictive value; pSLE = pediatric systemic lupus erythematosus; RF = rheumatoid factor

Introduction

Musculoskeletal Symptoms

Musculoskeletal (MSK) pain is pain that affects muscles, bones, ligaments, tendons, and nerves.⁵⁷ Childhood MSK pain is common, with estimated prevalence ranging from 2 to 50 percent.^{1,10,12} Young children especially may have difficulty characterizing their symptoms, which makes accurate assessment based on the patient history difficult. In addition, the presence of MSK pain can cause anxiety among children and their parents. Concerns about the presence of a serious condition such as arthritis or lupus, which could lead to permanent damage, may prompt consultation with a physician.⁵⁸

MSK pain can be divided into rheumatic and nonrheumatic causes. Nonrheumatic causes account for the majority of childhood MSK pain and are generally attributable to benign conditions including minor physical trauma (i.e., sprains and strains), overuse, and normal body growth.^{3,5} In contrast, rheumatic causes, such as an inflammatory arthritis, are much less prevalent and are generally chronic and require early diagnosis and treatment to prevent progression and disability. Common rheumatic causes of childhood MSK pain include pediatric systemic lupus erythematosus (pSLE), juvenile idiopathic arthritis (JIA), spondyloarthropathies (including enthesitis, juvenile ankylosing spondylitis, and reactive arthritis), acute rheumatic fever, and Henoch-Schonlein purpura. However, MSK pain is not universally present in children with JIA (16 percent of children with JIA do not report pain⁵⁹) and pSLE.

A complete history and physical examination is generally considered to be the best way to make a diagnosis of inflammatory arthritis.^{3,5} However, the complaint of MSK pain is often nonspecific and when combined with a lack of confidence in the MSK physical examination, can make it difficult to arrive at an accurate diagnosis.^{3,4} Hence, physicians may request additional laboratory tests. Serological tests such as antinuclear antibody (ANA), rheumatoid factor (RF), and cyclic-citrullinated peptide (CCP) may be ordered by physicians when children and adolescents are suspected of having a rheumatic cause for their MSK pain, despite the fact that the diagnostic performance and usefulness of these tests and the proper interpretation of the results for pediatric populations are largely uncertain.

This comparative effectiveness review provides a synthesis of the evidence on the test performance of ANA, RF, and CCP tests in children and adolescents with undiagnosed MSK pain and on the impact of test results on clinical decisionmaking and clinically important outcomes. The report is intended for a broad audience including: primary care physicians who may consider ordering ANA, RF, or CCP tests in a child with MSK pain; health payers who provide coverage for these tests; and parents or caregivers who would like to know whether these tests can determine if their child does or does not have a particular disease.

Pediatric Systemic Lupus Erythematosus and Juvenile Idiopathic Arthritis

Systemic lupus erythematosus (SLE) is an episodic, multisystem, autoimmune disease characterized by widespread inflammation of blood vessels and connective tissues.⁶⁰ It is estimated that the incidence of pSLE is 0.3 to 0.9 per 100,000 children per year⁶¹ and the prevalence is 3.3 to 8.8 per 100,000.⁶² The onset of pSLE is rare before 5 years of age and

uncommon before adolescence, after which the rates of occurrence stabilize.⁶⁰ The diagnosis of pSLE is generally based on the classification criteria of the American College of Rheumatology (ACR)^{55,63} which include specific signs, symptoms, and laboratory tests, including a positive ANA (see Appendix A). Left untreated, SLE is often progressive and can be fatal.⁶⁴ As awareness of the occurrence of pSLE has increased, early diagnosis has become more common⁶⁰ and rapid introduction of effective immunosuppressive treatment has led to improved outcomes.⁶⁴

JIA is the most common chronic inflammatory disease of children affecting approximately 1 in 1,000 children.^{65,66} Classification criteria developed by the International League of Associations for Rheumatology (ILAR)⁶⁷ are used worldwide to provide consistency across clinical research studies. The ILAR criteria have supplanted earlier criteria of the ACR⁶⁸ for the classification of juvenile rheumatoid arthritis (JRA) and the European League Against Rheumatism (EULAR)⁶⁹ for the classification of juvenile chronic arthritis (JCA). To maintain consistency, the acronym JIA will be used to represent JIA, JRA, and JCA throughout this report.

It is important to note that in all three of the published criteria patients are classified based on characteristic symptoms and signs, including the presence of objective arthritis for a minimum of 6 to 12 weeks. In spite of significant overlap, the classification criteria vary in terms of how the presence of RF is addressed. In the ILAR criteria, the presence of a positive RF on two occasions *excludes* five of the seven subtypes of JIA. In the EULAR criteria, a positive RF changes the classification from JCA to JRA. In the ACR criteria, RF is not considered at all in the classification. Therefore, depending on the criteria being used, the reported prevalence of RF will be different.

In adults with suspected rheumatoid arthritis, tests for RF⁷⁰ and, more recently, CCP antibodies are frequently requested as part of the diagnostic work-up.^{70,71} Although there is less evidence supporting the usefulness of these tests in children, they are often ordered as part of the diagnostic evaluation of a child suspected to have JIA.

Without effective treatment, JIA can progress and cause damage to cartilage, bone, and soft tissues, and may lead to severe disability, functional loss, and, in rare cases, organ failure and death.^{66,71} Although early diagnosis and treatment may reduce the progression of the disease and induce remission, only a minority of patients experience complete resolution of JIA prior to adulthood.⁶⁶

Antinuclear Antibody, Rheumatoid Factor, and Cyclic-Citrullinated Peptide Tests

The immune system is a defense system that fends off foreign invaders including bacteria and viruses. However, the immune system may malfunction and mislabel one's own body cells as foreign particles, and this may elicit an attack response. When the immune system attacks one's own body cells, it produces autoantibodies that target specific antigens naturally found in the body. ANA, RF, and CCP are examples of the autoantibodies specifically targeting the nuclear particles, the fragment crystallizable (Fc) portion of the immunoglobulin (Ig) G, and CCPs, respectively. An ANA test is often used to screen for autoimmune conditions,⁷⁴ especially when a diagnosis of SLE is suspected.

The gold standard for ANA testing is the indirect immunofluorescence (IIF) ANA test, which involves incubation of serial dilutions of the patient's sera with substrate cells, usually human

epithelial tumor line (HEp-2).⁷² If antibody to nuclear elements is present, binding to the substrate will be detected by fluorescein-conjugated anti-human Ig, which attaches to the antibody and is visually inspected using a fluorescence microscope. The assessment of fluorescence is based on the interpretation of this inspection and, as a result, may be somewhat subjective and vary from one laboratory to another. Each laboratory determines the cutoff used for a positive test, and as a result, titers from one laboratory cannot be compared with another. Research has shown that using a high titer ANA does not increase the positive predictive value for connective tissue disease.⁷³

The detection of antibodies may also be assessed using enzyme immunoassay (EIA). In EIA, an antigen is affixed to a surface, and then the patient serum sample is applied over the surface so ANA, if present, can bind to the antigen. EIA methods and expected results vary among manufacturers because there is no agreed standard for the antigen preparations that should be included or for the concentration(s) of the relevant antigen preparations.⁷⁴ Results of studies that compare IIF and EIA for ANA have been inconsistent, with some showing poor correlation,²⁶ and others demonstrating consistency.⁷⁴

RFs are Ig that react specifically with the Fc fragment of the IgG molecule. RFs are found in all Ig isotypes (i.e., IgA, IgG, IgD, IgM, and IgE), but the 19S IgM-RF is the most frequently used isotype for rheumatic disease testing, including arthritis.⁷⁵ The presence of RFs is typically determined by agglutination assays, nephelometry, or EIA. The agglutination assay method mainly employs latex beads as a substrate to which human or rabbit IgG is bound. Nephelometry is a photometric test in which complexes formed between the IgG and RF are detected by light scattering, which is dependent upon the concentration of those immune complexes. Latex agglutination and nephelometry only measure 19S IgM-RF, whereas EIAs have been designed to measure the various RF isotypes.⁷⁵

The CCP test detects the presence of autoantibodies to citrullinated peptides in serum.⁷⁶ Abnormal citrullination of various peptides is present in a variety of human diseases, including rheumatoid arthritis, psoriasis, and multiple sclerosis. However, the formation of antibodies to citrullinated peptides seems to be specific for adult patients with rheumatoid arthritis.⁷¹ Anti-CCP2 (a second-generation assay) is currently the most widely used anti-CCP assay.⁷¹ Anti-CCP antibodies and anti-citrullinated filaggrin antibodies are locally produced in inflamed joints, and citrullinated fibrin is found in the synovia of patients with rheumatoid arthritis.⁷⁷ In adults, a CCP test is usually ordered along with a RF test when evaluating a patient with inflammatory arthritis and when rheumatoid arthritis is considered on the differential diagnosis. The utility of the CCP test in pediatric rheumatic conditions is not clear.

Objectives of This Evidence Report

In order to better understand how the ANA, RF, and CCP tests perform in the clinical setting in which a child with MSK pain will be seen, it is important to know the prevalence of MSK complaints (including MSK pain and joint swelling) in children who do not have JIA and pSLE. It is also important to be aware of the rate of false positives for these tests (i.e., the proportion of otherwise healthy children who have a positive ANA, RF, or CCP). The appropriate interpretation of test performance also requires an understanding of the disease progression and changes in signs and symptoms in children with MSK pain who may or may not also have JIA or pSLE.

In addition to providing this background information, the objectives of this report were to assess the test performance of ANA, RF, and CCP tests in children and adolescents with

undiagnosed MSK pain and/or joint swelling compared with clinical diagnoses of pSLE and JIA; to explore the difference in test performance for accuracy modifiers including age, sex, race or ethnicity, comorbidities, and recent infections; and, to evaluate the impact of test results on clinical decisionmaking and clinically important outcomes such as referrals, ordering of additional tests, clinical management, and anxiety experienced by children and parents. We addressed the following Key Questions (KQs):

Key Questions

KQ 1. Prevalence and Incidence

KQ 1.1. In children and adolescents aged 18 years or less, what is the incidence and prevalence of undiagnosed MSK complaints?

KQ 1.2. In healthy children and adolescents aged 18 years or less, what is the incidence of positive test results in ANA, RF, and CCP?

KQ 2. Natural History

KQ 2.1. What proportion of children and adolescents aged 18 years or less with undiagnosed MSK pain is due to non-inflammatory etiologies?

KQ 2.2. What proportion of children and adolescents aged 18 years or less with undiagnosed MSK pain is due to inflammatory etiologies?

KQ 2.3. What proportion of children and adolescents aged 18 years or less experiences symptom resolution or recurrence?

KQ 3. Diagnostic Performance

KQ 3.1. In children and adolescents aged 18 years or less with undiagnosed MSK pain and/or joint swelling, what is the test performance (sensitivity, specificity, and positive and negative predictive values) of ANA for pSLE compared with a clinical diagnosis?

KQ 3.2. In children and adolescents aged 18 years or less with undiagnosed MSK pain and/or joint swelling, what is the test performance (sensitivity, specificity, and positive and negative predictive values) of ANA for JIA compared with a clinical diagnosis?

KQ 3.3. In children and adolescents aged 18 years or less with undiagnosed MSK pain and/or joint swelling, what is the test performance (sensitivity, specificity, and positive and negative predictive values) of RF for pSLE compared with a clinical diagnosis?

KQ 3.4. In children and adolescents aged 18 years or less with undiagnosed MSK pain and/or joint swelling, what is the test performance (sensitivity, specificity, and positive and negative predictive values) of RF for JIA compared with a clinical diagnosis?

KQ 3.5. In children and adolescents aged 18 years or less with undiagnosed MSK pain and/or joint swelling, what is the test performance (sensitivity, specificity, and positive and negative predictive values) of CCP for pSLE compared with a clinical diagnosis?

KQ 3.6. In children and adolescents aged 18 years or less with undiagnosed MSK pain and/or joint swelling, what is the test performance (sensitivity, specificity, and positive and negative predictive values) of CCP for JIA compared with a clinical diagnosis?

KQ 4. Accuracy Modifiers

KQ 4.1. In children and adolescents aged 18 years or less with undiagnosed MSK pain and/or joint swelling, do age, sex, race/ethnicity, comorbidities, and recent infections modify the diagnostic performance (sensitivity, specificity, and positive and negative predictive values) of ANA, RF, and CCP for pSLE compared with a clinical diagnosis?

KQ 4.2. In children and adolescents aged 18 years or less with undiagnosed MSK pain and/or joint swelling, do age, sex, race/ethnicity, comorbidities, and recent infections modify the diagnostic performance (sensitivity, specificity, and positive and negative predictive values) of ANA, RF, and CCP for JIA compared with a clinical diagnosis?

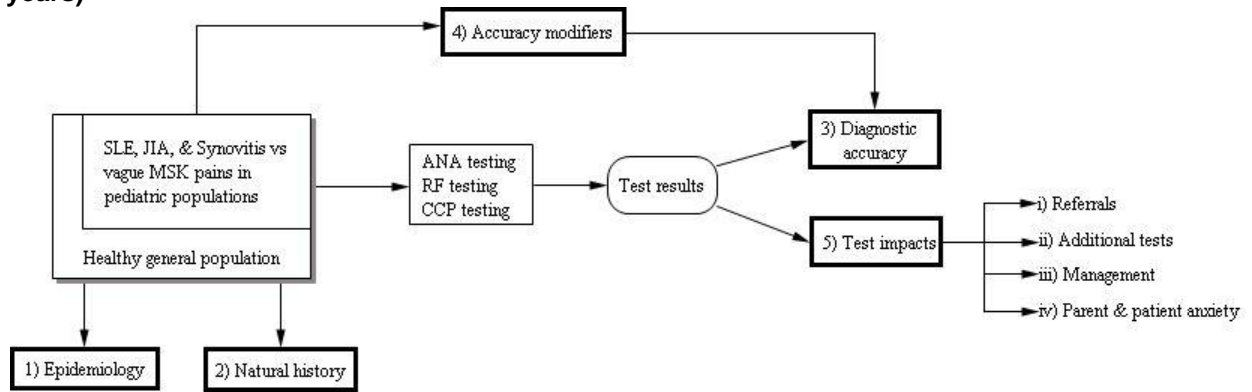
KQ 5. Clinical Impacts of Test Results

KQ 5. In children and adolescents aged 18 years or less with undiagnosed MSK pain and/or joint swelling, do ANA, RF, and CCP test results affect referral decisions, additional tests ordered, clinical management, and patient and parent anxiety due to the clinical uncertainty and additional tests?

Analytic Framework

The analytic framework (Figure 1) depicts the five KQs within the context of the pediatric population (≤ 18 years) with MSK complaints. In general, the figure illustrates how diagnostic accuracy may be modified by demographic and clinical factors. It also indicates how test results may influence four important areas including referral to specialists, additional tests, decisions regarding clinical management, and parents' and patients' level of anxiety. The epidemiology and natural history of the targeted rheumatic conditions are described independently of the test results.

Figure 1. Analytic framework for antibody testing for MSK complaints in pediatric populations (≤18 years)



ANA = antinuclear antibody; CCP = cyclic-citrullinated peptide; JIA = juvenile idiopathic arthritis; MSK = musculoskeletal; SLE = systemic lupus erythematosus; RF = rheumatoid factor

Methods

Topic Development and Refinement

In this chapter we document a prospectively developed protocol that was used to conduct this comparative effectiveness review. A core research team was assembled by the University of Alberta Evidence-based Practice Center. In consultation with the Agency for Healthcare Research and Quality (AHRQ), a panel of key informants was created to provide input in the development of the Key Questions (KQs) and scope of the evidence report. The public was invited to comment on these KQs over a period of 1 month. After reviewing the public comments, the KQs were finalized and submitted to AHRQ for approval. A technical expert panel was subsequently created to provide content and methodological expertise throughout the development of the comparative effectiveness review.

Search Strategy

The research librarian, in collaboration with the investigative team, developed and implemented search strategies designed to identify evidence relevant to questions of diagnostic performance and clinical impact of the tests (Appendix B).

KQs 1 and 2, the answers to which serve as background information, were addressed using a narrative approach by locating and summarizing information on the related disease prevalence, incidence, and natural history from the main search (described below) and additional searches using MEDLINE[®] and Google Scholar. As the primary focuses of this report, KQs 3, 4, and 5 were addressed by a rigorous systematic review process including a comprehensive search of the following electronic databases: MEDLINE[®], Cochrane Central Register of Controlled Trials, Cochrane Database of Systematic Reviews, Embase, CINAHL[®], Science Citation Index Expanded[®] and Social Sciences Citation Index[®] (both via Web of Science[®]), Academic Search Complete, Proquest Dissertations & Theses, and OCLC PapersFirst. We applied a diagnostic search filter and a child filter, when applicable. We conducted the original searches from 1960 to January 2010, and updated them in December 2010 and September 2011.

Search terms were identified by reviewing search strategies of systematic reviews on similar topics and by examining how potentially relevant studies were indexed in various databases (Appendix B). A combination of subject headings and text words was adapted for each electronic resource: (arthritis OR “lupus erythematosus” OR pain OR fibromyalgia OR “benign joint hypermobility” OR “joint instability” OR “patellofemoral pain syndrome” OR “arthralgia” OR “limb pain” OR “synovitis” OR “JIA” OR “JRA” OR “JSLE” OR “joint swelling”) AND (child* OR infant* OR kid* OR toddler* OR adoles* OR teen* OR pubescen* OR puberty* OR p?ediatric) AND (screening OR “natural history” OR “incidence” OR “prevalence” OR “referral” OR diagnosis OR “predictive value of tests” OR “reproducibility of results” OR “sex factors” OR “age factors” OR anxiety OR comorbidity) AND (“ANA test” OR “FANA test” OR “antinuclear antibod*” OR “rheumatoid factor*” OR “cyclic citrulline peptide” OR “anticyclicitrullinated peptide” OR “anti-CCP”).

In addition to the searches of electronic databases, we searched the following conference proceedings and scientific meetings: American College of Rheumatology, Joint meeting of the British Society for Rheumatology, Canadian Rheumatology Association, European League Against Rheumatism, International League of Associations for Rheumatology, and American

Academy of Pediatrics from 2005 to 2010. Additionally, we searched the bibliographies of the included studies and reviews for relevant studies. We set up search alerts for PubMed and Web of Science to identify any new and potentially relevant studies during the course of the review.

Results from the literature searches were entered into a Thomson Reuters Reference Manager 11.0.1[®] bibliographic database.

Study Selection

A two-stage selection was carried out. For the initial broad screening stage, each article was screened by two independent reviewers who assessed the relevance of the study based on its title and abstract using prespecified broad screening criteria. We excluded articles if they were judged clearly as (1) not primary studies reporting on prevalence of conditions, diagnostic test accuracy, or clinical impact, (2) not ANA, RF, or CCP tests, or (3) did not include a pediatric population. Articles were rated as “include,” “exclude,” or “unsure.” The full text of studies rated as “include” or “unsure” by both reviewers were retrieved. Discrepancies in decisions between reviewers were resolved through discussion or third party adjudication, if needed.

For the second level of screening the full-text of each article was further examined by two independent reviewers using a standard inclusion/exclusion form (Appendix C). This form was based on a specific and comprehensive set of criteria (Table 1). Each reviewer rated the article as “include,” “exclude,” or “unsure.” There was no restriction on study design, language, or publication year. The minimal requirement for inclusion was that studies must have recruited a population comprised of children and adolescents aged 18 years or less, examined the appropriate tests and reference standards, and provided information on (1) sensitivity (Sn) and specificity (Sp) or (2) clinically important outcomes. Discrepancies between the reviewers were resolved through discussion or third party adjudication, if needed. The corresponding author of the article was contacted when additional information was needed for making the inclusion/exclusion decisions.

Table 1. Inclusion and exclusion criteria

Article Type	- Studies reporting original research - Any language - No restriction on publication year, except studies of ANA published before 1980 (excluded because detection methods using animal substrates are no longer considered valid)
Participants	- Studies providing separate data for a population comprising children (≤18 years) with undiagnosed MSK pain or joint swelling, a diagnosis of pSLE or JIA, or index test results (i.e., ANA, RF, CCP)
Study Design	- Studies of any design that included at least 2 participants
Index Tests	- ANA, RF, CCP - The assay method of ANA using animal substrate was excluded (pre-1980 methodology) - The test of hidden RF was excluded (proposed as an alternate to RF in JIA in 1970s but does not relate to conventional RF tests)
Reference Standard	- Diagnosis of pSLE or JIA based on clinical criteria
Outcomes	- For KQ 3 and 4: Studies providing sufficient data to calculate Sn and Sp - For KQ 5: Studies providing numerical data or a narrative description regarding referral decisions, additional tests ordered, clinical management, and patient and parent anxiety due to the clinical uncertainty or additional tests

ANA = antinuclear antibody; CCP = cyclic-citrullinated peptide; JIA = juvenile idiopathic arthritis; KQ = Key Question; MSK = musculoskeletal; RF = rheumatoid factor; pSLE = pediatric systemic lupus erythematosus; Sn = sensitivity; Sp = specificity

Data Extraction

Data were extracted by a single reviewer using a standard data extraction form and verified by a second reviewer. Any disagreements were resolved through discussion or third party adjudication, if needed. We extracted data for the following categories: study characteristics, participant characteristics, index test, reference standard, and outcomes.

Quality Assessment

We assessed the methodological quality of each study using the QUADAS checklist.⁷ The tool assesses important common biases in diagnostic studies including spectrum, incorporation, and verification biases (Appendix C). Spectrum bias occurs when included patients do not represent the intended spectrum of severity for the target condition (i.e., JIA or pSLE).⁷⁸ Incorporation bias occurs when the index test (i.e., ANA, RF, or CCP) is incorporated in the reference standard (e.g., ILAR or ACR criteria).⁷⁸ Partial verification bias occurs when a nonrandom set of patients does not undergo the reference standard.⁷⁸ Differential verification bias occurs when a set of patients is verified with a second reference standard, especially when this selection depends on the index test result.⁷⁸ Disease progression bias occurs when the patient's condition changes between administering the index test and the reference standard. Information bias occurs when the results of the index test are interpreted knowing the results of the reference standard, and vice versa.⁷⁸

Two reviewers performed quality assessment independently. Decision rules regarding application of QUADAS were developed a priori. Discrepancies were resolved through discussion or third party adjudication, as needed.

Data Analysis and Synthesis

We summarized the general characteristics of studies using descriptive statistics. For the diagnostic performance, we constructed 2x2 tables and calculated Sn, Sp, and positive and negative predictive values. For visual interpretation we presented the results in forest plots. An a priori decision was made to not conduct meta-analysis due to the expected large degree of heterogeneity in participant characteristics and test positive thresholds across studies. When data were available, subgroup analyses were conducted by accuracy modifiers, as well as by assay methods used and JIA onset-types.

Rating the Body of Evidence

The strength of evidence was graded for KQs 3 to 5 using the AHRQ system for grading the strength of evidence (AHRQ Guidance for the Evaluation of Medical Tests (draft)).⁸ We assessed four domains including risk of bias, consistency, directness, and precision, and developed a summary of overall strength of evidence. The grading was done by two independent reviewers, and any discrepancy was resolved by discussion or third party adjudication, as needed. The “risk of bias” domain was scored as low, medium, or high risk of bias corresponding to the results of QUADAS; the “consistency” domain was scored as consistent, inconsistent, or unknown based on the visual interpretation of the forest plots; “directness” was scored as direct or indirect based on the relevance of the evidence to the corresponding KQ; “precision” was scored as precise or imprecise based on the width of 95 percent confidence intervals. The overall summary rating was evaluated as high, moderate, low, or insufficient.

Applicability

Applicability refers to how generalizable the findings of this report are to a wider range of populations that vary by age, sex, clinical presentation, disease severity, and clinical setting. It was assessed according to the AHRQ Guidance for the Evaluation of Medical Tests (draft).⁸

Peer Review and Public Commentary

Six experts in pediatric medicine, pediatric rheumatology, rheumatology, pathology, and diagnostic testing reviewed the draft report and provided feedback. Reviewer comments were considered by the University of Alberta Evidence-based Practice Center in preparation of the final report. All peer reviewer comments and the disposition of comments were submitted to AHRQ for assessment and approval.

Results: Part One

The aim of Key Questions (KQs) 1 and 2 was to provide background information for the interpretation of the results of KQs 3, 4, and 5. Studies were selected based on the availability of the evidence and their representativeness to the pediatric population of North America.

Key Question 1.1. Incidence and Prevalence of Undiagnosed Musculoskeletal Complaints in Children

In studies of the epidemiology of musculoskeletal (MSK) pain in children, 60–85 percent of school-aged children reported at least one episode of MSK pain within a 3 month period.^{6,79} More girls reported pain (65 percent) than boys (55 percent).⁷⁹ Up to 30 percent of children and adolescents reported having experienced chronic pain (including MSK pain) which lasted for more than 6 months.^{2,80-83} No studies reported the prevalence of joint swelling in children.

The prevalence of MSK pain ranges from 2 percent in 12-year-olds to 52 percent in 18-year-olds.⁹⁻¹¹ The prevalence of MSK pain increases steadily with age throughout childhood and adolescence.^{12,13} Haraldstad, et al.,⁷⁹ examined 1,238 Norwegian schoolchildren aged 8 to 18 years and found that the prevalence of back pain increased with age for both sexes.⁷⁹ A 4-year prospective study reported that the incidence of new-onset low back pain doubled with age from 13 percent in 12-year olds to 24 percent in 15-year-olds.⁸⁰ While some studies reported that low back, neck, shoulder, leg, and chronic pain were more prevalent in girls than boys,^{79,80,84} two studies found no such difference,^{85,86} and one found male predominance.¹³

Key Question 1.2: The Positivity of Antinuclear Antibody, Rheumatoid Factor, and Cyclic-Citrullinated Peptide Tests in Healthy Children

Nine studies examined the prevalence of a positive ANA in a total of 1,413 healthy children and found a range of 0 to 18 percent¹⁴⁻²² All of these studies examined children and adolescents 18 years old or less except Baig and Shere,¹⁸ which included 1- to 19-year olds and Youngchiyud, et al.,²¹ which included 12- to 20-year olds. The definition of a positive cutoff of ANA titer ranged from 1:5 to 1:40.

Studies on the positivity of the RF and CCP tests in healthy children were scarce. A commentary⁸⁷ indicated that most children with a positive RF test did not have JIA,^{36,88} and most children with JIA did not have a positive RF.⁸⁹ Kasapcopur, et al.,²³ investigated 118 healthy Turkish children, using a cutoff of >25 IU/ml, and found a rate of 3 percent RF positivity.²³ The prevalence of CCP positivity in healthy children was 0 and 0.6 percent as reported in two studies.^{24,25}

Key Question 2.1. Noninflammatory Causes of Pediatric Musculoskeletal Pain

De Inocencio¹² conducted a retrospective chart review on 317 children between 3 and 15 years of age in a primary care setting in Madrid.¹² Noninflammatory etiologies accounted for the overwhelming majority (96.7 percent) of the MSK pain for children seen in primary care. He found that physical trauma (43.6 percent) was the most common cause of pediatric MSK pain,

and bone and muscle contusions were the most common trauma subgroup. Other noninflammatory etiologies included overuse (24.0 percent), osteochondroses (10.3 percent), hypermobility (3.3 percent), non-specific pain (7.6 percent), growing pain (3.5 percent), and viral infection (4.5 percent). Non-specific pain, growing pains, and hypermobility were much more common in preschool children (3 to 5 years) than in both school-aged children (6 to 9 years) and adolescents (10 to 14 years).

Key Question 2.2. Inflammatory Causes of Pediatric Musculoskeletal Pain

De Inocencio¹² reported that inflammatory etiologies accounted for a small fraction (3.3 percent) of primary care visits from children with MSK pain, and included toxic synovitis (2.5 percent) and inflammatory arthritis (0.8 percent). This was consistent with an earlier study by McGhee, et al.,⁵⁹ who conducted a retrospective chart review of 226 children with MSK pain who were referred for an initial rheumatology consultation. Among a group of 111 patients whose only presenting complaint was MSK pain, one child (1 percent) had a rheumatic disease (ankylosing spondylitis), and none had pSLE or JIA. The majority (81 percent) had mechanical MSK or overuse syndromes as explanations of their pain. In addition, the same study observed that among the 76 children diagnosed with JIA, only 12 (16 percent) included pain as part of their main complaints. Earlier observations by Sherry, et al.,⁹⁰ stated that 14 percent of patients with a confirmed diagnosis of JIA reported no pain. Although the numbers differ, both studies confirm that the absence of MSK pain does not rule out a diagnosis of JIA.

Key Question 2.3. The Resolution or Recurrence of Pediatric Musculoskeletal Pain

The recurrence of MSK pain was common in children. Thirty-five percent of 14-year olds who previously complained of low back pain reported recurrent episodes at age 18 and into early adulthood.⁹¹ Mikkelsen, et al.,⁹² examined 1,628 school-aged children with weekly pain at baseline and found that 52 percent reported MSK pain at 1-year followup.

Results: Part Two

Literature Search

The search strategies identified 11,994 citations from electronic databases and hand searching (Figure 2). Initial broad screening identified 496 potentially relevant citations. Of these, we included one retrospective cohort study and 27 case-control studies; all 28 studies addressed KQ 3 (diagnostic performance). No study provided subgroup data to address KQ 4 (diagnostic modifiers). No study addressed KQ 5 (test impacts). See Appendix D for a list of excluded studies.

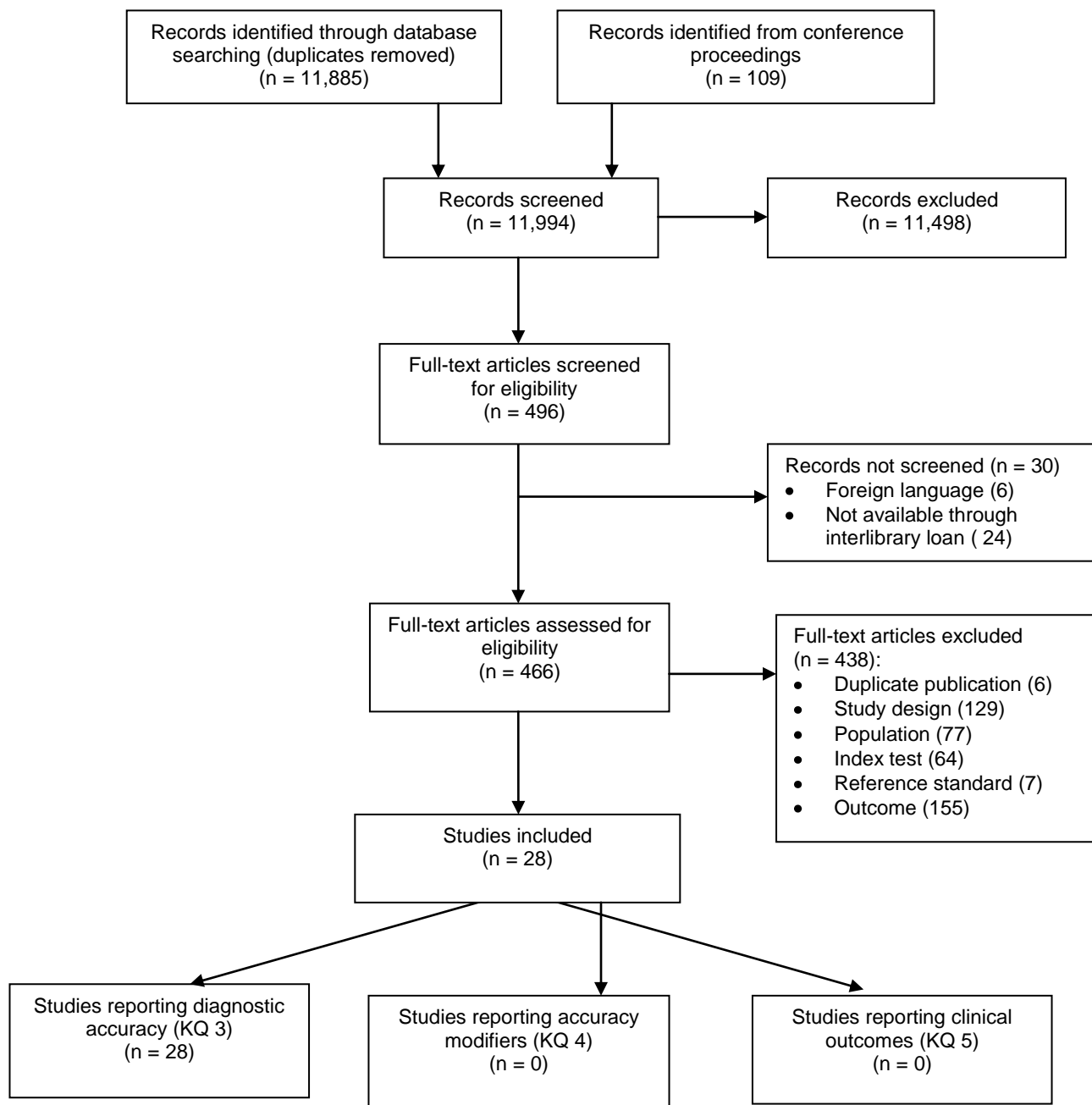
Characteristics of Included Studies

The 28 included studies were published between 1966 and 2009 (Appendix F). The mean age (where reported) of children ranged from 6.4 to 15 years of age. The studies were all published as full manuscripts in peer-reviewed journals. Thirteen studies^{24,30,31,33,37-39,41,43,44,46,48,49} were conducted in Europe, nine^{26,28,29,32,35,36,45,47,50} in North America, two^{40,42} in South America, three^{27,34,51} in Asia, and one²⁵ in Africa. Nine studies^{26,28,32,37,39,45,46,48,50} received funding from government, four^{24,30,34,51} from academic institutions, and four^{27,31,41,47} from noncommercial institutions. The remaining 11 studies^{25,29,33,35,36,38,40,42-44,49} did not report the source of funding.

One study³⁶ used a retrospective cohort design and included the spectrum of children with diagnosed and undiagnosed MSK pain. The remaining 27 studies used a case-control type design. The ANA test was examined in nine studies (two^{26,27} for pSLE and eight^{26,28-34} for JIA). The RF test was examined in 17 studies (one³⁵ for pSLE and 16^{28,30,33,35-47} for JIA). The CCP test was examined in seven studies^{24,25,30,48-51} for JIA. There were no studies that examined the CCP test for pSLE.

All of the studies included patients with MSK pain; none specifically addressed children with joint swelling.

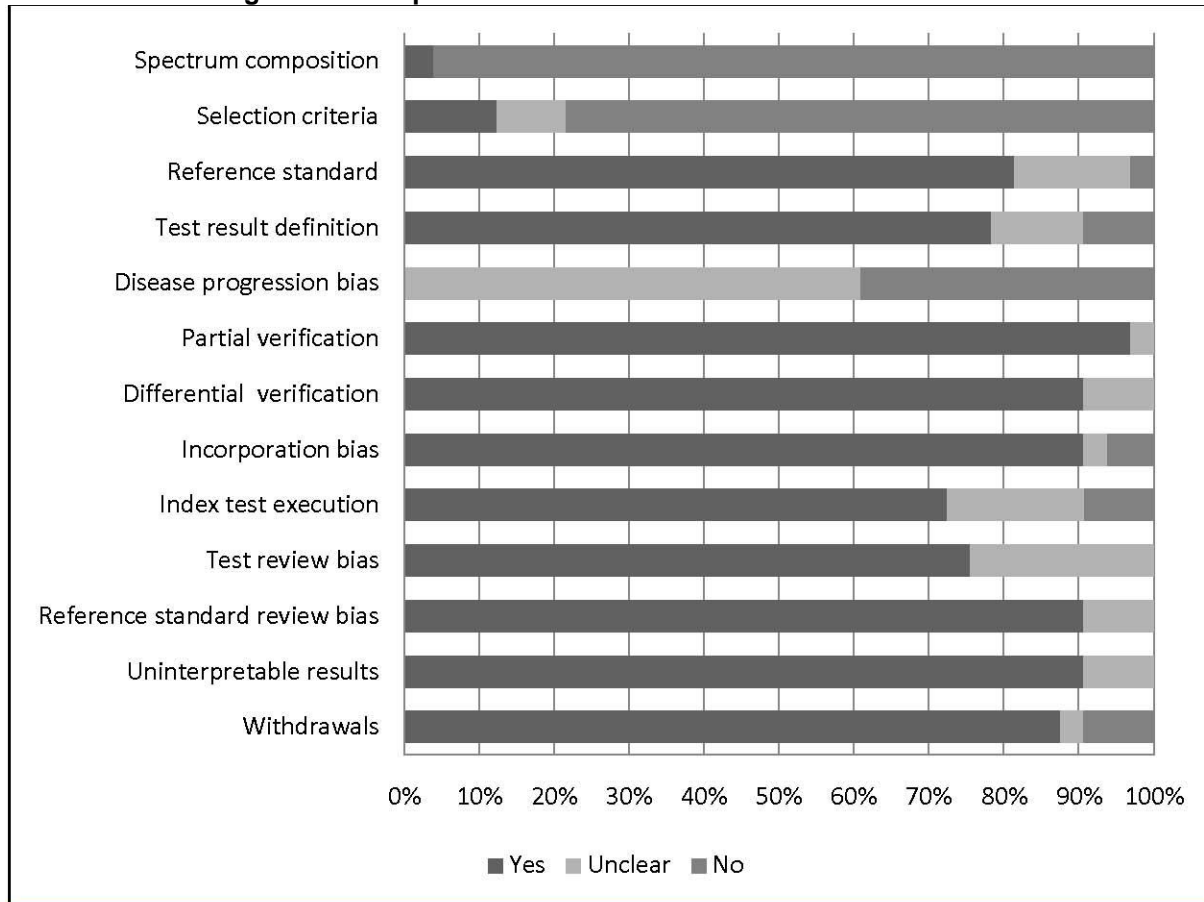
Figure 2. Results of literature search, retrieval, and selection



Methodological Quality of Included Studies

The methodological quality of the 28 included studies is summarized in Figure 3; summary tables are presented in Appendix E. Overall, there is substantial concern regarding spectrum bias. Most studies (97 percent) were rated “no” regarding the representativeness of the study population due to the case-control study design. The selection criteria of the population were not described adequately in most studies (83 percent). For studies examining ANA for pSLE, incorporation bias is a concern because ANA is considered one of the classification criteria for SLE.⁵⁵

Figure 3. Methodological quality of studies evaluating ANA, RF, and CCP tests for pSLE and JIA in children with undiagnosed MSK pain



ANA = antinuclear antibody test; CCP = cyclic-citrullinated peptide; JIA = juvenile idiopathic arthritis; MSK = musculoskeletal; pSLE = pediatric systemic lupus erythematosus; RF = rheumatoid factor

Key Question 3.1. Antinuclear Antibody Test for Pediatric Systemic Lupus Erythematosus

Study Characteristics

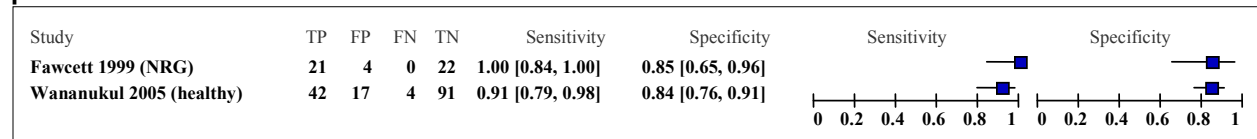
Two case-control studies^{26,27} including 201 children (67 pSLE, 134 controls) examined the prevalence of a positive ANA test in children with pSLE and control groups including healthy children and children scheduled for elective orthopedic surgery (Table 2). No information was provided in either study about the presence or absence of MSK pain or joint swelling in either the patient or the control groups.

The assay method of the index test in both studies was indirect immunofluorescence (IIF). Both studies used a positive cutoff titer of $\geq 1:40$.

Quantitative Results

The sensitivities (Sn) were 91 and 100 percent, the specificities (Sp) were 84 and 85 percent, the positive predictive values (PPV) were 71 and 84 percent, and negative predictive values (NPV) were 96 and 100 percent (Figure 4). Among control groups, the proportions of children who tested positive on the ANA test were 16 percent (healthy children) and 15 percent among patients scheduled for elective orthopedic surgeries.

Figure 4. Sensitivity and specificity of an ANA test for pSLE in children with undiagnosed MSK pain



ANA = antinuclear antibody test; FN = false negative; FP = false positive; MSK = musculoskeletal; NRG = nonrheumatic diseases; pSLE = pediatric systemic lupus erythematosus; TN = true negative; TP = true positive

Table 2. Description of studies evaluating an ANA test for pSLE in children with undiagnosed MSK pain

Author Year Location	Design	Funding	Source of Study Population	Control	Classification Criteria	Assay Method	Positive Threshold
Fawcett ²⁶ 1999	Case-control	Government	NR	Underwent elective orthopedic surgical procedures	ACR	IIF	Discernible fluorescence pattern at titer 1:40
Wananukul ²⁷ 2005	Case-control	Non-commercial institution	NR	Scheduled for elective surgery (adenotonsillectomy, herniorrhaphy or plastic surgery)	ACR	IIF	Titer \geq 1:40

ANA = antinuclear antibody test; ACR = American College of Rheumatologists; IIF = indirect immunofluorescence method; MSK = musculoskeletal; pSLE = pediatric systemic lupus erythematosus; NR = not reported

Key Question 3.2. Antinuclear Antibody Test for Juvenile Idiopathic Arthritis

Study Characteristics

Eight case-control studies^{26,28-34} including 1,382 children (1,067 JIA, 315 controls) examined the prevalence of a positive ANA test in children with JIA and control groups (Table 3). In four studies all participants were under 18 years of age; two studies^{28,34} included a small number of young adults. Two studies^{26,31} did not report age.

For the control groups, three studies³²⁻³⁴ included healthy children, five^{26,28-31} included children with nonrheumatic conditions, and one³² included children with other rheumatic diseases. Three studies^{29,31,34} recruited JIA patients from general hospitals or clinics, and the source was unreported by the remaining studies. For the classification of JIA, four studies^{26,28,32,34} used the ACR criteria, two^{30,31} used the ILAR criteria, one³³ used the EULAR criteria, and the classification criteria were unreported in one study.²⁹ Only one study (Nordal, et al.,³¹) looked at children with MSK pain who either had a diagnosis of JIA or acute lymphoblastic leukemia; none of the other studies commented on the presence or absence of MSK pain or joint swelling in either the patient or control groups. The frequency of uveitis or iridocyclitis was reported by three studies: 18 percent of JIA patients in Nordal, et al.,³¹ 6 percent of JIA patients in Osborn, et al.,³² and 3 percent in Wakhlu, et al.³⁴

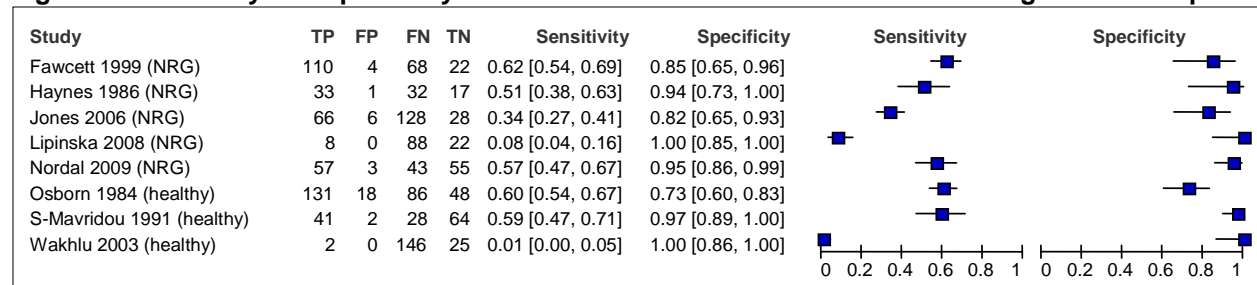
IIF was used in all studies except Jones, et al.,²⁹ who did not report the method used. In studies that reported the positive cutoff titers, the titers ranged from >1:20 to >1:320. Fawcett²⁶ and Nordal, et al.,³¹ examined multiple assay methods of ANA for JIA (Appendix F).

Quantitative Results

The Sn ranged from 1 to 62 percent (median = 54 percent); the Sp ranged from 73 to 100 percent (median = 95 percent); PPV ranged from 88 to 100 percent (median = 96 percent); and NPV ranged from 15 to 70 percent (median = 30 percent) (Figure 5). Analyses by subtypes of JIA are presented in Appendix G.

Among the healthy controls, the proportion of children (three groups³²⁻³⁴) who tested positive for ANA ranged from 0 to 27 percent (median = 3). Among controls with nonrheumatic conditions,^{26,28-31} the proportion of children with positive tests ranged from 0 to 18 percent (median = 6). There were two studies that reported very low Sn. Both had zero percent prevalence of a positive ANA among controls. In one,³⁰ this could be explained by their use of a high cutoff titer of >1:320. In the other study³⁴ the reason for the disparity is unclear.

Figure 5. Sensitivity and specificity of an ANA test for JIA in children with undiagnosed MSK pain



ANA = antinuclear antibody test; FN = false negative; FP = false positive; JIA = juvenile idiopathic arthritis; MSK = musculoskeletal; NRG = nonrheumatic disease group; TN = true negative; TP = true positive

Table 3. Description of studies evaluating an ANA test for JIA in children with undiagnosed MSK pain

Author Year Location	Design	Funding	Source of Study Population	Control	Classification Criteria	Assay Method	Positive Threshold
Fawcett ²⁶ 1999 U.S.	Case-control	Government	NR	NRG: Undergoing elective orthopedic surgical procedures	ACR	IIF	Positive if a clearly discernible fluorescence pattern appears at 1:40 serum dilution
Haynes ²⁸ 1986 U.S.	Case-control	Government and non-commercial institution	NR	NRG: Age-matched children with nonrheumatic diseases	ACR	IIF	Reading of $\geq 1+$ in fluorescence at 1:20 dilution
Jones ²⁹ 2006 North America	Case-control	NR	Randomly selected patients from multiple medical centers	NRG: Patients with acute lymphoblastic leukemia	NR	NR	Titer > 1:80
Lipinska ³⁰ 2008 Poland	Case-control	Academic institution	NR	NRG: Children with functional CV system dysfunction	ILAR	IIF	Titer > 1:320
Nordal ³¹ 2009 Norway	Case-control	Non-commercial institution	NR	NRG: Children undergoing elective outpatient procedures; no diagnosis of inflammatory diseases	ILAR	IIF	Titer > 1:80
Osborn ³² 1984 U.S.	Case-control	Government	Pediatric clinic	Healthy and ORG	ACR	IIF	Titer > 1:40
Siamopoulou-Mavridou ³³ 1991 Greece	Case-control	NR	NR	Healthy	EULAR	IIF	Titer > 1:40
Wakhiu ³⁴ 2003 India	Case-control	Academic institution	Immunology clinic	Healthy	ACR	IIF	Titer: > 1:40

ANA = antinuclear antibody test; ACR = American College of Rheumatology; CV = cardiovascular; EULAR = The European League Against Rheumatism; IIF = indirect immunofluorescence method; ILAR = International League of Associations for Rheumatology; JIA = juvenile idiopathic arthritis; MSK = musculoskeletal; NR = not reported; NRG = nonrheumatic disease group; ORG = other rheumatic diseases group

Key Question 3.3. Rheumatoid Factor Test for Pediatric Systemic Lupus Erythematosus

Study Characteristics

One case-control study by Hanson, et al.,³⁵ examined the prevalence of a positive IgM-RF test in 14 children with pSLE and 32 controls. The study was published in 1966. The assay method for the RF test was latex fixation. As the study was conducted prior to the development of the ACR criteria for the classification of SLE, patients were diagnosed based on criteria developed by Cook, et al.,⁹³ and Urbach.⁹⁴ The control groups comprised a mix of healthy children and children with other rheumatic conditions and ulcerative colitis. The study did not comment on the presence or absence of MSK pain or joint swelling in either the patients or controls.

Quantitative Results

The Sn was 29 percent and Sp was 88 percent.³⁵ The proportion of children who tested positive for RF was 13 percent.

Key Question 3.4. Rheumatoid Factor Test for Juvenile Idiopathic Arthritis

Study Characteristics

One retrospective cohort study by Eichenfield, et al.,³⁶ examined the records of pediatric patients who had a RF test and were seen in the walk in clinic, inpatient service, emergency department, or Pediatric Rheumatology Center at The Children's Hospital of Philadelphia (Table 4). Among the 437 patient records that were evaluated, 105 had a diagnosis of JIA according to ACR criteria. The remaining 332 patients (combined into one control group) had a mix of MSK complaints (n = 201) or symptoms suggestive of an underlying autoimmune disease (n = 131).

Fifteen case-control studies examined the prevalence of a positive RF in children with JIA and controls (Table 4). The diagnosis was based on the ACR criteria in nine studies,^{28,36,39-42,45-47} the EULAR criteria in four,^{33,37,38,44} and the ILAR criteria in one.³³ Two^{35,43} did not state which classification criteria were used. In one study, 62 percent of patients had chronic bilateral iridocyclitis.⁴⁶ For the control groups, nine studies^{33,37-41,44,46,47} used healthy children, three^{28,30,45} used children with nonrheumatic conditions, one⁴² used children with other rheumatic conditions, and two studies^{35,43} used a mixed group that included healthy children and children with nonrheumatic and other rheumatic conditions. None of the case-control studies commented on the presence or absence of MSK pain or joint swelling in the patient or control groups. Studies reporting data on non-IgM RF tests are presented in Appendix F.

Quantitative Results

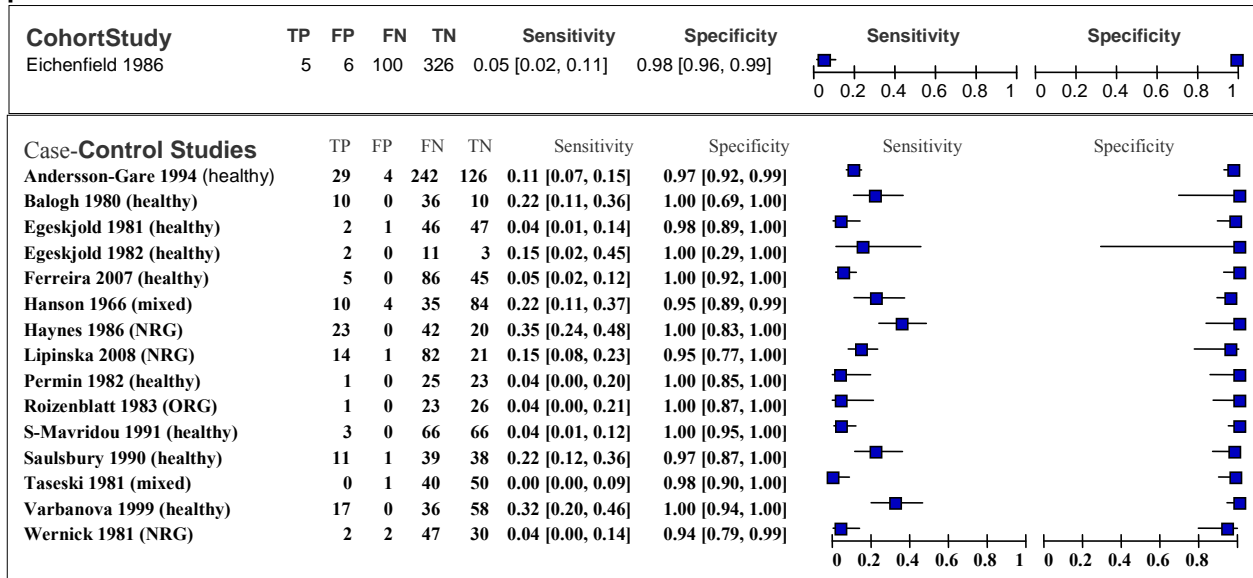
Results from the cohort study by Eichenfield, et al.,³⁶ show an Sn of 5 percent and a Sp of 98 percent (Figure 6). This is consistent with the RF results from the same center's total JIA population (6.9 percent) as reported in their paper. The authors also analyzed the post-test probability of JIA based on the reported Sn and Sp. The analyses take into account the pretest probability or prevalence of JIA, which in this cohort was 24 percent. The results showed that

probability of JIA increased to 45 percent with a positive test. They also showed that in a “typical” primary care practice, the probability of JIA went from 0.3 percent to 0.7 percent with a positive test.

Fifteen case-control studies, including 1,647 children (986 JIA, 661 controls) examined the prevalence of a positive IgM-RF test (Figure 6). Sn ranged from 0 to 35 percent (median = 11 percent), Sp ranged from 94 to 100 percent (median = 100 percent), PPV ranged from 0 to 100 percent (median = 100 percent), and NPV ranged from 20 to 71 percent (median = 48 percent).

The proportion of healthy children who tested positive ranged from 0 to 3 percent (median = 0). For the controls with nonrheumatic conditions, the proportion ranged from 0 to 6 percent (median = 5). For the controls (two groups) that included a mix of healthy children and children with other conditions, the proportion that tested positive ranged from 2 to 5 percent. There does not appear to be a relationship between the cutoff titer used (if reported) and Sn. Analyses by subtypes of JIA are presented in Appendix G.

Figure 6. Sensitivity and specificity of an RF (IgM) test for JIA in children with undiagnosed MSK pain



FN = false negative; FP = false positive; IgM = immunoglobulin M; JIA = juvenile idiopathic arthritis; MSK = musculoskeletal; NRG = nonrheumatic disease group; ORG = other rheumatic diseases group; RF = rheumatoid factor; TN = true negative; TP = true positive.

Table 4. Description of studies evaluating an RF (IgM) test for JIA in children with undiagnosed MSK pain

Author Year Location	Funding	Source of Study Population	Non-disease Group	Classification Criteria	Assay Method	Positive Threshold
Cohort Study						
Eichenfield ³⁶ 1986 U.S.	NR	Consecutive patients from pediatric hospital	NA (due to cohort design)	ACR	Latex fixation test	Titer > 1:80
Case-Control Studies						
Andersson-Gare ³⁷ 1994 Sweden	Government	Epidemiological survey in south-western Sweden	Healthy	EULAR	EIA	Mean of control group + 2 sd
Balogh ³⁸ 1980 Hungary	NR	Hospital consecutive patients	Healthy: Age- and sex-matched	EULAR/WHO workshop	Latex fixation test	NR
Egeskjold ³⁹ 1981 Denmark	Government	NR	Healthy: Age- and sex-matched	ACR	IIF	Titer > 1:9
Egeskjold ⁴⁶ 1982 Denmark	Government	NR	Healthy	ACR	IIF	Maximum of peak 2 displacement beyond normal range
Ferreira ⁴⁰ 2007 Brazil	NR	Randomly selected patients from multiple centers of pediatric rheumatology	Healthy	ACR	Latex fixation test	Latex: 20 IU/ml
Hanson ³⁵ 1966 U.S.	NR	NR	Healthy, NRG, ORG	Unclear	Latex fixation test	Titer > 1:160
Haynes ²⁸ 1986 U.S.	Government and non-commercial institution	NR	NRG: Age-matched children with nonrheumatic diseases	ACR	EIA	Mean of control + 2 sd
Lipinska ³⁰ 2008 Poland	Academic institution	NR	NRG: Age- and sex-matched children with functional cardiovascular system dysfunction	ILAR	EIA	24 RU/ml
Permin ⁴¹ 1982 Denmark	Non-commercial institution	NR	Healthy	ACR	IIF	Titer > 1:10
Roizenblatt ⁴² 1983 Brazil	NR	Pediatric clinic	ORG: Age- and sex-matched hypermobile children	ACR	EIA	Mean of control group + 2 sd

Table 4. Description of studies evaluating an RF (IgM) test for JIA in children with undiagnosed MSK pain (continued)

Author Year Location	Funding	Source of Study Population	Non-disease Group	Classification Criteria	Assay Method	Positive Threshold
Saulsbury ⁴⁷ 1990 U.S.	Non-commercial institution	NR	Healthy	ACR	EIA	Titer > 1:20
Siamopoulou-Mavridou ³³ 1991 Greece	NR	NR	Healthy: Age- and sex-matched children without rheumatic disease	EULAR	Latex fixation test	Mean optical density of healthy control + 3 sd
Taseski ⁴³ 1981 Yugoslavia	NR	NR	Healthy and ORG (collagen diseases)	NR	Latex slide test	Agglutination visually detectable
Varbanova ⁴⁴ 1999 Bulgaria	NR	NR	Healthy	EULAR	EIA	Mean IU of healthy control + 2 sd
Wernick ⁴⁵ 1981 U.S.	Government	NR	NRG (scoliosis and neurologic diseases)	ACR	Solid phase radioimmunoassay	Mean of normal control + 2 sd

ACR = American College of Rheumatology; EULAR = The European League Against Rheumatism; EIA = enzyme immunoassay; IgM = immunoglobulin M; IIF = indirect immunofluorescence; ILAR = International League of Associations for Rheumatology; IU = international unit; JIA = juvenile idiopathic arthritis; ml = milliliter; MSK = musculoskeletal; NA = not applicable; NR = not reported; NRG = nonrheumatic disease group; ORG = other rheumatic diseases group; RF = rheumatoid factor; RU = relative unit; sd= standard deviation; WHO = World Health Organization

Key Question 3.5. Cyclic-Citrullinated Peptide Test for Pediatric Systemic Lupus Erythematosus

No studies provided information to address this question.

Key Question 3.6. Cyclic-Citrullinated Peptide Test for Juvenile Idiopathic Arthritis

Study Characteristics

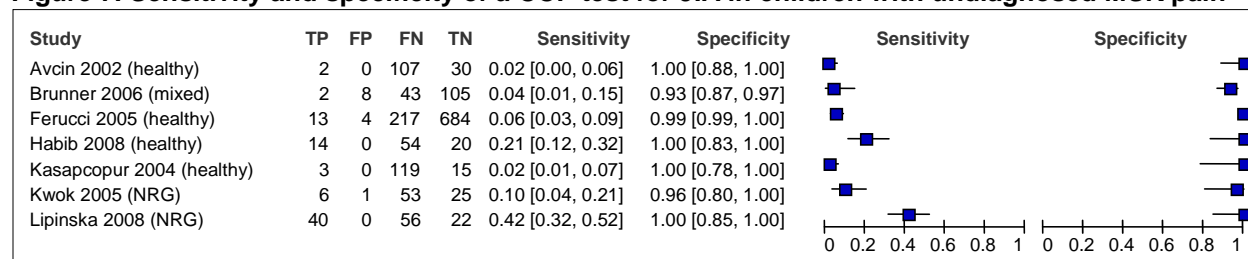
Seven case-control studies^{24,25,30,48-51} including 1,643 participants (729 JIA, 914 control) examined the prevalence of a positive CCP test in children with JIA and controls (Table 5). None of the studies reported on the presence or absence of MSK pain or joint swelling in either the patient or control groups.

The control group in four studies^{24,25,48,50} consisted of healthy children; two studies used a nonrheumatic patient control group (cardiovascular dysfunction,³⁰ allergies and idiopathic thrombocytopenia⁵¹). One study⁴⁹ used a mixed group of healthy children and children with other autoimmune diseases as their comparator. Four studies^{24,25,30,51} used the ILAR criteria for classification of JIA; one study⁵⁰ used ACR criteria. All seven studies used the EIA method; however, different cutoff points were used. It is not clear from the reported methods which anti-CCP assays were used in the studies.

Quantitative Results

The Sn ranged from 2 to 42 percent and Sp ranged from 93 to 100 percent (Figure 7). The PPV ranged from 20 to 100 percent and NPV ranged from 11 to 71 percent. The proportion of healthy controls (four groups) that tested positive for CCP ranged from 0 to 0.6 percent (median = 0). The proportion of controls with nonrheumatic conditions (two groups) that tested positive ranged from 0 to 4 percent (median = 2). Among the mixed controls (one group), 7 percent tested positive. Subgroup analyses by subtypes of JIA are presented in Appendix G.

Figure 7. Sensitivity and specificity of a CCP test for JIA in children with undiagnosed MSK pain



CCP = Cyclic-citrullinated peptide; FN = false negative; FP = false positive; JIA = juvenile idiopathic arthritis; MSK = musculoskeletal; NRG = nonrheumatic disease group; TN = true negative; TP = true positive

Table 5. Description of studies evaluating a CCP test for juvenile idiopathic arthritis in children with undiagnosed MSK pain

Author Year Location	Design	Funding	Source of Study Population	Control	Classification Criteria	Assay Method	Positive Threshold
Avcin 2002 ⁴⁸ Italy and Slovenia	Case-control	Government	NR	Healthy	NR	EIA	70 units
Brunner 2006 ⁴⁹ Germany	Case-control	NR	NR	Healthy, other autoimmune pathologies, NRG (undergoing cardiac therapy)	NR	EIA	2.5 RU
Ferucci 2005 ⁵⁰ U.S.	Case-control	Government	Cohort from Cincinnati and NIAMS registry	Healthy	ACR	EIA	5 units/ml
Habib 2008 ²⁵ Egypt	Case-control	NR	NR	Healthy	ILAR	EIA	20 units/ml
Lipinska 2008 ³⁰ Poland	Case-control	Academic institution	NR	NRG (functional cardiovascular system dysfunction)	ILAR	EIA	5 RU
Kasapcopur 2004 ²⁴ Turkey	Case-control	Academic institution	Consecutive patients admitted to hospital outpatient department	Healthy	ILAR	EIA	5 RU
Kwok 2005 ⁵¹ Hong Kong	Case-control	Academic institution	NR	NRG (allergy, idiopathic thrombocytopenia, and hepatitis C)	ILAR	EIA	20 units

ACR = American College of Rheumatology; CCP = cyclic-citrullinated peptide; EIA = enzyme immunoassay; ILAR = International League of Associations for Rheumatology; ml = milliliter; MSK = musculoskeletal; NR = not reported; RU = relative unit

Key Question 4.1. Accuracy Modifiers of Antinuclear Antibody, Rheumatoid Factor, Cyclic-Citrullinated Peptide Test for Pediatric Systemic Lupus Erythematosus

No studies provided data on accuracy modifiers (age, sex, race or ethnicity, comorbidities, recent infections) for any of the tests for pSLE.

Key Question 4.2. Accuracy Modifiers of Antinuclear Antibody, Rheumatoid Factor, Cyclic-Citrullinated Peptide Test for Juvenile Idiopathic Arthritis

No studies provided data on accuracy modifiers (age, sex, race or ethnicity, comorbidities, recent infections) for any of the tests for JIA.

Key Question 5. Clinical Impacts due to the Results of Antinuclear Antibody, Rheumatoid Factor, and Cyclic-Citrullinated Peptide Tests

No studies provided information to address this question.

Rating the Body of Evidence

The body of evidence was assessed using the AHRQ system for grading the strength of evidence (Table 6). All case-control studies were assessed as “high risk of bias” primarily due to spectrum bias and lack of adequate reporting of selection criteria. Therefore, strength of evidence derived from case-control studies was initially assessed as “low”. This assessment was downgraded to “insufficient” when other limitations were noted in any of the other domains. All studies were considered to provide “indirect” evidence due to the use of surrogate outcomes (i.e., test performance) instead of health outcomes.

Table 6. Strength of evidence for ANA, RF, and CCP tests for pSLE and JIA in children with undiagnosed MSK pain

Key Question	N Studies, (N disease; N control)	Outcome	Risk of Bias	Consistency	Direct-ness	Precision	Overall Strength of Evidence
ANA – pSLE	2 c-c (67; 134)	Sn	High	Consistent	Indirect	Precise	Insufficient
		Sp		Consistent	Indirect	Precise	Insufficient
ANA – JIA	8 c-c (1,067; 315)	Sn	High	Inconsistent	Indirect	Imprecise	Insufficient
		Sp		Consistent	Indirect	Precise	Insufficient
IgM-RF – pSLE	1 c-c (14; 32)	Sn	High	Unknown (single study)	Indirect	Imprecise	Insufficient
		Sp		Indirect	Imprecise	Insufficient	
IgM-RF – JIA	1 cohort (437)	Sn	Medium	Unknown (single study)	Indirect	Imprecise	Low
		Sp	Medium		Indirect	Imprecise	Low
	15 c-c (986; 661)	Sn	High	Inconsistent	Indirect	Imprecise	Insufficient
		Sp	High	Consistent	Indirect	Precise	Insufficient
CCP – pSLE	No studies	NA	NA	NA	NA	NA	Insufficient
CCP – JIA	7 c-c (729; 914)	Sn	High	Inconsistent	Indirect	Imprecise	Insufficient
		Sp		Consistent	Indirect	Precise	Insufficient
Accuracy modifiers	No studies	NA	NA	NA	NA	NA	Insufficient
Clinical impact of tests	No studies	NA	NA	NA	NA	NA	Insufficient

ANA = antinuclear antibody; c-c = case-control; CCP = cyclic-citrullinated peptide; IgM = immunoglobulin M; JIA = juvenile idiopathic arthritis; N = number; MSK = musculoskeletal; NA = not applicable; RF = rheumatoid factor; pSLE = pediatric systemic lupus erythematosus; Sn = sensitivity; Sp = specificity

Applicability

Applicability refers to how generalizable the findings of this report are to a wider range of populations that vary by age, sex, clinical presentation, disease severity, and clinical setting. The study populations were relatively heterogeneous in terms of the ethnicity, and a wide range of conditions were included. The age of the participants was similarly broad, as was the age of disease onset (generally between age 1 and 15 years when it was reported). The settings in which the tests were conducted also varied. Disease activity and severity were rarely described, but when reported more than half of the study participants had active disease. The diagnostic performance of the tests did not appear to differ significantly across the spectrum of patients and institutional sources, which would normally imply that the results are generalizable to a broader population.

Only one³⁶ of the 28 studies examined the performance of these tests in children with undiagnosed MSK pain—the primary population of interest for this review. In the study by Eichenfield, et al.,³⁶ even with a highly selected population of children who had undergone an RF test, only 4.8 percent of children with JIA had a positive RF, and 6 of 332 (1.8 percent) of the

remaining children—including 115 with only MSK pain—were RF positive. The authors conducted further analyses which demonstrated that in the primary care setting, the pretest probability of 0.3 percent only increased to 0.7 percent with a positive test.

None of the remaining studies reported on the presence or absence of MSK pain in either the patients or controls, and in all of these studies, the patients had known diagnoses of either JIA or pSLE. For this reason, the applicability of the tests to children with undiagnosed MSK pain is unclear. In the clinical setting of MSK pain, physicians are primarily interested in the degree to which the test results might change the probability that a patient with a positive or negative test may or may not have the disease (the positive predictive value [PPV] and negative predictive value [NPV], respectively). The PPV and NPV of these tests will change dramatically depending on the baseline prevalence (or pretest probability) of disease. Therefore, case-control studies that compare children with disease to a healthy population in a 1:1 ratio (a 50 percent disease prevalence) may overestimate test performance and utility. In contrast, in children with undiagnosed MSK pain seen in a primary care setting where prevalence of JIA or pSLE is less than one percent¹² the same test will identify more false positives than true positives. We developed a series of hypothetical scenarios to demonstrate this (Table 7).

Table 7. Hypothetical scenarios for PPV at different baseline disease prevalence

	PPV	NPV
ANA – pSLE (Sn = 91%, Sp = 84%)		
Baseline prevalence 1%	5%	100%
Baseline prevalence 50%	85%	90%
ANA – JIA (Sn = 36%, Sp = 96%)		
Baseline prevalence 1%	8%	99%
Baseline prevalence 50%	90%	60%
RF – JIA (Sn = 12%, Sp = 98%)		
Baseline prevalence 1%	6%	99%
Baseline prevalence 50%	86%	53%
CCP – JIA (Sn = 9%, Sp = 99%)		
Baseline prevalence 1%	8%	99%
Baseline prevalence 50%	90%	52%

ANA = antinuclear antibody; CCP = cyclic-citrullinated peptide; JIA = juvenile idiopathic arthritis; NPV = negative predictive value; PPV = positive predictive value; pSLE = pediatric systemic lupus erythematosus; RF = rheumatoid factor; Sn = sensitivity; Sp = specificity

Summary and Discussion

Summary

Incidence and Prevalence of Musculoskeletal Pain in Children and Adolescents

Studies that have investigated the prevalence of musculoskeletal (MSK) pain in children report a wide range of prevalence from 2 to 52 percent. Noninflammatory causes of MSK pain account for the majority of diagnoses (97 percent). No studies reported the prevalence of joint swelling in children.

Prevalence of Test Positivity in Healthy Children and Adolescents

From the studies included in our review, among the healthy control groups, the median antinuclear antibody (ANA) positivity is three percent, median RF positivity is zero percent, and CCP positivity is less than one percent.

Test Performance of ANA, RF, and CCP in Children and Adolescents With Undiagnosed MSK Pain

Only one retrospective cohort study examined the test performance of rheumatoid factor (RF) to diagnose juvenile idiopathic arthritis (JIA) among children with MSK pain. It demonstrated a sensitivity (Sn) of 5 percent and a specificity (Sp) of 98 percent. Fifteen case-control studies did not specifically address the test performance of RF among children with MSK pain. The strength of evidence is low for both Sn and Sp (Table 8). Further evidence is likely to change our confidence in the estimates of performance, and is likely to change the estimates.

The 12 case-control studies looking at other test-disease combinations did not specifically address the prevalence of positive tests for ANA or cyclic-citrullinated peptide (CCP) among children presenting with MSK pain. The strength of evidence is insufficient to determine the test performance of ANA or CCP to diagnose JIA or pediatric systemic lupus erythematosus (pSLE) in children with undiagnosed MSK pain (Table 9).

A general pattern of high Sp and low Sn was observed for almost all the test-disease combinations; however, the design of case-control studies may lead to bias.⁵²⁻⁵⁴ The selective inclusion of cases with established disease (i.e., JIA or pSLE) is likely to lead to an overestimation of Sn. The inclusion of healthy controls is expected to decrease the likelihood of false positive test results and lead to an overestimation of Sp.

Implications

There is insufficient evidence to determine the test performance of ANA or CCP in children with undiagnosed MSK pain. The strength of evidence is low for the utility of RF in the diagnosis of JIA in children with undiagnosed MSK pain. A result of high Sp and low Sn was observed for almost all the test-disease combinations. The generally low Sn suggests that it is inappropriate to use these tests in isolation (i.e., without clinical assessment) to make a diagnosis of JIA and pSLE. In spite of the high Sp, the low prevalence of JIA and pSLE in the target

population (i.e., children with undiagnosed MSK pain) makes the tests of limited diagnostic value. The presence of other clinical characteristics (e.g., morning stiffness, joint swelling, malar rash, cytopenia) may increase the pretest probability of the disease in question. While both the Sn and Sp for ANA for pSLE were high, this test in isolation has limited diagnostic value for children with undiagnosed MSK given the very low prevalence of pSLE, and up to 18 percent prevalence of false positive ANA in the general population.

Limitations

The generally insufficient strength of evidence is primarily attributable to the high risk of spectrum bias in the case-control studies, a result of the distinct disease and control groups not being representative of the target population of children with undiagnosed MSK pain. For studies examining ANA for pSLE, incorporation bias is a concern because ANA is considered one of the classification criteria for SLE.⁵⁵

There is no evidence with which to assess the impact of potential accuracy modifiers, and there is no evidence with which to assess the clinical utility of the tests including the impact of the test results on referrals, ordering of additional tests, patient management, and patient and parent anxiety levels.

In addition to the issues identified above, there are general limitations for systematic reviews such as publication bias. We addressed this issue by conducting a comprehensive search of the published literature for potentially relevant studies. Search strategies included combinations of subject headings and free text words. Even though we applied a diagnostic search filter to the search strategies of the electronic databases, our searches identified over 11,000 records. Furthermore, these searches were supplemented by hand searching for grey literature (i.e., unpublished or difficult to find studies). There is also a possibility of study selection bias. However, we employed at least two independent reviewers to identify potentially relevant studies, and feel confident that the studies that were excluded from this report were done so for consistent and appropriate reasons.

Conclusion

Most of the evidence from the 28 studies included in this review was not applicable to the population of interest as studies examined children with known disease rather than with undiagnosed MSK pain. No studies addressed children with joint swelling. No study provided a complete investigation on accuracy modifiers. No studies examined clinically important outcomes such as the impact of the test results on referrals, ordering of additional tests, patient management, and patient and parent anxiety levels.

Because the Sn and Sp of these tests have yet to be verified, current evidence does not support their use as diagnostic tests for children with undiagnosed MSK pain. They have a potential application as an adjunct to a clinical assessment that suggests the presence of an inflammatory arthritis or connective tissue disease.

Future Research

The following general recommendations for future research are based on the preceding discussion of the evidence.

- In order to better understand the natural history of MSK pain in children and the probability of a diagnosis of JIA or pSLE in this population, prospective cohort studies of children and adolescents with MSK pain are needed. Given the low prevalence of JIA and pSLE, a sufficiently large number of participants is required.
- For the research to be generalizable, researchers need to use consistent test methodology and cutoffs as well as consistent and well-accepted clinical criteria for the diagnoses of JIA and pSLE.
- Potential accuracy modifiers of test performance need to be examined, including age, sex, race, history of recent infections, presence of clinical characteristics other than MSK pain (e.g., morning stiffness, joint swelling, uveitis, malar rash, cytopenias).
- The clinical impact of these tests (e.g., referral decisions, additional tests ordered, clinical management, quality of life, psychological distress of child and/or parents) should be assessed in cohort studies.
- Efforts are needed to improve the overall quality of reporting of primary studies of diagnostic test accuracy. The STARD checklist includes 25 items that address the level of detail that should be specified within such studies including descriptions of participants, test methods, statistical methods, and results.⁵⁶ This could be considered as a guide for authors reporting studies that evaluate diagnostic tests.

Table 8. Summary of evidence of the diagnostic characteristics of ANA, RF, and CCP tests for pSLE and JIA in children with undiagnosed MSK pain

Key Questions	N Studies, Sample Size	Sensitivity Range (median)*	Specificity Range (median)	PPV Range (median)	NPV Range (median)	Strength of Evidence
KQ 3: Test performance						
3.1 ANA – pSLE	2 case-control, 201	91-100%	84-85%	71-84%	96-100%	Insufficient
3.2 ANA – JIA	8 case-control, 1,382	1-62% (54)	73-100% (95)	88-100% (96)	15-70% (30)	Insufficient
3.3 RF (IgM) – pSLE	1 case-control, 46	29%	88%	50%	74%	Insufficient
3.4 RF (IgM) – JIA	1 cohort study, 437	5%	98%	45%	77%	Low
	15 case-control, 1,647	0-35% (11)	94-100% (100)	0-100% (100)	20-71% (48)	Insufficient
3.5 CCP – pSLE	No studies					Insufficient
3.6 CCP – JIA	7 case-control, 1,643	2-42% (6)	93-100% (100)	20-100% (100)	11-71% (28)	Insufficient
KQ 4: Accuracy modifiers	No studies	NA	NA	NA	NA	Insufficient
KQ 5: Clinical impacts	No studies	NA	NA	NA	NA	Insufficient

*Median not presented if ≤ 4 studies.

ANA = antinuclear antibody; CCP = cyclic-citrullinated peptide; IgM = immunoglobulin M; JIA = juvenile idiopathic arthritis; KQ = Key Question; NA = not applicable; NPV = negative predictive value; PPV = positive predictive value; N = number; pSLE = pediatric systemic lupus erythematosus; RF = rheumatoid factor

References and Included Studies

1. Mikkelsen M, Salminen JJ, Kautiainen H. Non-specific musculoskeletal pain in preadolescents. Prevalence and 1-year persistence. *Pain*. 1997;73:29-35.
2. Palermo TM. Impact of recurrent and Chronic pain on child and family daily functioning: a critical review of the literature. *J Dev Behav Pediatr*. 2000;21(1):58-69.
3. Junnila J, Cartwright VW. Chronic musculoskeletal pain in children: Part II. Rheumatic causes. *Am Fam Physician*. 2006;74(2):293-300.
4. Malleson PN, Beauchamp RD. Rheumatology: 16. Diagnosing musculoskeletal pain in children. *CMAJ*. 2001;165(2):183-8.
5. Junnila J, Cartwright VW. Chronic musculoskeletal pain in children: Part I. Initial Evaluation. *Am Fam Physician*. 2006;74(1):115-22.
6. Clinch J, Eccleston C. Chronic musculoskeletal pain in children: assessment and management. *Rheumatology (Oxford)*. 2009;48:466-74.
7. Whiting J, Rutjes AW, Dinnes J, et al. Development and validation of methods for assessing the quality of diagnostic accuracy studies. *Health Technol Assess*. 2004;8(25):1-234.
8. Agency for Healthcare Research and Quality. AHRQ guidance for the evaluation of medical tests [draft]. Rockville, MD: Agency for Healthcare Research and Quality; 2010.
9. Goodman JE, McGrath PJ. The epidemiology of pain in children and adolescents: a review. *Pain*. 1991;46:247-64.
10. Abu-Arafeh I, Russell G. Recurrent limb pain in school. *Arch Dis Child*. 1996;74:336-9.
11. Kristjansdottir G. Prevalence of pain combinations and overall pain: a study of headache, stomach pain and back pain among school children. *Scand J Soc Med*. 1997;25:58-63.
12. de Inocencio J. Epidemiology of musculoskeletal pain in primary care. *Arch Dis Child*. 2004;89:431-4.
13. Burton AK, Clarke RD, McClune TD, et al. The natural history of low back pain in adolescents. *Spine*. 1996;21(20):2323-8.
14. Wananukul S, Voramethkul W, Kaewopas Y, et al. Prevalence of positive antinuclear antibodies in healthy children. *Asian Pac J Allergy Immunol*. 2005;23:153-7.
15. Petty RE, Cassidy JT, Sullivan DB. Clinical correlates of antinuclear antibodies in juvenile rheumatoid arthritis. *J Pediatr*. 1973;83:386-9.
16. Goel KM, Shanks RA, Whaley K, et al. Autoantibodies in childhood connective tissue diseases and in normal children. *Arch Dis Child*. 1975;50:419-23.
17. Arroyave CM, Mary JG, Kent CR, et al. The frequency of antinuclear antibody (ANA) in children by use of mouse kidney (MK) and human epithelial cells (HEp-2) as substrates. *J Allergy Clin Immunol*. 1988;82:741-4.
18. Baig MM, Shere SJ. Prevalence of autoantibodies in Saudi population. *J Med*. 1989;286-90.
19. Martini A, Lorini R, Zanaboni D, et al. Frequency of autoantibodies in normal children. *Am J Dis Child*. 1989;143:493-6.
20. Cabral DA, Petty RE, Fung M, et al. Persistent antinuclear antibodies in children without identifiable inflammatory rheumatic or autoimmune disease. *Pediatrics*. 1992;89:441-4.
21. Youngchaiyud U, Youngchaiyud P, Chandanyingyong D, et al. Incidence of autoantibodies in Thais. *J Med Assoc Thai*. 1981;64:445-8.

22. Allen RC, Dewez P, Stuart L, et al. Antinuclear antibodies using HEP-2 cells in normal children and in children with common infections. *J Paediatr.* 1991;27:39-42.
23. Kasapcopur O, Ozbakir F, Arisoy N, et al. Frequency of antinuclear antibodies and rheumatoid factor in healthy Turkish children. *Turk J Pediatr.* 1999;41(1):67-71.
24. Kasapcopur O, Altun S, Aslan M, et al. Diagnostic accuracy of anti-cyclic citrullinated peptide antibodies in juvenile idiopathic arthritis. *Ann Rheum Dis.* 2004;63(12):1687-9.
25. Habib HM, Mosaad YM, Youssef HM. Anti-cyclic citrullinated peptide antibodies in patients with juvenile idiopathic arthritis. *Immunol Invest.* 2008;37(8):849-57.
26. Fawcett PT, Rose CD, Gibney KM, et al. Use of ELISA to measure antinuclear antibodies in children with juvenile rheumatoid arthritis. *J Rheumatol.* 1999;26(8):1822-6.
27. Wananukul S, Voramethkul W, Kaewopas Y, et al. Prevalence of positive antinuclear antibodies in healthy children. *Asian Pac J Allergy Immunol.* 2005;23(2-3):153-7.
28. Haynes DC, Gershwin ME, Robbins DL. Autoantibody profiles in juvenile arthritis. *J Rheumatol.* 1986;13(2):358-63.
29. Jones O, Spencer C, Bowyer S, et al. A multicenter case-control study on predictive factors distinguishing childhood leukemia from juvenile rheumatoid arthritis. *Pediatrics.* 2006;117(5):e840-e844.
30. Lipinska J, Smolewska E, Brozik H, et al. Anti-CCP antibodies in children with Juvenile Idiopathic Arthritis (JIA) - diagnostic and clinical significance. *Cent Eur J Immunol.* 2008;33(1):19-23.
31. Nordal EB, Songstad NT, Berntson L, et al. Biomarkers of chronic uveitis in juvenile idiopathic arthritis: predictive value of antihistone antibodies and antinuclear antibodies. *J Rheumatol.* 2009;36(8):1737-43.
32. Osborn TG, Patel NJ, Moore TI, et al. Use of the HEP-2 cell substrate in the detection of antinuclear antibodies in juvenile rheumatoid arthritis. *Arthritis Rheum.* 1984;27(11):1286-9.
33. Siamopoulou-Mavridou A, Mavridis AK, Terzoglou C, et al. Autoantibodies in Greek juvenile chronic arthritis patients. *Clin Exp Rheumatol.* 1991;9(6):647-52.
34. Wakhlu A, Gupta D, Aggarwal A, et al. Low levels of anti-histone antibodies in north Indian children with juvenile rheumatoid arthritis. *Indian J Med Res.* 2003;118:204-7.
35. Hanson V, Kornreich HK, Drexler E. Rheumatoid factor in children with lupus erythematosus. A serologic study. *Am J Dis Child.* 1966;112(1):28-32.
36. Eichenfield AH, Athreya BH, Doughty RA, et al. Utility of rheumatoid factor in the diagnosis of juvenile rheumatoid arthritis. *Pediatrics.* 1986;78(3):480-4.
37. Andersson GB, Fasth A. Serum concentration of hyaluronan, IgM and IgA rheumatoid factors in a population based study of juvenile chronic arthritis. *Scand J Rheumatol.* 1994;23(4):183-90.
38. Balogh Z, Meretey K, Falus A, et al. Serological abnormalities in juvenile chronic arthritis: a review of 46 cases. *Ann Rheum Dis.* 1980;39(2):129-34.
39. Egeskjold EM, Permin H, Horbov S, et al. Anti-IgG antibodies in juvenile rheumatoid arthritis. *Acta Paediatr Scand.* 1981;70(5):711-6.
40. Ferreira RA, Silva CHM, Silva DAO, et al. Is measurement of IgM and IgA rheumatoid factors (RF) in juvenile rheumatoid arthritis clinically useful? *Rheumatol Int.* 2007;27(4):345-9.
41. Permin H, Egeskjold EM. IgE anti-IgG antibodies in patients with juvenile and adult rheumatoid arthritis including Felty's syndrome. *Allergy.* 1982;37(6):421-7.
42. Roizenblatt S, Goldenberg J, Gabriel A, Jr., et al. 7S IgG rheumatoid factor and hidden 19S IgM rheumatoid factor in juvenile chronic arthritis. *Allergol Immunopathol.* 1993;21(5):197-200.

43. Taseski B, Dezelic N, Dezelic G, et al. A study of rheumatoid factors in juvenile rheumatoid arthritis (chronic polyarthritis) by the photometric latex test. *Z Rheumatol.* 1981;40(1):17-20.
44. Varbanova BB, Baleva M, Nikolov K, et al. Prevalence of IgM-, IgA- and IgG-rheumatoid factors in seronegative polyarticular disease compared to pauciarticular disease in juvenile chronic arthritis as measured by ELISA. *Adv Exp Med Biol.* 1999;455:61-8.
45. Wernick R, LoSpalluto JJ, Fink CW, et al. Serum IgG and IgM rheumatoid factors by solid phase radioimmunoassay. A comparison between adult and juvenile rheumatoid arthritis. *Arthritis Rheum.* 1981;24(12):1501-11.
46. Egeskjold EM, Johansen A, Permin H, et al. The significance of antinuclear antibodies in juvenile rheumatoid arthritis associated with chronic bilateral iridocyclitis. *Acta Paediatr Scand.* 1982;71(4):615-20.
47. Saulsbury FT. Prevalence of IgM, IgA and IgG rheumatoid factors in juvenile rheumatoid arthritis. *Clin Exp Rheumatol.* 1990;8(5):513-7.
48. Avcin T, Cimaz R, Falcini F, et al. Prevalence and clinical significance of anti-cyclic citrullinated peptide antibodies in juvenile idiopathic arthritis. *Ann Rheum Dis.* 2002;61(7):608-11.
49. Brunner JK, Sitzmann FC. Anticyclic citrullinated peptide antibodies in juvenile idiopathic arthritis. *Mod Rheumatol.* 2006;16(6):372-5.
50. Ferucci ED, Majka DS, Parrish LA, et al. Antibodies against cyclic citrullinated peptide are associated with HLA-DR4 in simplex and multiplex polyarticular-onset juvenile rheumatoid arthritis. *Arthritis Rheum.* 2005;52(1):239-46.
51. Kwok JSY, Hui KH, Lee TL, et al. Anticyclic citrullinated peptide: diagnostic and prognostic values in juvenile idiopathic arthritis and rheumatoid arthritis in a Chinese population. *Scand J Rheumatol.* 2005;34(5):359-66.
52. Lijmer JG, Mol BW, Heisterkamp S, et al. Empirical evidence of design-related bias in studies of diagnostic tests. *JAMA.* 1999;282:1061-6.
53. Whiting P, Rutjes AW, Reitsma JB, et al. Sources of variation and bias in studies of diagnostic accuracy: a systemic review. *Ann Intern Med.* 2004;140:189-202.
54. Pai M, Flores LL, Pai N, et al. Diagnostic accuracy of nucleic acid amplication tests for tuberculous meningitis: A systematic review and meta-analysis. *Lancet Infect Dis.* 2003;3:633-43.
55. Tan EM, Cohen AS, Fries JF, et al. The 1982 revised criteria for the classification of systemic lupus erythematosus. *Arthritis Rheum.* 1982;25:1271-7.
56. Bossuyt PM, Reitsma JB, Bruns DE, et al. The STARD statement for reporting studies of diagnostic accuracy: explanation and elaboration. *Ann Intern Med.* 2003;138(1):W1-W12.
57. Jacewicz M. Symptoms and diagnosis of musculoskeletal disorders. *MERCK*. 2006. www.merck.com/mmhe/sec05/ch059/ch059b.html
58. de Inocencio J. Musculoskeletal pain in primary pediatric care analysis of 1000 consecutive general pediatric clinic visits. *Pediatrics.* 1998;102(6):e63.
59. McGhee JL, Burks FN, Sheckels JL, et al. Identifying children with chronic arthritis based on chief complaints: absence of predictive value for musculoskeletal pain as an indicator of rheumatic disease in children. *Pediatrics.* 2002;110(2:Pt 1):9.
60. Petty RE, Laxer RM. *Textbook of pediatric rheumatology.* 5 ed. Philadelphia: Elsevier; 2005.
61. Malleson PN, Fung MY, Rosenburg AM. The incidence of pediatric rheumatic diseases: result from the Canadian Pediatric Rheumatology Association Disease Registry. *J Rheumatol.* 1996;23:1981-7.
62. Kamphuis S, Silverman ED. Prevalence and burden of pediatric-onset systemic lupus erythematosus. *Nat Rev Rheumatol.* 2010;6(9):538-46.

63. Hochberg MC. Updating the American College of Rheumatology revised criteria for the classification of systemic lupus erythematosus [letter]. *Arthritis Rheum.* 1997;40:1725.
64. Benseler SM, Silverman ED. Systemic lupus erythematosus. *Pediatr Clin N Amer.* 2005;52:443.
65. Sahwney S, Woo P. Diagnosis and management of juvenile idiopathic arthritis: current status. *Indian Pediatr.* 2001;38:1083-90.
66. Hayward K, Wallace CA. Recent developments in anti-rheumatic drugs in pediatrics: treatment of juvenile idiopathic arthritis. *Arthritis Res Ther.* 2009;11(1):216-27.
67. Petty RE, Southwood TR, Manners P, et al. International League of Associations for Rheumatology Classification of Juvenile Idiopathic Arthritis: Second Revision, Edmonton, 2001. *J Rheumatol.* 2004;31(2):390-2.
68. Brewer EJ, Bass J, Baum J, et al. Current proposal revision of JRA criteria. *Arthritis Rheum.* 1977;20 (Suppl. 2):189-90.
69. EULAR. European League Against Rheumatism Bulletin No. 4: nomenclature and classification of arthritis in children. 1977.
70. Fabien N, Olsson NO, Goetz J, et al. Prevalence of autoantibodies to cyclic citrullinated peptide in patients with rheumatic diseases other than rheumatoid arthritis: a French multicentre study. *Clinic Rev Allerg Immunol.* 2008;34:34-44.
71. Niewold TB, Harrison MJ, Paget SA. Anti-CCP antibody testing as a diagnostic and prognostic tool in rheumatoid arthritis. *Q J Med.* 2007;100:193-201.
72. American College of Rheumatology. Position statement on methodology of testing for antinuclear antibodies. American College of Rheumatology. 2009. [www.rheumatology.org/practice/clinical/position/ana_position_stmt.pdf#search="methodologyoftestingforantinuclear"](http://www.rheumatology.org/practice/clinical/position/ana_position_stmt.pdf#search=). Accessed February 3, 2011.
73. Vaile JH, Dyke L, Kherani R, et al. Is high titre ANA specific for connective tissue disease? *Clin Exp Rheumatol.* 2000;18(4):433-8.
74. Keren DF. Antinuclear antibody testing. *Clin Lab Med.* 2002;22(2):447-74.
75. Syed RH, Gilliam BE, Moore TL. Rheumatoid factors and anticyclic citrullinated peptide antibodies in pediatric rheumatology. *Curr Rheumatol Rep.* 2008;10:156-63.
76. Lee AN, Beck CE, Hall HM. Rheumatoid factor and anti-CCP autoantibodies in rheumatoid arthritis: a review. *Clin Lab Sci.* 2008;21:15-8.
77. Masson-Bessiere C, Sebbag M, Girbal-Neuhauser E, et al. The major synovial targets of the rheumatoid arthritis-specific antifilaggrin autoantibodies are deiminated forms of the alpha- and beta-chains of fibrin. *J Immunol.* 2001;166(6):4177-84.
78. Leeflang MMG, Deeks JJ, Gatsonis C, et al. Systematic reviews of diagnostic test accuracy. *Ann Intern Med.* 2008;149(12):889-97.
79. Haraldstad K, Sorum R, Eide H, et al. Pain in children and adolescents: prevalence, impact on daily life, and parents' perception, a school survey. *Scand J Caring Sci.* 2010;doi: 10.1111/j.1471-6712.2010.00785.x.
80. Perquin CW, Hazebroek-Kampschreur AA, Hunfeld JA, et al. Pain in children and adolescents: a common experience. *Pain.* 2000;87:51-8.
81. Roth-Isigkeit A, Thyen U, Raspe HH, et al. Reports of pain among German children and adolescents: an epidemiological study. *Acta Paediatr.* 2004;93:258-63.
82. Brattberg G. Do pain problems in young school children persist into early adulthood? A 13-year follow-up. *Eur J Pain.* 2004;8:187-99.
83. Manchikanti L, Singh V, Datta S, et al. Comprehensive review of epidemiology, scope, and impact of spinal pain. *Pain Physician.* 2009;12:E35-E70.

84. Diepenmaat AC, van der Wal MF, de Vet HC, et al. Neck/shoulder, low back, and arm pain in relation to computer use, physical activity, stress, and depression among Dutch adolescents. *Pediatrics*. 2006;117(2):412-6.
85. Olsen TL, Anderson RL, Dearwater SR, et al. The epidemiology of low back pain in an adolescent population. *Am J Public Health*. 1992;82:606-8.
86. Sundblad GMB, Saartok T, Engstrom LM. Prevalence and co-occurrence of self-rated pain and perceived health in school-children: age and gender differences. *Eur J Pain*. 2007;11(2):171-80.
87. Jarvis JN. Commentary - ordering lab tests for suspected rheumatic disease. *Pediatr Rheumatol*. 2008;6(19):1-3.
88. Jarvis JN. The unique clinical presentation of children with chronic arthritis: putting the pediatrics in pediatric rheumatology. *Curr Prob Pediatr Adolesc Health Care*. 2006;36:80-2.
89. Cuesta I, Kerr KL, Jarvis JN. Subspecialty referrals for pauciarticular juvenile rheumatoid arthritis. *Arch Pediatr Adolesc Med*. 2000;154(2):122-5.
90. Sherry DD, Bohnsack J, Salmonson K, et al. Painless juvenile rheumatoid arthritis. *J Pediatr*. 1990;116(6):921-3.
91. Salminen JJ, Erkintalo MO, Pentti J, et al. Recurrent low back pain and early disc degeneration in the young. *Spine*. 1999;13:1316-21.
92. Mikkelsen M. Musculoskeletal pain and fibromyalgia in preadolescents: prospective 1-year follow-up study. *Turun yliopisto Sarja D Osa*. 1998;320.
93. Cook CD, Wedgwood RJP, Craig JM, et al. Systemic lupus erythematosus. Description of 37 cases in children and a discussion of endocrine therapy in 32 of the cases. *Pediatrics*. 1960;26:570.
94. Urbach F. Lupus erythematosus. *Pediatr Clin N Amer*. 1961;8:873.

Acronyms

ACR	American College of Rheumatology
AHRQ	Agency for Healthcare Research and Quality
ANA	Antinuclear antibody
ARA	American Rheumatism Association
CCP	Cyclic-citrullinated peptide
CDSR	Cochrane Database of Systematic Reviews
CINAHL	Cumulative Index to Nursing and Allied Health Literature
ELISA	Enzyme linked immunosorbent assay
EPC	Evidence-based Practice Center
EULAR	European League Against Rheumatism
FANA	Fluorescent antinuclear antibody test
FN	False negative
FP	False positive
Ig	Immunoglobulin
IIF	Indirect immunofluorescence
ILAR	International League of Associations for Rheumatology
IU	International unit
JIA	Juvenile idiopathic arthritis
JRA	Juvenile rheumatoid arthritis
KQ	Key Question
LST	Latex slide test
MSK	Musculoskeletal
NIAMSK	National Institute of Arthritis and Musculoskeletal and Skin Diseases
NPV	Negative predictive value
NRG	Nonrheumatic disease group
OD	Optical density
ORG	Other rheumatic diseases group
PPV	Positive predictive value
pSLE	Pediatric systemic lupus erythematosus
RF	Rheumatoid factor
SD	Standard deviation
Sn	Sensitivity
Sp	Specificity
SSC	Standard sensitized test
TN	True negative
TP	True positive
WHO	World Health Organization

Appendix A. American College of Rheumatology Criteria for Classification of Systemic Lupus Erythematosus

1997 Update of the 1982 American College of Rheumatology revised criteria for classification of systemic lupus erythematosus

1. Malar Rash	Fixed erythema, flat or raised, over the malar eminences, tending to spare the nasolabial folds
2. Discoid rash	Erythematous raised patches with adherent keratotic scaling and follicular plugging; atrophic scarring may occur in older lesions
3. Photosensitivity	Skin rash as a result of unusual reaction to sunlight, by patient history or physician observation
4. Oral ulcers	Oral or nasopharyngeal ulceration, usually painless, observed by physician
5. Nonerosive arthritis	Involving 2 or more peripheral joints, characterized by tenderness, swelling, or effusion
6. Pleuritis or pericarditis	1. Pleuritis--convincing history of pleuritic pain or rubbing heard by a physician or evidence of pleural effusion OR 2. Pericarditis--documented by electrocardiogram or rub or evidence of pericardial effusion
7. Renal disorder	1. Persistent proteinuria > 0.5 grams per day or > than 3+ if quantitation not performed OR 2. Cellular casts--may be red cell, hemoglobin, granular, tubular, or mixed
8. Neurologic disorder	1. Seizures--in the absence of offending drugs or known metabolic derangements; e.g., uremia, ketoacidosis, or electrolyte imbalance OR 2. Psychosis--in the absence of offending drugs or known metabolic derangements, e.g., uremia, ketoacidosis, or electrolyte imbalance
9. Hematologic disorder	1. Hemolytic anemia--with reticulocytosis OR 2. Leukopenia--< 4,000/mm ³ on ≥ 2 occasions OR 3. Lymphopenia--< 1,500/mm ³ on ≥ 2 occasions OR 4. Thrombocytopenia--<100,000/mm ³ in the absence of offending drugs
10. Immunologic disorder	1. Anti-DNA: antibody to native DNA in abnormal titer OR 2. Anti-Sm: presence of antibody to Sm nuclear antigen OR 3. Positive finding of antiphospholipid antibodies on: 1. an abnormal serum level of IgG or IgM anticardiolipin antibodies, 2. a positive test result for lupus anticoagulant using a standard method, or 3. a false-positive test result for at least 6 months confirmed by Treponema pallidum immobilization or fluorescent treponemal antibody absorption test
11. Positive antinuclear antibody	An abnormal titer of antinuclear antibody by immunofluorescence or an equivalent assay at any point in time and in the absence of drugs

The classification is based on 11 criteria. For the purpose of identifying patients in clinical studies, a person is defined as having SLE if any 4 or more of the 11 criteria are present, serially or simultaneously, during any interval of observation.

From: Hochberg MC. Updating the American College of Rheumatology revised criteria for the classification of systemic lupus erythematosus [letter]. Arthritis Rheum 1997;40:1725. Reprinted with permission from John Wiley and Sons.

Appendix B. Literature Search Strings

Title: ANA RF Anti CCP Testing Search Strategies

Database: MEDLINE , 1950 - present

Search name: ANA RF Anti CCP Testing -MEDLINE -

Notes: limits: human, 1960-2009

Date searched: Jan 21, 2010

Results: 5,389

<p>1. citrulline/ 2. exp Peptides, Cyclic/ 3. 1 and 2 4. ((anti adj ccp) or (citrullinated adj peptide*)).mp. 5. ((citrulline adj antibod*) or (anti-citrulline adj antibod*)).ti,ab. 6. exp Antibodies, Antinuclear/ 7. ((antinuclear adj antibod*) or (antinuclear adj factor*)).ti,ab. 8. (ana adj titer).ti,ab. 9. (ANA adj2 test*).ti,ab. 10. (FANA adj2 test*).ti,ab. 11. exp Rheumatoid Factor/ 12. (rheumatoid adj factor*).ti,ab. 13. or/3-12 14. exp Lupus Erythematosus, Systemic/ 15. (JSLE or SLE or "lupus erythematosus").ti,ab. 16. exp Pain/di, et 17. Growth/ph 18. (grow* and (pain or pains)).ti,ab. 19. 16 and (17 or 18) 20. musculoskeletal diseases/ or arm/ or leg/ or extremities/ 21. 16 and 20 22. Fibromyalgia/ 23. fibromyalgia.ti,ab. 24. exp arthralgia/ 25. arthralgia.ti,ab. 26. ((joint* adj pain*) or (limb* adj pain*)).ti,ab. 27. limp*.ti,ab. 28. benign.ti,ab. 29. exp Joint Instability/ 30. (joint adj (instability or hypermobility)).ti,ab. 31. 28 and (29 or 30) 32. Patellofemoral Pain Syndrome/ 33. (patellofemoral adj pain adj syndrome).ti,ab. 34. exp Synovitis/ or synovitis.mp. 71. exp demography/ 72. age factors/ or "age of onset"/ 73. sex factors/ 74. infection/ or infection*.ti,ab. 75. anxiety/ or (anxious* or anxiety).ti,ab. 76. comorbidity/ 77. or/71-76 78. exp Rheumatic Diseases/di, co, et, im, pa, pp 79. exp Connective Tissue Diseases/di, co, et, im, pa, pp 80. exp arthritis/di, co, et, im, pa, pp 81. arthritis, rheumatoid/di, co, et, im, pa, pp 82. arthritis, juvenile rheumatoid/di, co, et, im, pa, pp 83. exp Lupus Erythematosus, Systemic/di, co, et, im,</p>	<p>35. or/14-15,19-27,31-34 36. Arthritis/ 37. (\$arthritis or (\$articular adj arthritis)).ti,ab. 38. or/36-37 39. exp child/ or (adolesc* or early or juvenile).ti,ab. 40. (JIA or JRA).ti,ab. 41. or/39-40 42. 38 and 41 43. exp Arthritis, Juvenile Rheumatoid/ 44. ((juvenile or early) adj (rheumatoid or idiopathic) adj arthritis).ti,ab. 45. or/42-44 46. or/14-15,19,21-27,31-34,45 47. incidence/ 48. prevalence/ 49. exp disease progression/ 50. Natural History/ 51. natural history.ti,ab. 52. or/47-51 53. exp Mass Screening/ 54. exp "referral and consultation"/ 55. (screen* or refer*).ti,ab. 56. or/53-55 57. exp "Reproducibility of Results"/ 58. exp "Sensitivity and Specificity"/ 59. Predictive Value of Tests/ 60. (di or bl or cl or im).fs. 61. exp Diagnostic Errors/ 62. early diagnosis/ 63. exp delayed diagnosis/ 64. Diagnosis, Differential/ 65. or/57-64 66. (cost or costs or economic*).ti,ab. 67. exp "Costs and Cost Analysis"/ 68. cost-benefit analysis/ 69. ec.fs. 70. or/66-68 95. (Pubert* or Pubescen* or Prepubescen*).mp. 96. exp Pediatrics/ 97. (Pediatric* or Paediatric* or Padiatric*).mp. 98. exp Schools/ 99. (Nursery school* or Kindergar* or Primary school* or Secondary school* or Elementary school* or High school* or Highschool*).mp. 100. or/85-99 101. adolescent/ and adult/ 102. 100 not 101 103. (52 or 56) and 35 and 102 104. 13 and 102 and 65 and (84 or 46) 105. 13 and 46 and 102</p>
---	---

pa, pp 84. or/78-83 85. exp infant/ 86. (Infant* or infancy or Newborn* or Baby* or Babies or Neonat* or Preterm* or Prematur* or Postmatur*).mp. 87. exp Child/ 88. (Child* or Schoolchild* or School age* or Preschool* or Kid or kids or Toddler*).mp. 89. exp Adolescent/ 90. Adoles*.mp. 91. (Teen* or Boy* or Girl*).mp. 92. exp Minors/ 93. minors*.mp. 94. exp Puberty/	106. 13 and (56 or 65) and 46 and 102 107. 13 and 77 and 102 108. (13 or 84) and 70 109. 13 and (35 or 45) and 102 110. 77 and 65 and 13 and 102 111. (52 or 56) and (45 or 84) and 102 112. 13 and (56 or 65) and 70 113. 13 and 70 114. or/103-113 115. humans/ NOT (humans/ and animals) 116. 114 AND 115
--	--

Topic: ANA RF Anti CCP Testing Search Strategies

Database: EMBASE, 1980 - present

Search name: ANA RF Anti CCP Testing - EMBASE-amy

Notes: limits of: human, 1980-2009

Date searched: Jan 21, 2010

Results: 4,849

1. cyclic citrullinated peptide/ 2. ((anti adj ccp) or (citrullinated adj peptide*)).mp. 3. ((citrulline adj antibod*) or (anti-citrulline adj antibod*)).ti,ab. 4. exp Antinuclear Antibody/ 5. ((antinuclear adj antibod*) or (antinuclear adj factor*)).ti,ab. 6. (ana adj titer).ti,ab. 7. (ANA adj2 test*).ti,ab. 8. (FANA adj2 test*).ti,ab. 9. exp Rheumatoid Factor/ 10. rheumatoid factor*.ti,ab. 11. or/1-10 12. exp Systemic Lupus Erythematosus/ 13. (JSLE or SLE or "lupus erythematosus").ti,ab. 14. (grow* and (pain or pains)).ti,ab. 15. musculoskeletal diseases/ or arm/ or leg/ or extremities/ 16. pain/di, et 17. 15 and 16 18. exp arthralgia/ 19. arthralgia.ti,ab. 20. ((joint* adj pain*) or (limb* adj pain*)).ti,ab. 21. limp*.ti,ab. 22. Fibromyalgia/ 23. fibromyalgia.ti,ab. 24. benign.ti,ab. 25. exp Joint Instability/ or Joint hypermobility/ 26. (joint adj (instability or hypermobility)).ti,ab. 27. 24 and (25 or 26) 28. Patellofemoral Pain Syndrome/ 29. patellofemoral joint/ and pain/ 30. (patellofemoral adj pain adj syndrome).ti,ab. 31. knee pain/ or ankle pain/ 32. exp Synovitis/ or synovitis.mp. 33. "complex regional pain syndrome"/	37. or/35-36 38. exp child/ or (adolesc* or early or juvenile).ti,ab. 39. (JIA or JRA).ti,ab. 40. or/38-39 41. 37 and 40 42. exp Juvenile Rheumatoid Arthritis/ 43. ((juvenile or early) adj (rheumatoid or idiopathic) adj arthritis).ti,ab. 44. or/41-43 45. incidence/ or prevalence/ or seasonal variation/ 46. exp disease course/ 47. natural history.ti,ab. 48. or/45-47 49. exp mass screening/ or screening/ 50. exp "referral and consultation"/ 51. (screen* or refer*).ti,ab. 52. or/49-51 53. Differential Diagnosis/ 54. exp Reproducibility/ 55. exp "sensitivity and specificity"/ 56. Predictive Value of Tests/ 57. serodiagnosis/ 58. (di or bl or cl or im).fs. 59. exp Diagnostic Error/ 60. "diagnostic techniques and procedures"/ 61. diagnostic procedure/ 62. early diagnosis/ 63. Diagnostic Accuracy/ 64. physical examination/ 65. blood examination/ 66. "Pain Assessment"/ 67. or/53-66 68. (cost or costs or economic*).ti,ab. 69. exp economic aspect/ 70. cost-benefit analysis/ 71. ec.fs.
--	---

34. or/12-23,27-33 35. Arthritis/ 36. (\$arthritis or (\$articular adj arthritis)).ti,ab. 75. gender/ or sex difference/ 76. infection/ or infection*.ti,ab. 77. anxiety/ or (anxious* or anxiety).ti,ab. 78. comorbidity/ 79. or/73-78 80. exp newborn/ 81. (Infant* or infancy or Newborn* or Baby* or Babies or Neonat* or Preterm* or Prematur* or Postmatur*).mp. 82. exp Child/ 83. (Child* or Schoolchild* or School age* or Preschool* or Kid or kids or Toddler*).mp. 84. exp Adolescent/ 85. Adoles*.mp. 86. (Teen* or Boy* or Girl*).mp. 87. (minors* or juvenil*).mp. 88. exp Adolescence/ 89. (Pubert* or Pubescen* or Prepubescen*).mp. 90. exp Pediatrics/	72. or/68-70 73. exp demography/ or geographic distribution/ 74. age/ 91. (Pediatric* or Paediatric* or Peadiatric*).mp. 92. exp school/ or high school/ or kindergarten/ or middle school/ or nursery school/ or primary school/ 93. (Nursery school* or Kindergar* or Primary school* or Secondary school* or Elementary school* or High school* or Highschool*).mp. 94. or/80-93 95. (48 or 52) and 67 and 34 and 94 96. 11 and 94 and 67 and (34 or 44) 97. 11 and 94 and (34 or 44) 98. 11 and (67 or 52) and (34 or 44) and 94 99. 11 and 79 and 94 100. 11 and 79 and 67 101. 11 and 72 and (44 or 94) 102. 11 and 72 and (34 or 44) and 94 and 67 103. 11 and 94 and 67 104. or/95-103 105. adolescent/ and adult/ 106. 104 not 105 107. humans/ and animals/ 108. 106 not 107
---	--

Topic: ANA RF Anti CCP Testing Search Strategies

Database: CINAHL, 1960-present

Notes: limits of: human, 1980-2009

Date searched: Jan 21, 2010

Topic: ANA RF Anti CCP Testing Search Strategies

Database: Cochrane Library – CDSR and CENTRAL

Notes: limits: human, 1960-2009

Date searched: Feb 2010

Results: 216

#1(citrulline OR anti-ccp) or (rheumatoid factor) or (citrullinated peptide) or (antinuclear AND (antibod* OR factor*)) or (ANA OR FANA) and test* in Cochrane Reviews and Clinical Trials

#2 (lupus) or (JSLE OR SLE) or (limb or grow*) AND pain or (fibromyalgia) or (arthralgia) in Cochrane Reviews and Clinical Trials

#3 (joint AND (instability OR hypermobility)) or (patellofemoral pain syndrome) or (synovitis) or (JRA OR JIA) or (juvenile OR early) AND (rheumatoid OR idiopathic) AND arthritis in Cochrane Reviews and Clinical Trials

#4 (#2 OR #3)

#5 (incidence OR prevalence) or (disease AND (progression OR history)) or (natural history) in Cochrane Reviews and Clinical Trials

#6 (mass screening) or (screen*) or (refer*) or (consultation) or (referral and consultation) in Cochrane Reviews and Clinical Trials

#7 (sensitivity OR specificity) or (diagnostic error*) or (early diagnosis) or (delayed diagnosis) or (differential diagnosis) in Cochrane Reviews and Clinical Trials

#8 (demography) or (age of onset) OR (age factors) or (sex factors) or (infection) or (anxiety OR anxious*) in Cochrane Reviews and Clinical Trials

#9 (child*) or (adolescen*) or (infant* OR infancy OR newborn* OR baby OR babies) or (pediatric* OR paediatric*) in Cochrane Reviews and Clinical Trials

#10 (#1 AND #4 AND #7 AND #9) 42 edit delete #11 (#1 AND #4 AND #9)

#12 (#1 AND #8 AND #9) 185 edit delete #13 (#1 AND #4 AND #6 AND #9)

#14 (#4 AND #6 AND #9) 642 edit delete #15 ((#5 OR #6) AND #4 AND #9)

#16 (#10 OR #11 OR #12 OR #13 OR #14 OR #15)

#17 (#16), from 1960 to 2009

#18 (#10 OR #11 OR #12 OR #13)

#19 (#10 OR #11 OR #12 OR #13), from 1960 to 2009

Topic: ANA RF Anti CCP Testing Search Strategies

Database: Web of Science (ISI): Science Expanded, Social Sciences Expanded

Notes: limits: english only, human, 1980

Date searched: 9Feb10

Results: 856 (line #17)a

#17= #14 OR #13 OR #12 OR #10 OR #8

Refined by: Languages=(ENGLISH)

Databases=SCI-EXPANDED, SSCI Timespan=1960-2009

16= #14 OR #13 OR #12 OR #10 OR #8

Databases=SCI-EXPANDED, SSCI Timespan=1960-2009

15= #14 OR #12 OR #10 OR #8

Databases=SCI-EXPANDED, SSCI Timespan=1960-2009

14= #7 AND #6 AND #5 AND #1

Databases=SCI-EXPANDED, SSCI Timespan=1960-2009

13= #7 AND #6 AND #1

Databases=SCI-EXPANDED, SSCI Timespan=1960-2009

12= #11 AND #7 AND #5 AND #2

Databases=SCI-EXPANDED, SSCI Timespan=1960-2009

11= #4 OR #3

Databases=SCI-EXPANDED, SSCI Timespan=1960-2009

10= #9 AND #7 AND #2 AND #1

Databases=SCI-EXPANDED, SSCI Timespan=1960-2009

9= #5 OR #4

Databases=SCI-EXPANDED, SSCI Timespan=1960-2009

8= #7 AND #5 AND #2 AND #1

Databases=SCI-EXPANDED, SSCI Timespan=1960-2009

7= Topic=(infant*) OR Topic=(child*) OR Topic=(adolescen*) OR Topic=(pediatric* OR paediatric*)

Databases=SCI-EXPANDED, SSCI Timespan=1960-2009

6= Topic=(demography) OR Topic=((age factor*) OR (age of onset)) OR Topic=(sex factor* OR gender) OR

Topic=(infection*) OR Topic=(anxiety*) OR Topic=(comorbid*)

Databases=SCI-EXPANDED, SSCI Timespan=1960-2009

5= Topic=(early diagnos*) OR Topic=(differential diagnos*) OR Topic=(delayed diagnos*) OR Topic=(diagnostic error*)

Databases=SCI-EXPANDED, SSCI Timespan=1960-2009

4= Topic=(screen*) OR Topic=(referral OR refer*) OR Topic=(consultation OR consult*)

Databases=SCI-EXPANDED, SSCI Timespan=1960-2009

3= Topic=(incidence) OR Topic=(prevalence) OR Topic=(disease progression) OR Topic=(disease history) OR

Topic=(natural history)

Databases=SCI-EXPANDED, SSCI Timespan=1960-2009

2= Topic=(JSLE OR SLE OR lupus OR JIA OR JRA) OR Topic=((limb pain) OR (grow* pain)) OR

Topic=(fibromyalgia OR arthralgia) OR Topic=(joint AND (instability OR hypermobility)) OR Topic=((patellofemoral

pain syndrome) OR synovitis) OR Topic=((juvenile OR early) AND (rheumatoid OR idiopathic) and arthritis)

Databases=SCI-EXPANDED, SSCI Timespan=1960-2009

1= Topic=(antinuclear AND (antibod* OR factor* OR test*)) OR Topic=(anti-ccp OR (citrullinated peptide)) OR

Topic=((ANA OR FANA) AND (test*)) OR Topic=(rheumatoid factor*)

Topic: ANA RF Anti CCP Testing Search Strategies

Database: Academic Science Complete (EBSCO)

Notes: Limits: human, 1960

Date searched: 9Feb10

Results: 226

S20= S17 or S18 Limiters - Published Date from: 19600101-20091231
 S19= S17 or S18
 S18= S14 NOT S16
 S17= S14 and S15
 S16= Animal
 S15= Human
 S14= S8 or S9 or S10 or S11 or S12 or S13
 S13= S2 and S5 and S7
 S12= S2 and S4 and S7
 S11= S1 and S2 and S4 and S7
 S10= S1 and S2 and S7
 S9= S1 and S3 and S6 and S8
 S8= S1 and S2 and S3 and S7
 S7= Children or Pediatric or Paediatric or Infant or Adolescent or Adolescence
 S6= Demography or Gender or Anxiety or Infection
 S5= Incidence or Prevalence or Natural history
 S4= Screening or Referral or Consultation
 S3= Diagnostic errors or Diagnosis
 S2= Lupus or Rheumatoid arthritis or Fibromyalgia or Synovitis
 S1= citrulline or anti-ccp or Rheumatoid factor

Database	Dates searched	Date search ran	Number of results
MEDLINE (OVID)	1960-2009	14Jan10	5,389
EMBASE (OVID)	1980-2009	14Jan10	4,849
CINAHL (EBSCO)	1960-2009	14Jan10	374
CDSR & CENTRAL (Cochrane)	1960-2009	9Feb10	216
Web of Science (ISI)	1960-2009	9Feb10	856
Academic Search Complete (EBSCO)	1960-2009	9Feb10	226
TOTAL RESULTS			11,910

Search alerts:

- 1) PubMed
- 2) Web of Science
- 3) Scopus – Forward searching only

Appendix C. Forms

ANA, RF, and CCP Testing in Pediatric Populations INCLUSION/EXCLUSION FORM

Reviewer's initial: _____

Ref ID: _____

Criteria	Decision
1. REPORT OF PRIMARY RESEARCH	
2. POPULATION (at least two of the followings)	
a. Children and adolescents aged 18 years or younger	
AND (any of the followings)	
b1. Diagnosed with SLE or JIA/JRA/Still's disease	<input type="checkbox"/>
b2. With undiagnosed limb pains	<input type="checkbox"/>
b3. Given index test results	<input type="checkbox"/>
3. INDEX TEST (any of the followings)	
a. Antinuclear antibody (ANA)*	<input type="checkbox"/>
b. Rheumatoid factor (RF)†	<input type="checkbox"/>
c. Cyclic-citrullinated peptide (CCP)	<input type="checkbox"/>
EXCLUDE IF	
*Using non-human substrate for ANA or published before 1980	<input type="checkbox"/>
†Testing for hidden RF	<input type="checkbox"/>
4. REFERENCE STANDARD	
a. Clinical diagnosis of SLE or JIA/JRA/Still's disease	
5. REPORT OF NUMERICAL DATA (any of the following)	
a. Data sufficient to compute a 2x2 table	<input type="checkbox"/>
b. Data on subsequent impacts due to test results	<input type="checkbox"/>

Comments: _____

REVIEWER'S

DECISION[‡]: _____

FINAL DECISION: _____

Foreign language (if applicable): _____

[‡]Reason for exclusion (if applicable): _____

Article excluded, but may inform the followings:

- Background
- KQ1a
- KQ1b
- KQ2

QUADAS CODING GUIDE

1. Spectrum composition

This item examines the degree of similarity between the study's recruited sample and our population of interest (children with MSK symptoms) for the ANA review.

Yes The spectrum of patients included in the study was representative of those in whom the test will be used in practice. For cohort design, the recruited population should reflect a general children population (<18 years) with undiagnosed MSK pain.*

No† If the characteristics of participants were too specific. For example, in cohort design, the undiagnosed MSK pain children all have "stunted" growth. In the case arm for case-control design, all the JIA patients have uveitis. For case-control design, one study arm being the known disease (SJE/JIA) and the other arm being the healthy children with or without MSK pain.

Unclear Not reported or insufficient evidence by the study.

* Cohort and case-control study designs are illustrated in the following file: 'Ab testing_study designs.jpg' located in directory 'Z:\common\Arthritis Testing for children (AHRQ)\05_Data Extraction_Forms'.

† Note that if the participants' characteristics are drastically different from our desired population, it should be excluded altogether. If it is the case, describe in the comment section.

2. Selection criteria

This item refers to whether the study provides clear inclusion/exclusion criteria for participant selection.

Yes Detailed information on the source and inclusion/exclusion criteria was reported

No Information on the source and inclusion/exclusion criteria was poorly or not reported.

Unclear Information was partly reported but not sufficient to score a "Yes".

3. Reference standard

This item refers to whether the reference standard used can truly classify the disease status in participants.

Yes The study used widely accepted clinical standard, including but not limited to the ACR, ILAR, and Vancouver standards, for SLE or JIA.

No The reported standard was unlikely to correctly classify the disease status.

Unclear Not reported or insufficient evidence by the study.

4. Test result definition

This item examines whether or not the serological cutoff of positive index test is clearly stated.

Yes Serological cutoff was clearly stated (ie: titer, IU/mL, etc).

No Serological cutoff was not stated.

Unclear Not reported or insufficient evidence by the study.

5. Disease progression bias

Over a long period of time, disease status might change due to spontaneous recovery or disease progression. This item specifies a reasonable time frame in which the underlying disease status is expected not to change drastically.

Yes The time between the index test and reference standard was 6 months or less.

No The time between the index test and reference standard was longer than 6 months.

Unclear Not reported or insufficient evidence by the study.

6. Partial verification bias

This item refers to whether the selection of participants to receive confirmation using the reference standard might be biased.

Yes All participants, or a random selection of participants, who received the index test went on to receive verification by a reference standard. For cohort design, generally all participants will go through the same reference standard, whereas in case-control design, each study arm could be examined based on different reference standards (for example, the known JIA might use ILAR by rheumatologist, whereas, the healthy controls are based on general physician assessment).

No $\geq 20\%$ of the participants received the index test did not receive verification by the reference standard.

Unclear Not reported or insufficient evidence by the study.

7. Differential verification bias

This item examines whether or not all participants are receiving the same reference standard to verify the disease status.

Yes Same reference standard was used in all participants. Case-control design by default are likely to fall into this category since the patients were generally classified using the same reference standard.

No Different reference standards were used in $\geq 20\%$ of participants.

Unclear Not reported or insufficient evidence by the study.

8. Incorporation bias

This item examines whether the index test forms part of the reference standard. If it is the case, overestimated agreement is likely to result.

If no explicit statement indicating ANA is not part of the ACR standard, a “no” should be scored. On the contrary, if RF/CCP is compared against the JIA’s ILAR standard, assume a score of “yes” unless an explicit statement indicates that RF/CCP was, indeed, part of the reference standard.

Yes	Using RF/CCP for SLE/JIA, assumes a “yes” unless an explicit statement indicated otherwise. Using ANA for SLE, assume a “no” unless an explicit statement indicated otherwise.
No	Using RF/CCP for SLE/JIA, assumes a “yes” unless an explicit statement indicated otherwise. Using ANA for SLE, assume a “no” unless an explicit statement indicated otherwise.
Unclear	Not reported or insufficient evidence by the study.

9. Index test execution

Sufficient detail on carrying out the index test is important in tracking down the reasons for abnormal diagnostic results and in replicating the test in a different setting.

Yes	The index test was sufficiently described including information on both the assay method and substrate used.
No	Either or both the assay method and substrate used was not described.
Unclear	Not reported or insufficient evidence by the study.

10. Test review bias

This item examines whether the index test results are analyzed without the knowledge of the reference standard.

Yes	Index test results were interpreted without the knowledge of reference standard. If the index test was done and analyzed prior the execution of reference standard, it will automatically score a “yes”.
No	Index test results were certainly or likely to be interpreted with the knowledge of reference standard.
Unclear	Not reported or insufficient evidence by the study.

11. Reference standard review bias

This item examines whether the reference standard results are analyzed without the knowledge of the index test.

Yes	Reference standard results were interpreted without the knowledge of index test.
No	Reference standard results were interpreted with knowledge of index test. ANA for SLE will automatically score a “no” unless an explicit statement indicated otherwise.
Unclear	Not reported or insufficient evidence by the study.

12. Uninterpretable results

This item examines the reporting of the uninterpretable results from either index test or reference standard.

Yes	The study has no uninterpretable/indeterminate/intermediate results (ie: suspected, instead of definite, diagnosis of SLE/JIA). Or, if existed, these results were adequately
-----	---

reported.

No It was clear that the uninterpretable/indeterminate/intermediate results occur but was not reported.

Unclear No insufficient evidence by the study.

13. Withdrawals

This item examines the reporting of withdrawals.

Yes It was clear what happened to all patients who entered the study, for example if a flow diagram of study participants was reported explaining any withdrawals or exclusions, or the numbers recruited match those in the analysis. The drop out was not greater than 12% from the original sample size, and was appropriately accounted for.

No It appears that >12% patients recruited did not complete the study

Unclear Not reported or insufficient evidence by the study.

Guidelines for completing the QUADAS checklist

1) Spectrum composition

- Condition 1: In cohort design, the initial recruited sample consists solely of children with undiagnosed MSK pain. YES
- Condition 2: In case-control design, case arm as known disease (JIA/SLE) and control arm as healthy children. NO
- Condition 3: In case-control design, case arm as known disease (JIA/SLE) and control arm as other disease condition (ORG/NRG). NO
- Condition 4: In case-control design, case arm as known disease (JIA/SLE) along with specific characteristics (ie: JIA with uveitis or polyarticular JIA) and control arm as healthy children. NO
- Condition 5: In case-control design, case arm as known disease (JIA/SLE) along with specific characteristics (ie: JIA with uveitis or polyarticular JIA) and control arm as other disease condition (ORG/NRG). NO

2) Inclusion/exclusion criteria

- Condition 1: 3-5 of the following 5 selection criteria were used: Age, source of participants, method of diagnosis (for case-control design), condition of JIA/SLE (for case-control design), or any relevant demographic/clinical characteristic. YES
- Condition 2: 0-2 of the following 5 selection criteria were used: Age, source of participants, method of diagnosis (for case-control design), condition of JIA/SLE (for case-control design), or any relevant demographic/clinical characteristic. NO

3) Reference standard

- Condition 1: The diagnosis of JIA/SLE was based on a widely accepted diagnostic criteria including the ACR (American College of Rheumatology), ILAR (International League of Associations for Rheumatology), and Vancouver criteria. YES
- Condition 2: The diagnosis of JIA/SLE was based on other diagnostic criteria different from the ones listed under condition 1. UNSURE
- Condition 3: Patients were described as diagnosed with JIA/SLE by a healthcare professional. YES
- Condition 4: Patients were only described as diagnosed with JIA/SLE without further details. UNSURE

4) Test result definition

- Condition 1: Serological cutoff was explicitly stated (ie: in titer, IU/mL, or AU), thus what constitutes a positive/negative test was clear. YES
- Condition 2: Serological cutoff was not explicitly stated, thus what constitutes a positive/negative test was unclear. UNSURE
- Condition 3: Serological cutoff was not used, only mean measures were provided. NO

5) Disease progression bias

- Condition 1: The interval between the index test and reference standard was 6 months or less. YES
- Condition 2: The interval between the index test and reference standard was more than 6 months. NO
- Condition 3: In case control design, the mean disease duration of JIA/SLE was greater than 6 months and no reason to suggest that the index test was conducted within 6 months of the diagnosis. NO

6) Partial verification bias

- Condition 1: The whole or a random sample of participants with an index test was selected to receive the clinical diagnosis. YES
- Condition 2: Only a non-random sample of participants was selected, based on their index test results, to receive the diagnosis. NO
- Condition 3: In most case-control studies, in which, the diagnosis was generally done before the index test. YES

7) Differential verification bias

- Condition 1: Only one set of diagnostic criteria was used for JIA/SLE. YES
- Condition 2: More than 1 sets of diagnostic criteria were used for JIA/SLE. NO

8) Incorporation bias

- Condition 1: RF/CCP as index test against JIA/SLE diagnosis. YES
- Condition 2: ANA as index test against JIA diagnosis. YES
- Condition 3: ANA as index test against SLE diagnosis. NO
- Condition 4: ANA as index test against SLE diagnosis using the ACR criteria. NO
- Condition 5: ANA as index test against SLE diagnosis using other criteria. NO
- Condition 6: ANA as index test against SLE diagnosis, but explicitly stated that ANA was excluded from the diagnostic criteria. YES

9) Index test execution

- Condition 1: Both the assay method and substrate used were described. YES
- Condition 2: Either the assay method or substrate used was described. UNSURE
- Condition 3: Neither the assay method nor the substrate used was described. NO

10) Test review bias

- Condition 1: The index test was done before the diagnosis of JIA/SLE. YES
- Condition 2: The diagnosis of JIA/SLE was done before index test and explicitly stated that the interpretation of the index test results was blinded to the diagnosis. YES
- Condition 3: The diagnosis of JIA/SLE was done before index test but not explicitly stated that the interpretation of the index test results was blinded to the diagnosis. YES
- Condition 4: The index test results were likely to be interpreted with the knowledge of JIA/SLE diagnosis. UNSURE
- Condition 5: The index test results were interpreted and skewed with the knowledge of JIA/SLE diagnosis. NO

11) Reference standard review bias

- Condition 1: The diagnosis of JIA/SLE was done before the index test. YES
- Condition 2: The index test was done before the diagnosis of JIA/SLE and explicitly stated that the interpretation of diagnosis was blinded to index test results. YES
- Condition 3: The index test was done before the diagnosis of JIA/SLE but not explicitly stated that the interpretation diagnosis was blinded to index test results. UNSURE
- Condition 4: The diagnoses of JIA/SLE were likely to be interpreted with the knowledge of index test. NO

12) Uninterpretable results

- Condition 1: No uninterpretable results (ie: serological cutoff is used and patients were classified as positive or negative). YES
- Condition 2: The uninterpretable results were adequately described. YES
- Condition 3: The uninterpretable results were likely to exist but not reported by authors. NO
- Condition 4: Could not determine whether all the study results were reported. UNSURE

13) Withdrawal

- Condition 1: No withdrawal. YES
- Condition 2: Withdrawal was small ($\leq 10\%$) and with description on reasons of dropouts. YES
- Condition 3: Withdrawal was small ($\leq 10\%$) and without description on reasons of dropouts. UNSURE
-
-
- Condition 4: Withdrawal was large ($>10\%$) and with description on reasons of dropouts. UNSURE
- Condition 5: Withdrawals was large ($>10\%$) and without description on reasons for dropouts. NO

Appendix D. List of Excluded Studies

EXCLUDED STUDIES (N = 438)

The following studies failed to meet at least one of the pre-specified inclusion criteria.

Study design (N = 129)

The following studies were excluded because they were not reports of primary research.

1. Miller E, Uleryk E, Doria AS. Evidence-based outcomes of studies addressing diagnostic accuracy of MRI of juvenile idiopathic arthritis. *AJR* 2009;American(5):1209-18.
2. Kallel-Sellami M, Baili-Klila L, Zerzeri Y, et al. Pediatric systemic lupus erythematosus with C1q deficiency. *Annals of the New York Academy of Sciences* 2007;1108:193-6.
3. Melegari A, Mascia MT, Sandri G, et al. Immunodeficiency and autoimmune phenomena in female hyper-IgM syndrome. *Annals of the New York Academy of Sciences* 2007;1109:106-8.
4. Mseddi M, Dammak A, Marrekchi S, et al. Subacute cutaneous lupus erythematosus in childhood: a case report [French]. *Archives de Pediatrie* 2007;14(2):164-6.
5. Gough-Palmer A, McHugh K. Investigating hip pain in a well child. *BMJ: British Medical Journal* 2007;334(7605):1216-7.
6. Banerjee P, Crain B. 2-year-old girl with right leg weakness. *Brain Pathology* 2008;18(4):608-10.
7. Abbassian A. The limping child: a clinical approach to diagnosis. *British Journal of Hospital Medicine* 2007;68(5):246-50.
8. Trueman CA. Joint disease: the future arthritis burden. *Caring* 2009;28(2):8-13.
9. Gilbert NF, Deavers MT, Madewell JE, et al. A 16-year-old girl with pain and swelling in the medial clavicle. *CLIN ORTHOP RELATED RES* 2008;466(12):3158-62.
10. Brown RJ. Introduction to the special issue on medically unexplained symptoms: background and future directions. *Clinical Psychology Review* 2007;27(7):769-80.
11. Wyndham M. The limping child. *Community Practitioner* 2007;80(9):42.
12. Syed RH, Gilliam BE, Moore TL. Rheumatoid factors and anticyclic citrullinated peptide antibodies in pediatric rheumatology. *Current Rheumatology Reports* 2008;10(2):156-63.
13. Yokota S, Mori M, Imagawa T, et al. Proposal for juvenile idiopathic arthritis guidance on diagnosis and treatment for primary care pediatricians and nonpediatric rheumatologists. *Modern Rheumatology* 2007;17(5):353-63.
14. Iwata N, Mori M, Miyamae T, et al. Sjogren's syndrome associated with childhood-onset systemic lupus erythematosus [Japanese]. *Nihon Rinsho Meneki Gakkai Kaishi* 2008;31(3):166-71.
15. van Holsbeeck MT. A role for US screening in juvenile idiopathic arthritis. *Pediatric Radiology* 2007;37(7):623-4.
16. Gottlieb BS, Ilowite NT. Systemic lupus erythematosus in children and adolescents. *Pediatrics in Review* 2006;27(9):323-30.
17. Al-Mendalawi MD. Juvenile systemic lupus erythematosus in Bahrain. A tertiary referral center experience. *Saudi Medical Journal* 2009;30(9):1240-1.
18. Quartier P, Prieur AM. Systemic lupus erythematosus [French]. *Archives de Pediatrie* 2003;10(4):367-73.
19. Gandon-Laloum S. Growth and limb pains [French]. *Archives de Pediatrie* 2006;13(6):550-2.
20. Bizzaro N, Wiik A. Appropriateness in anti-nuclear antibody testing: from clinical request to strategic laboratory practice. *Clinical & Experimental Rheumatology* 2004;22(3):349-55.
21. Jarvis JN. The unique clinical presentation of children with chronic arthritis: putting the pediatrics in pediatric rheumatology. *Current Problems in Pediatric & Adolescent Health Care* 2006;36(3):80-2.
22. Kaur S, Thami GP. Antinuclear antibody-negative systemic lupus erythematosus: revisited. *INDIAN J PEDIATR* 2003;70(2):185-6.
23. Leung AK, Lemay JF. The limping child. *Journal of Pediatric Health Care* 2004;18(5):219-23.
24. Gunther KP, Thielemann F, Bottesi M. Anterior knee pain in children and adolescents. Diagnosis and conservative treatment [German]. *Orthopade* 2003;32(2):110-8.

25. Shea KG, Pfeiffer R, Curtin M. Idiopathic anterior knee pain in adolescents. *Orthopedic Clinics of North America* 7 A.D.;34(3):377-83.
26. Siegel DM. Antinuclear Antibody (ANA) Testing. *Pediatrics in Review* 2003;24(9):320-1.
27. Ganley TJ, Gaugles RL, Moroz LA. Consultation with the specialist: patellofemoral conditions in childhood. *Pediatrics in Review* 70 A.D.;27(7):264-9.
28. Kashikar-Zuck S, Graham TB, Huenefeld MD, et al. A review of biobehavioral research in juvenile primary fibromyalgia syndrome. *ARTHRITIS CARE RES* 2000;13(6):388-97.
29. Locham KK, Singh J, Garg R, et al. ANA negative lupus erythematosus. *Indian Pediatrics* 2000;37(5):540-2.
30. Solomon DH, Shmerling RH, Schur PH, et al. A computer based intervention to reduce unnecessary serologic testing. *J RHEUMATOL* 1999;26(12):2578-84.
31. Carreno PL. Practical value of immunologic changes in juvenile chronic arthritis] [Spanish]. *Anales Espanoles de Pediatria* 1993;39:Suppl-30.
32. Modesto CC. Antinuclear antibodies and juvenile chronic rheumatoid arthritis] [Spanish]. *Anales Espanoles de Pediatria* 1996;44(4):305-9.
33. Lightfoot RW, Jr. Cost effective use of laboratory tests in rheumatology. *Bulletin on the Rheumatic Diseases* 1997;46(6):1-3.
34. Martinez-Cordero E, Orozco BG, Martinez ME. Evaluation of disease activity by laboratory tests in juvenile rheumatoid arthritis. *Journal of Investigational Allergology & Clinical Immunology* 1995;5(4):216-20.
35. Brady M. The child with a limp. *Journal of Pediatric Health Care* 1993;7(5):226-8.
36. Leak AM. Autoantibody profile in juvenile chronic arthritis. *ANN RHEUM DIS* 1988;47(3):178-82.
37. Southwood TR, Malleson PN. Antinuclear antibodies and juvenile chronic arthritis (JCA): search for a specific autoantibody associated with JCA. *ANN RHEUM DIS* 1991;50(9):595-8.
38. Schaller JG. Juvenile rheumatoid arthritis: Series I. *Arthritis & Rheumatism* 1977;20(2:Suppl):70.
39. Chamberlain MA. Referral of chronic arthritics. *British Medical Journal Clinical Research Ed* 1984;288(6414):347-8.
40. Ballardini P, Busachi CA, Amoresano A, et al. Prevalence of rheumatoid factors in juvenile rheumatoid arthritis. *Clinical & Experimental Rheumatology* 2010;9(5):548.
41. Kanski JJ. Juvenile arthritis and uveitis. *Survey of Ophthalmology* 1990;34(4):253-67.
42. Mayet WJ, Bachmann M, Hermann E, et al. The Ro/SS-A antigen-antibody system [German]. *Zeitschrift fur Rheumatologie* 1988;47(2):80-5.
43. Sievers K, Nissila M, Sievers UM. Cerebral vasculitis visualized by angiography in juvenile rheumatoid arthritis simulating brain tumor. *Acta Rheumatologica Scandinavica* 1968;14(3):222-32.
44. Oppermann J. Phagocytized aggregates in joint effusion cells of children with rheumatoid arthritis [German]. *Allergie und Asthma* 1969;15(4):218-22.
45. Irias JJ. Hydralazine-induced lupus erythematosus-like syndrome. *American Journal of Diseases of Children* 1975;129(7):862-4.
46. Bloch-Michel H, Waltzing P, Brauner M, et al. The rosette rheumatoid test in seronegative juvenile-infantile polyarthritis [French]. *Annales de Medecine Interne* 1972;123(4):375-9.
47. Yodfat Y, Yossipovitch Z, Cohen I, et al. A family with a high incidence of juvenile rheumatoid arthritis. *ANN RHEUM DIS* 1972;31(2):92-4.
48. Bresnihan FP, Ansell BM. Effect of penicillamine treatment on immune complexes in two cases of seropositive juvenile rheumatoid arthritis. *ANN RHEUM DIS* 1975;35(5):463-5.
49. Gregg S. Rheumatoid arthritis in childhood. *Arizona Medicine* 1971;28(8):577-85.
50. Baum J, Fink C. Juvenile rheumatoid arthritis in monozygotic twins: a case report and review of the literature. *Arthritis & Rheumatism* 1968;11(1):33-6.
51. Brizard J. Chronic inflammatory rheumatism in children [French]. *Cahiers de Medecine* 1972;13(3):167-79.
52. Bruneau CD, Edmonds JP, Hughes GR, et al. Detection and characterization of DNA-anti-DNA complexes in a patient with systemic lupus erythematosus. *Clinical & Experimental Immunology* 1977;28(3):433-6.
53. Ansell BA. Rheumatoid disease, laboratory data. *CRC Critical Reviews in Clinical Laboratory Sciences* 1972;3(2):193-201.
54. Day NK, Geiger H, McLean R, et al. C2 deficiency. Development of lupus erythematosus. *Journal of Clinical Investigation* 1973;52(7):1601-7.
55. Kornreich H, Malouf NN, Hanson V. Acute hepatic dysfunction in juvenile rheumatoid arthritis. *J Pediatric* 1971;79(1):27-35.
56. Mackay IR. Autoimmune disease. *Medical Journal of Australia* 1969;1(13):696-9.
57. Schaller JG. Diagnosis and treatment of arthritis in children. *Medical Times* 1977;105(11):65-74.
58. Kolle G. Juvenile rheumatoid arthritis and related collagen diseases. Clinical aspects [German]. *Monatsschrift fur Kinderheilkunde* 1976;124(12):779-85.
59. Hausteiner UF, Sonnichsen N. Lupus erythematosus in childhood--a virus-induced autoimmune disease? [German]. *Padiatrie und Grenzgebiete* 1974;13(2-3):163-76.
60. Boone JE, Baldwin J, Levine C. Juvenile rheumatoid arthritis. *Pediatric Clinics of North America* 1974;21(4):885-915.

61. Levinson JE. Juvenile rheumatoid arthritis. *POSTGRAD MED* 1972;51(6):88-94.
62. Dumas M, Loyau G. Diagnostic problems raised by polyarthritis of an inflammatory nature [French]. *Semaine des Hopitaux* 1972;48(9):645-8.
63. Smiley WK. The eye in juvenile rheumatoid arthritis. *Transactions of the Ophthalmological Societies of the United Kingdom* 1974;94(3):817-29.
64. Macaubas C, Nguyen K, Milojevic D, et al. Oligoarticular and polyarticular JIA: Epidemiology and pathogenesis. *Nature Reviews Rheumatology* 2009;5(11):616-26.
65. Goldenberg DL. Diagnosis and Differential Diagnosis of Fibromyalgia. *American Journal of Medicine* 2009;122(12 SUPPL.):S14-S21.
66. Hunter PJ, Wedderburn LR. Pediatric rheumatic disease: Can molecular profiling predict the future in JIA? *Nature Reviews Rheumatology* 2009;5(11):593-4.
67. Ortega-Hernandez O-D, Shoenfeld Y. Infection, vaccination, and autoantibodies in chronic fatigue syndrome, cause or coincidence. *Contemporary Challenges in Autoimmunity Annals of the New York Academy of Sciences* 2009;1173:600-9.
68. McGonagle D, Benjamin M. Towards a new clinico-immunopathological classification of juvenile inflammatory arthritis. *J RHEUMATOL* 2009;36(8):1573-4.
69. Ringold S, Cron RQ. The temporomandibular joint in juvenile idiopathic arthritis: Frequently used and frequently arthritic. *Pediatric Rheumatology* 2009;7(11).
70. Nandi M, Ganguli SK, Mondal R, et al. Clinico-serological profile of juvenile idiopathic arthritis. *Indian Pediatrics* 2009;46(7):640-1.
71. Goeb V, Jouen F, Gilbert D, et al. Diagnostic and prognostic usefulness of antibodies to citrullinated peptides. *Joint Bone Spine* 2009;76(4):343-9.
72. Gomez AP, Atrio AIS, Nieves AIT, et al. Differential diagnostic protocol of arthritis in infancy and adolescence [Spanish]. *Medicine* 2009;10(29):1972-5.
73. Gomez AP, Atrio AIS, Hernandez FA, et al. Juvenile idiopathic arthritis [Spanish]. *Medicine* 2009;10(29):1933-41.
74. Syed RH, Gilliam BE, Moore TL. Rheumatoid factors and anticyclic citrullinated peptide antibodies in pediatric rheumatology. *Current Rheumatology Reports* 2008;10(2):156-63.
75. Unsal E, Arli AO, Akman H. Rhupus arthropathy as the presenting manifestation in Juvenile SLE: A case report. *Pediatric Rheumatology* 2007;5(7).
76. Hinze C, Brunner HI. Pediatric rheumatology - Its own specialty. *Nature Clinical Practice Rheumatology* 2008;4(6):279.
77. Olsen NJ, Wandstrat AE, Karp DR. Development of risk profiles for systemic lupus erythematosus. *Future Rheumatology* 2007;2(5):507-14.
78. Silverman E. Pediatric systemic lupus erythematosus. *Future Rheumatology* 2007;2(1):23-50.
79. Weiss JE, Ilowite NT. Juvenile Idiopathic Arthritis. *Rheumatic Disease Clinics of North America* 2007;33(3):441-70.
80. Yildiz B, Kural N. IgG1 deficiency and high IgA level with juvenile idiopathic arthritis. *European Journal of Pediatrics* 2007;166(11):1179-80.
81. Buc M, Parnicka Z, Rovensky J, et al. Juvenile idiopathic arthritis an overview of current status in immunopathogenesis, immunogenetic, and immunotherapy [Slovak]. *Rheumatologia* 2006;20(4):209-14.
82. Gottlieb BS, Ilowite NT. Systemic lupus erythematosus in children and adolescents. *Pediatrics in Review* 2006;27(9):323-30.
83. Lopez-Longo F-J, Rodriguez-Mahou M, Sanchez-Ramon S, et al. Anti-cyclic citrullinated peptide versus anti-Sa antibodies in diagnosis of rheumatoid arthritis in an outpatient clinic for connective tissue disease and spondyloarthritis. *J RHEUMATOL* 2006;33(8):1476-81.
84. Ravelli A, Martini A. Remission in juvenile idiopathic arthritis. *Clinical and experimental rheumatology* 2006;24(6 SUPPL. 43):S105-S110.
85. Uma SA. Systemic lupus erythematosus in children-clinical spectrum and management. *Indian Journal of Practical Pediatrics* 2005;7(4):352-8.
86. Childhood SLE. *South African Medical Journal* 2005;95(6):363.
87. Kamoun M. Diagnostic performance and predictive value of anti-citrullinated peptide antibodies for diagnosis of rheumatoid arthritis: Toward more accurate detection? *Clinical Chemistry* 2005;51(1):12-3.
88. Migliorini P, Baldini C, Rocchi V, et al. Anti-Sm and anti-RNP antibodies. *Autoimmunity* 2005;38(1):47-54.
89. Symmons D. Commentary: Juvenile idiopathic arthritis - Issues of definition and causation. *International Journal of Epidemiology* 2005;34(3):671-2.
90. Vander CB, Peene I, Cantaert T, et al. Anti-citrullinated protein/peptide antibodies (ACPA) in rheumatoid arthritis: Specificity and relation with rheumatoid factor. *Autoimmunity Reviews* 2005;4(7):468-74.
91. Wiik AS. Anti-nuclear autoantibodies: Clinical utility for diagnosis, prognosis, monitoring, and planning of treatment strategy in systemic immunoinflammatory diseases. *Scandinavian Journal of Rheumatology* 2005;34(4):260-8.
92. Yablanski K, Yordanova V. Diagnostic significance of the antibodies to cyclic citrullinated peptides in patients with rheumatoid

- arthritis. *Clinical Application of Immunology* 2005;4(1-2):435-40.
93. Maddison PJ. Is it SLE? *Best Practice and Research in Clinical Rheumatology* 2002;16(2):167-80.
 94. Agarwal V, Mahajan A. Laboratory tests in rheumatology: Rational utilization and interpretation. *JK Science* 2004;6(1):52-5.
 95. Bodman-Smith MD, Fife MF, Wythe H, et al. Anti-BiP antibody levels in juvenile idiopathic arthritis (JIA). *Rheumatology* 2004;43(10):1305-6.
 96. Klein-Gitelman M. Pediatric lupus versus adult lupus role of the laboratory. *Clinical and Applied Immunology Reviews* 2004;4(5):333-50.
 97. Petty RE, Southwood TR, Manners P, et al. International League of Associations for Rheumatology Classification of Juvenile Idiopathic Arthritis: Second Revision, Edmonton, 2001. *J RHEUMATOL* 2004;31(2):390-2.
 98. Ravelli A. Toward an understanding of the long-term outcome of juvenile idiopathic arthritis. *Clinical and experimental rheumatology* 2004;22(3):271-5.
 99. van Venrooij WJ, Hazes JM, Visser H. Anticitrullinated protein/peptide antibody and its role in the diagnosis and prognosis of early rheumatoid arthritis. *Netherlands Journal of Medicine* 2002;60(10):383-8.
 100. Lewis JG, Florkowski CM, Elder PA, et al. Rheumatoid factor and false positive sex-hormone binding globulin. *Clinica Chimica Acta* 2003;332(1-2):139-44.
 101. Slettjord CN, Nossent HC, Goodson NJ, et al. Increased mortality in early inflammatory polyarthritis: Comment on the article by Goodson et al. *Arthritis and rheumatism* 2003;48(5):1462-3.
 102. Goumy L. Idiopathic juvenile polyarthritis [French]. *Presse Medicale* 2001;30(3):137-42.
 103. Hull RG. Guidelines for management of childhood arthritis. *Rheumatology* 2001;40(11):1309-12.
 104. Malleson PN, Beauchamp RD. *Rheumatology*: 16. Diagnosing musculoskeletal pain in children. *Canadian Medical Association Journal* 2001;165(2):183-8.
 105. Rauz S, Murray PI, Southwood TR. Juvenile idiopathic arthritis and uveitis: The classification conundrum. *Eye* 2000;14(6):817-20.
 106. Diagnosis, development and treatment principles of systemic lupus erythematosus [French]. *Annales de Dermatologie et de Venerologie* 2000;127(SPEC.ISS. 1):A167-A173.
 107. Illei GG, Klippel JH. Why is the ANA result positive? *Bulletin on the Rheumatic Diseases* 1999;48(1):1-4.
 108. Pelkonen PM. Juvenile arthritis with oligoarticular onset. *Bailliere's Clinical Rheumatology* 1998;12(2):273-86.
 109. Rosenberg AM. Systemic lupus erythematosus in children. *Springer Seminars in Immunopathology* 1994;16(2-3):261-79.
 110. Tibbitts GM. Juvenile rheumatoid arthritis: Old challenges, new insights. *POSTGRAD MED* 1994;96(2):75-87.
 111. Tucker LB. Juvenile rheumatoid arthritis. *Current Opinion in Rheumatology* 1993;5(5):619-28.
 112. Hanson V. The subtypes of juvenile rheumatoid arthritis and their significance. *Ryumachi* 1991;31(6):592-8.
 113. Bonagura VR, Ilowite NT, Hatam L, et al. Expression of the major rheumatoid factor cross-reactive idiotype in pediatric patients with systemic lupus erythematosus. *Clinical Immunology and Immunopathology* 1991;60(2):232-43.
 114. Laxer RM. Introduction. *J RHEUMATOL* 1990;17(SUPPL. 26):1.
 115. Rhodes VJ. Physical therapy management of patients with juvenile rheumatoid arthritis. *Physical Therapy* 1991;71(12):910-9.
 116. Southwood TR, Malleson PN. Antinuclear antibodies and juvenile chronic arthritis (JCA): Search for a specific autoantibody associated with JCA. *ANN RHEUM DIS* 1991;50(9):593-8.
 117. Ansell BM. Juvenile chronic arthritis, juvenile rheumatoid arthritis, and inflammatory arthropathies of childhood. *Current Opinion in Rheumatology* 1990;2(5):799-803.
 118. Miller III JJ. Immunologic abnormalities of juvenile arthritis. *Clinical Orthopaedics and Related Research* 1990;(259):23-30.
 119. Martini A. Immunological approach to juvenile rheumatoid arthritis [French]. *Annales de Pediatrie* 1988;35(8):555-8.
 120. John M, John V, Oppermann J. Laboratorial findings in the case of juvenile chronic arthritis [German]. *Zeitschrift fur Klinische Medizin* 1985;40(19):1445-8.
 121. Wahn V. Anti-immunoglobulins (rheumatoid factors) - Immunobiology and relevance in pediatric practice [German]. *Monatsschrift fur Kinderheilkunde* 1985;133(4):196-200.
 122. Moore TL, Weiss TD. Immunologic studies in juvenile arthritis. *Bulletin on the Rheumatic Diseases* 1982;32(3):25-9.
 123. Gupta K, Chintu C, Raghu MB. Juvenile rheumatoid arthritis in Zambian children. *East African Medical Journal* 1981;58(5):344-53.
 124. Ulybina OV, Shakhbazyan IE, Komissarova IA. Significance of some cytochemical and immunological indices in the clinical characteristics of rheumatoid arthritis children [Russian]. *Pediatriya - Zhurnal im GN Speranskogo* 1980;59(4):40-4.
 125. Keren DF. Antinuclear antibody testing. *CLIN LAB MED* 2002;22(2):447-74.

126. Price S. Autoantibody profiles linked to ethnicity in pediatric SLE. *Nature Reviews Rheumatology* 2009;5(4):182.
127. Schaller J, Wedgwood RJ. Juvenile rheumatoid arthritis: a review. *Pediatrics* 1972;50(6):940.
128. De AT, Davey R, Solanki K, et al. Systemic lupus erythematosus (SLE) in a paediatric unit. *New Zealand Medical Journal* 2010;123(1313):77-9.
129. Kobayashi R, Mii S, Nakano T, et al. Neonatal lupus erythematosus in Japan: A review of the literature. *Autoimmunity Reviews* 2009;8(6):462-6.

Population (N = 77)

The following studies were excluded because the study population was not consisted primarily of individuals ≤ 18 years with a diagnosis of JIA, SLE, or with undiagnosed MSK pain .

1. Allen RC, Dewez P, Stuart L, et al. Antinuclear antibodies using HEp-2 cells in normal children and in children with common infections. *Journal of Paediatrics & Child Health* 1991;27(1):39-42.
2. Alpigiani MG, Cerboni M, Bertini I, et al. Endocrine autoimmunity in young patients with juvenile chronic arthritis. *Clinical & Experimental Rheumatology* 2002;20(4):565-8.
3. Arinola OG, Salimonu LS. Rheumatoid factor in sera of Nigerian school children with urinary schistosomiasis. *African Journal of Medicine & Medical Sciences* 2000;29(3-4):329-30.
4. Attar SM, Bunting PS, Smith CD, et al. Comparison of the anti-cyclic citrullinated peptide and rheumatoid factor in rheumatoid arthritis at an arthritis center. *Saudi Medical Journal* 2009;30(3):446-7.
5. Awoyinfa JD, Batubo TA, McFarlane H, et al. Rheumatoid factor in Nigerians. *African Journal of Medical Sciences* 1971;2(1):49-55.
6. Barone C, Bartoloni C, Gentiloni N, et al. Systemic lupus erythematosus with only IgE-class antinuclear antibodies. *Arthritis & Rheumatism* 1981;24(11):1441-3.
7. Berkun Y, Zandman-Goddard G, Barzilai O, et al. Infectious antibodies in systemic lupus erythematosus patients. *Lupus* 2009;18(13):1129-35.
8. Bonaguri C, Melegari A, Dall'Aglio P, et al. An Italian multicenter study for application of a diagnostic algorithm in autoantibody testing. *Annals of the New York Academy of Sciences* 2009;1173:124-9.
9. Bortolotti F, Vajro P, Balli F, et al. Non-organ specific autoantibodies in children with chronic hepatitis C. *Journal of Hepatology* 1996;25(5):614-20.
10. Carreno L, Lopez-Longo FJ, Monteagudo I, et al. Immunological and clinical differences between juvenile and adult onset of systemic lupus erythematosus. *Lupus* 1999;8(4):287-92.
11. Che Maraina CH, Rusni D, Azwany YN. Do antibodies against modified citrullinated vimentin have a value in the ACR criteria for rheumatoid arthritis? 2010 p. 62.
12. Correia ML, Carvalho S, Fortuna J, et al. Comparison of three anti-CCP antibody tests and rheumatoid factor in RA and control patients. *Clinical Reviews in Allergy and Immunology* 2008;34(1):21-5.
13. DeHoratius RJ, Pillarisetty R, Messner RP, et al. Anti-nucleic acid antibodies in systemic lupus erythematosus patients and their families. Incidence and correlation with lymphocytotoxic antibodies. *Journal of Clinical Investigation* 1975;56(5):1149-54.
14. Dias BL, Imamura EU, Izumi AP, et al. Juvenile idiopathic arthritis with dry synovitis: clinical case and review of literature [Portuguese]. *Acta Reumatologica Portuguesa* 2009;34(3):541-5.
15. El-Awady HM, El-Wakkad ASE-D, Saleh MT, et al. Serum Melatonin in Juvenile Rheumatoid Arthritis: Correlation with Disease Activity. *Pakistan Journal of Biological Sciences* 2007;10(9):1471-6.
16. El-Chennawi FA, Mosaad YM, Habib HM, et al. Comparative study of antinuclear antibody detection by indirect immunofluorescence and enzyme immunoassay in lupus patients. *Immunological Investigations* 2009;38(8):839-50.
17. El-Sehemy MS, Al-Saaran AM, Baddour NM, et al. Comparative clinical prospective therapeutic study between cyclophosphamide, cyclosporine and azathioprine in the treatment of lupus nephritis. *The Egyptian journal of immunology / Egyptian Association of Immunologists* 2006;13:39-52.
18. Ferreira M, Salgueiro AB, Estrada J, et al. [Lupus erythematosus]. [Portuguese]. *Acta Medica Portuguesa* 2008;21(2):199-204.
19. Frot AS, Barbarot S, Poignant S, et al. Ecchymotic angioedema revealing childhood systemic lupus erythematosus with anti-C1q antibodies [French]. *Annales de Dermatologie et de Venereologie* 2008;135(8-9):584-6.
20. Ganesh R, Ramalingam V, Eswara RT, et al. Antinuclear antibodies in Mycobacterium tuberculosis infection. *INDIAN J PEDIATR* 2008;75(11):1188.
21. Gillespie JP, Lindsley CB, Linshaw MA, et al. Childhood systemic lupus erythematosus with negative antinuclear antibody test. *J Pediatric* 1981;98(4):578-81.
22. Gregorio GV, Jones H, Choudhuri K, et al. Autoantibody prevalence in chronic hepatitis B virus infection: effect in interferon alfa. *Hepatology* 1996;24(3):520-3.
23. Gregorio GV, Pensati P, Iorio R, et al. Autoantibody prevalence in children with liver disease due to chronic hepatitis C virus (HCV) infection. *Clinical & Experimental Immunology* 1998;112(3):471-6.
24. Gregorio GV, Davies ET, Mieli-Vergani G, et al. Significance of extractable nuclear antigens in childhood autoimmune liver disease. *Clinical & Experimental Immunology* 1995;102(2):308-13.
25. Hafez M, El-Battoty MF, Hawas S, et al. Susceptibility to and severity of rheumatoid

- arthritis in multicase families. *British Journal of Rheumatology* 1991;30(3):181-5.
26. Hamade IH, Al Shamsi HN, Al DH, et al. Uveitis survey in children. *British Journal of Ophthalmology* 2009;93(5):569-72.
 27. Hazzan R, Mukamel M, Yacobovich J, et al. Risk factors for future development of systemic lupus erythematosus in children with idiopathic thrombocytopenic purpura. *Pediatric Blood & Cancer* 2006;47(5):657-9.
 28. Hedberg H. The total complement activity of synovial fluid in juvenile forms of arthritis. *Acta Rheumatologica Scandinavica* 1971;17(4):279-85.
 29. Hodkinson B, Meyer PW, Musenge E, et al. The diagnostic utility of the anti-CCP antibody test is no better than rheumatoid factor in South Africans with early rheumatoid arthritis. *Clinical Rheumatology* 2010;29(6):615-8.
 30. Itoh Y, Fukunaga Y, Igarashi T, et al. Autoimmunity in chronic fatigue syndrome in children. *Japanese Journal of Rheumatology* 1998;8(4):429-37.
 31. Jundt JW, Creager AH. Star Complexes - Febrile Illnesses Associated with Sore Throat, Arthritis, and Rash. *Southern Medical Journal* 1993;86(5):521-8.
 32. Kang M, Wang HW, Cheng PX, et al. Lack of association between mannose-binding lectin gene polymorphisms and juvenile idiopathic arthritis in a Han population from the Hubei province of China. *Arthritis Research & Therapy* 2006;8(4):R85.
 33. Kasapcopur O, Ozbakir F, Arisoy N, et al. Frequency of antinuclear antibodies and rheumatoid factor in healthy Turkish children. *Turkish Journal of Pediatrics* 1999;41(1):67-71.
 34. Kenesi-Laurent M-A, Kaplan G, Kahn M-F. Oligoarthritis with antinuclear antibodies. A new syndrome equivalent to juvenile oligoarthritis with ANA [French]. *Revue du Rhumatisme et des Maladies Osteo-Articulaires* 1991;58(1):1-6.
 35. Kotaniemi K, Kaipainen-Seppanen O, Savolainen A, et al. A population-based study on uveitis in juvenile rheumatoid arthritis. *Clinical & Experimental Rheumatology* 1999;17(1):119-22.
 36. Krishnan C, Kaplan MH. Immunopathologic studies of systemic lupus erythematosus. II. Antinuclear reaction of gamma-globulin eluted from homogenates and isolated glomeruli of kidneys from patients with lupus nephritis. *Journal of Clinical Investigation* 1967;46(4):569-79.
 37. Kurahara D, Tokuda A, Grandinetti A, et al. Ethnic differences in risk for pediatric rheumatic illness in a culturally diverse population. *J RHEUMATOL* 2002;29(2):379-83.
 38. Kuwabara N, Itoh Y, Igarashi T, et al. Autoantibodies to lens epithelium-derived growth factor/transcription co-activator P75 (LEDGF/P75) in children with chronic nonspecific complaints and with positive antinuclear antibodies. *Autoimmunity* 2009;42(6):492-6.
 39. Lehman TJ, Curd JG, Zvaifler NJ, et al. The association of antinuclear antibodies, antilymphocyte antibodies, and C4 activation among the relatives of children with systemic lupus erythematosus. *Arthritis & Rheumatism* 1982;25(5):556-61.
 40. Levcovitz H, Fletcher MA, Phillips P, et al. Segregation of lymphocyte low-molecular-weight DNA and antinuclear-antibodies in a family with systemic lupus erythematosus in first cousins. *Human Genetics* 1988;80(3):253-8.
 41. Limaye S, Carr V, Kirkpatrick P, et al. Antibodies to cyclic citrullinated peptide in patients with chronic arthritis attending an arthritis-monitoring clinic. *Journal of Clinical Rheumatology* 2005;11(3):150-2.
 42. Linnik MD, Hu JZ, Heilbrunn KR, et al. Relationship between anti-double-stranded DNA antibodies and exacerbation of renal disease in patients with systemic lupus erythematosus. *Arthritis and rheumatism* 2005;52:1129-37.
 43. Lu M-C, Hsieh S-C, Lai N-S, et al. Comparison of anti-agalactosyl IgC antibodies, rheumatoid factors, and anti-cyclic citrullinated peptide antibodies in the differential diagnosis of rheumatoid arthritis and its mimics. *Clinical and experimental rheumatology* 2007;25(5):716-21.
 44. Macarthur C, Wright JG, Srivastava R, et al. Variability in physicians' reported ordering and perceived reassurance value of diagnostic tests in children with 'growing pains'. *ARCH PEDIATR ADOLESC MED* 1996;150(10):1072-6.
 45. Maury CP, Teppo AM, Wafin F, et al. Class-specific rheumatoid factors, DR antigens, and amyloidosis in patients with rheumatoid arthritis. *ANN RHEUM DIS* 1988;47(7):546-52.
 46. Menard HA. Anti-CCP versus anti-Sa antibodies for the diagnosis of RA: Commentary. *Nature Clinical Practice Rheumatology* 2007;3(2):76-7.
 47. Milde EJ, Tonder O. Demonstration of rheumatoid factor in tissue by mixed agglutination with tissue sections. *Arthritis & Rheumatism* 1968;11(4):537-45.
 48. Mine T, Tanaka H, Ishida Y, et al. Juvenile rheumatoid arthritis manifesting in only limping due to flexion contraction of the knee. *Clinical Rheumatology* 2007;26(3):433-5.
 49. Moore TL, Bandlamudi R, Alam SM, et al. Parvovirus infection mimicking systemic lupus erythematosus in a pediatric population. *SEMIN ARTHRITIS RHEUM* 1999;28(5):314-8.
 50. Nair JR, Mewar D. Evaluation of the clinical utility of anti-CCP antibody test in daily clinical practice. 2010 p. i166.

51. Oen K, El-Gabalawy HS, Canvin JMG, et al. HLA associations of seropositive rheumatoid arthritis in a Cree and Ojibway population. *J RHEUMATOL* 1998;25(12):2319-23.
52. Orme A, Luthy KE. The case of the silent crippler. *Journal of Pediatric Health Care* 2009;23(3):180-5.
53. Pachman LM, Jayanetra P, Rothberg RM. Rheumatoid sera and soluble complexes: nitroblue tetrazolium dye test and hexose monophosphate shunt activation. *Pediatrics* 1973;52(6):823.
54. Pottel H, Wiik A, Lochter H, et al. Clinical optimization and multicenter validation of antigen-specific cut-off values on the INNO-LIA[trademark] ANA update for the detection of autoantibodies in connective tissue disorders. *Clinical and experimental rheumatology* 2004;22(5):579-88.
55. Prinz JC, Kutasi Z, Weisenseel P, et al. "Borrelia-associated early-onset morphea": A particular type of scleroderma in childhood and adolescence with high titer antinuclear antibodies? Results of a cohort analysis and presentation of three cases. *Journal of the American Academy of Dermatology* 2009;60(2):248-55.
56. Reiff A, Takei S, Sadeghi S, et al. Etanercept therapy in children with treatment-resistant uveitis. *Arthritis & Rheumatism* 2001;44(6):1411-5.
57. Sarma PK, Agrawal S, Aggarwal A, et al. Systemic lupus erythematosus with major organ involvement in a patient with systemic-onset juvenile idiopathic arthritis. *J RHEUMATOL* 2007;34(4):893-4.
58. Savolainen E, Kaipainen-Seppanen O, Kroger L, et al. Total incidence and distribution of inflammatory joint diseases in a defined population: results from the Kuopio 2000 arthritis survey. *J RHEUMATOL* 2003;30(11):2460-8.
59. Shen JY, Chen SL, Wu YX, et al. Pulmonary hypertension in systemic lupus erythematosus. *Rheumatology International* 1999;18:147-51.
60. Slater CA, Davis RB, Shmerling RH. Antinuclear antibody testing. A study of clinical utility. *Arch Int Med* 1996;156(13):1421-5.
61. Southwood TR, Petty RE, Malleon PN, et al. Psoriatic arthritis in children. *Arthritis & Rheumatism* 1989;32(8):1007-13.
62. Souza LDL, Gallinaro AL, Abdo CHN, et al. Effect of Musculoskeletal Pain on Sexuality of Male Adolescents and Adults with Juvenile Idiopathic Arthritis. *J RHEUMATOL* 2009;36(6):1337-42.
63. Szanto A, Szodoray P, Kiss E, et al. Clinical, serologic, and genetic profiles of patients with associated Sjogren's syndrome and systemic lupus erythematosus. *Human Immunology* 2006;67(11):924-30.
64. Tampona M, Brescia V, Fontana A, et al. Application of a combined protocol for rational request and utilization of antibody assays improves clinical diagnostic efficacy in autoimmune rheumatic disease. *ARCH PATHOL LAB MED* 2007;131(1):112-6.
65. Tsiakalou V, Tsangaridou E, Polioudaki H, et al. Optimized detection of circulating anti-nuclear envelope autoantibodies by immunofluorescence. *BMC Immunology* 2006;7:20.
66. Ursum J, Bos WH, van de Stadt RJ, et al. Different properties of ACPA and IgM-RF derived from a large dataset: further evidence of two distinct autoantibody systems. *Arthritis Research & Therapy* 2009;11(3):R75.
67. Valladares GC, Rus MA, Sanchez-Molina Acosta MI, et al. Measurement of anti-CCP antibodies for the diagnosis of rheumatoid arthritis [Spanish]. *Quimica Clinica* 2003;22(6):397-402.
68. van-Jaarsveld CH, ter-Borg EJ, Jacobs JW, et al. The prognostic value of the antiperinuclear factor, anti-citrullinated peptide antibodies and rheumatoid factor in early rheumatoid arthritis. *Clinical and experimental rheumatology* 1999;17:689-97.
69. Vojdani A. Antibodies as predictors of complex autoimmune diseases. *International Journal of Immunopathology and Pharmacology* 2008;21(2):267-78.
70. Waller M. Correlation between diagnosis and results of serologic tests for antiglobulin antibodies. *American Journal of Medicine* 1973;54(6):731-4.
71. Wandstrat AE, Carr-Johnson F, Branch V, et al. Autoantibody profiling to identify individuals at risk for systemic lupus erythematosus. *Journal of Autoimmunity* 2006;27(3):153-60.
72. Wijeyesinghe U, Russell AS. Outcome of high titer antinuclear antibody positivity in individuals without connective tissue disease: A 10-year follow-up. *Clinical Rheumatology* 2008;27(11):1399-402.
73. Willemze A, Ioan-Facsinay A, El-Gabalawy H. Anti-citrullinated protein antibody response associated with synovial immune deposits in a patient with suspected early rheumatoid arthritis. *J RHEUMATOL* 2008;35(11):2282-4.
74. Yachha SK, Srivastava A, Chetri K, et al. Autoimmune liver disease in children. *Journal of Gastroenterology & Hepatology* 2001;16(6):674-7.
75. Yi Z, Xin T, Zhanguo L. Prevalence and clinical significance of antibodies to citrullinated fibrinogen (ACF) in Chinese patients with rheumatoid arthritis. *Clinical Rheumatology* 2007;26(9):1505-12.
76. Zha QL, He YT, Yu JP. Correlations between diagnostic information and therapeutic efficacy in rheumatoid arthritis analyzed with decision tree

- model. Chinese journal of integrated traditional and Western medicine 2006;26:871-6.
77. Zierhut M, Doycheva D, Biester S, et al. How to manage therapy-resistant rheumatic uveitis [German]. Aktuelle Rheumatologie 2007;32(5):278-80.

Index test (N = 64)

The following studies were excluded because the index tests were not ANA, RF, and/or CCP.

1. Adib N, Silman A, Thomson W. Outcome following onset of juvenile idiopathic inflammatory arthritis: II. predictors of outcome in juvenile arthritis. *Rheumatology* 2005;44(8):1002-7.
2. Anderson CJ, Neas BR, Uchiumi T, et al. Autoantibodies to the 20-kDa ribosomal proteins: Identification, characterization, and new aspects on prevalence in systemic lupus erythematosus. *Clin Immunol* 2001;98(2):249-57.
3. Ansell BM, Holborow J, Zutshi D, et al. Comparison of three serological tests in adult rheumatoid arthritis and Still's disease (juvenile rheumatoid arthritis). *Ann N Y Acad Sci* 1969;168(1):21-9.
4. Bader-Meunier B, Armengaud JB, Haddad E, et al. Initial presentation of childhood-onset systemic lupus erythematosus: a French multicenter study. *J Pediatr* 2005;146(5):648-53.
5. Barnes MG, Grom AA, Thompson SD, et al. Subtype-specific peripheral blood gene expression profiles in recent-onset juvenile idiopathic arthritis. *Arthritis Rheum* 2009;60(7):2102-12.
6. Barnett EV, NORTH AF, Jr., CONDEMI JJ, et al. Antinuclear factors in systemic lupus erythematosus and rheumatoid arthritis. *Ann Intern Med* 1965;63:100-8.
7. Barraclough D, Russell AS, Percy JS. Diagnosis and follow-up of children referred to a rheumatic disease unit. *Med J Aust* 1977;1(25):920-3.
8. Beresford MW, Cleary G, Sills JA, et al. Delayed diagnosis: JIA in children with learning difficulties. *Rheumatology* 2005;44:292.
9. Boone MI, Moore TL, Cruz OA. Screening for uveitis in juvenile rheumatoid arthritis. *J Pediatr Ophthalmol Strabismus* 1998;35(1):41-3.
10. Calabro JJ, Parrino GR, Atchoo PD, et al. Chronic iridocyclitis in juvenile rheumatoid arthritis. *Arthritis Rheum* 1970;13(4):406-13.
11. Chen Y-S, Yang Y-H, Lin Y-T, et al. Risk of infection in hospitalised children with systemic lupus erythematosus: A 10-year follow-up. *Clin Rheumatol* 2004;23(3):235-8.
12. Chudwin DS, Ammann AJ, Cowan MJ, et al. Significance of a positive antinuclear antibody test in a pediatric population. *Am J Dis Child* 1983;137(11):1103-6.
13. Cleary AG, Lancaster GA, Annan F, et al. Nutritional impairment in juvenile idiopathic arthritis. *Rheumatology* 2004;43(12):1569-73.
14. Craft AW, Eastham EJ, Bell JI, et al. Serum ferritin in juvenile chronic polyarthritis. *Ann Rheum Dis* 1977;36(3):271-3.
15. De IJ. Epidemiology of musculoskeletal pain in primary care. *Arch Dis Child* 2004;89(5):431-4.
16. Dippell J. Hidden IgM rheumatoid factor in chronic juvenile arthritis. An open multicenter diagnostic study [German]. *Monatsschr Kinderheilkd* 1989;Organ(9):606-9.
17. Ehrmann FD, Bernatsky S, Abrahamowicz M, et al. Consultation with an arthritis specialist for children with suspected juvenile rheumatoid arthritis: a population-based study. *Arch Pediatr Adolesc Med* 2008;162(6):538-43.
18. Foster HE, Eltringham MS, Kay LJ, et al. Delay in access to appropriate care for children presenting with musculoskeletal symptoms and ultimately diagnosed with juvenile idiopathic arthritis. *Arthritis Rheum* 2007;57(6):921-7.
19. Fujikawa S, Okuni M. Clinical analysis of 570 cases with juvenile rheumatoid arthritis: results of a nationwide retrospective survey in Japan. *Acta Paediatr Jpn* 1997;39(2):245-9.

20. Goel KM, Shanks RA. Follow-up study of 100 cases of juvenile rheumatoid arthritis. *Ann Rheum Dis* 1974;33(1):25-31.
21. Griffiths B, Mosca M, Gordon C. Assessment of patients with systemic lupus erythematosus and the use of lupus disease activity indices. *Best Pract Res Clin Rheumatol* 2005;19(5 SPEC. ISS.):685-708.
22. Guillaume S, Prieur AM, Coste J, et al. Long-term outcome and prognosis in oligoarticular-onset juvenile idiopathic arthritis. *Arthritis Rheum* 2000;43(8):1858-65.
23. Heinlen LD, McClain MT, Kim X, et al. Anti-Ro and anti-nRNP response in unaffected family members of SLE patients. *Lupus* 2003;12(4):335-7.
24. Ilowite NT, Wedgwood JF, Bonagura VR. Expression of the major rheumatoid factor cross-reactive idiotype in juvenile rheumatoid arthritis. *Arthritis Rheum* 1989;32(3):265-70.
25. Jesus AA, Silva CA, Carneiro-Sampaio M, et al. Anti-C1q antibodies in juvenile-onset systemic lupus erythematosus. *Ann N Y Acad Sci* 2009;1173:235-8.
26. Kobayashi S, Wada N, Kubo M. Autoantibodies to native human type II collagen in the sera of patients with juvenile rheumatoid arthritis. *Japan J Rheumatol* 1991;3(4):265-74.
27. Konijnenberg AY, De Graeff-Meeder ER, Kimpen JL, et al. Children with unexplained chronic pain: do pediatricians agree regarding the diagnostic approach and presumed primary cause? *Pediatrics* 2004;114(5):1220-6.
28. Kornreich HK, Drexler E, Hanson V. Antinuclear factors in childhood rheumatic diseases. *J Pediatr* 1966;69(6):1039-45.
29. Kotaniemi K, Savolainen A, Karma A, et al. Recent advances in uveitis of juvenile idiopathic arthritis. *Surv Ophthalmol* 2003;48(5):489-502.
30. Kotaniemi K. Uveitis in juvenile idiopathic arthritis. *Acta Ophthalmol Scand* 2002;80(2):226.
31. Krumrey-Langkammerer M, Hafner R. Evaluation of the ILAR criteria for juvenile idiopathic arthritis. *J Rheumatol* 2001;28(11):2544-7.
32. Len CA, Terreri MT, Puccini RF, et al. Development of a tool for early referral of children and adolescents with signs and symptoms suggestive of chronic arthropathy to pediatric rheumatology centers. *Arthritis Rheum* 2006;55(3):373-7.
33. Maeno N, Takei S, Fujikawa S, et al. Antiagalactosyl IgG antibodies in juvenile idiopathic arthritis, juvenile onset Sjogren's syndrome, and healthy children. *J Rheumatol* 2004;31(6):1211-7.
34. Marini R, Costallat LT. Young age at onset, renal involvement, and arterial hypertension are of adverse prognostic significance in juvenile systemic lupus erythematosus. *Rev Rhum (English Edition)* 1999;66(6):303-9.
35. Mason T, Reed AM, Nelson AM, et al. Radiographic progression in children with polyarticular juvenile rheumatoid arthritis: a pilot study. *Ann Rheum Dis* 2005;64(3):491-3.
36. McBride JD, Gabriel FG, Fordham J, et al. Screening autoantibody profiles in systemic rheumatic disease with a diagnostic protein microarray that uses a filtration-assisted nanodot array luminometric immunoassay (NALIA). *Clin Chem* 2008;54(5):883-90.
37. Mikkelsen M, Sourander A, Salminen JJ, et al. Widespread pain and neck pain in schoolchildren. A prospective one-year follow-up study. *Acta Paediatr* 1999;88(10):1119-24.
38. Minden K, Mingels A, Niewerth M, et al. Juvenile idiopathic arthritis and uveitis: Epidemiology including data from a national database [German]. *Klin Monbl Augenheilkd* 2007;224(6):469-72.
39. Miller JJ. Immunologic abnormalities of juvenile arthritis. *Clin Orthop Relat Res* 1990;(259):23-30.
40. Moore T, Dorner RW, Zuckner J. Hidden rheumatoid factor in seronegative juvenile rheumatoid arthritis. *Ann Rheum Dis* 1974;33(3):255-7.

41. Moore TL, Dorner RW, Zuckner J. Complement-fixing hidden rheumatoid factor in children with benign rheumatoid nodules. *Arthritis Rheum* 1978;21(8):930-4.
42. Moore TL, Dorner RW, Weiss TD, et al. Hidden 19S IgM rheumatoid factor in juvenile rheumatoid arthritis. *Pediatr Res* 1980;14(10):1135-8.
43. Moore TL, Dorner RW, Osborn TG, et al. Hidden 19S IgM rheumatoid factors. *Semin Arthritis Rheum* 1988;18(1):72-5.
44. Moore TL, el-Najdawi E, Dorner RW. IgM rheumatoid factor plaque-forming cells in juvenile rheumatoid arthritis. *Arthritis Rheum* 1987;30(3):335-8.
45. Moore TL, Dorner RW, Sheridan PW, et al. Precipitation of 19S IgM rheumatoid factor-IgG circulating immune complexes in patients with juvenile arthritis by polyethylene glycol and separation by immobilized protein A. *Clin Exp Immunol* 1984;56(2):247-52.
46. Moore TL, Dorner RW, Weiss TD, et al. Specificity of hidden 19S IgM rheumatoid factor in patients with juvenile rheumatoid arthritis. *Arthritis Rheum* 1981;24(10):1283-90.
47. Moore TL, Zuckner J, Baldassare AR, Weiss TD, Dorner RW. Complement-fixing hidden rheumatoid factor in juvenile rheumatoid arthritis. *ARTHRITIS RHEUM* 1978; 21(8):935-941.
48. Nesher G, Moore TL, Grisanti MW, et al. Antiperinuclear factor in juvenile rheumatoid arthritis. *Ann Rheum Dis* 1992;51(3):350-2.
49. Neuer G, Bustin M, Michels H, et al. Autoantibodies to the chromosomal protein HMG-17 in juvenile rheumatoid arthritis. *Arthritis Rheum* 1992;35(4):472-5.
50. Oen K, Reed M, Malleson PN, et al. Radiologic outcome and its relationship to functional disability in juvenile rheumatoid arthritis. *J Rheumatol* 2003;30(4):832-40.
51. Okano T, Satoh M, Akizuki M. Immunologic tests: Anti SS-A/Ro, anti SS-B/La antibody [Japanese]. *Nippon Rinsho - Nippon Rinsho* 2005;63:Suppl-8.
52. Perilloux BC, Shetty AK, Leiva LE, et al. Antinuclear antibody (ANA) and ANA profile tests in children with autoimmune disorders: a retrospective study. *Clin Rheumatol* 2000;19(3):200-3.
53. Petty RE, Cassidy JT, Sullivan DB. Clinical correlates of antinuclear antibodies in juvenile rheumatoid arthritis. *J Pediatr* 1973;83(3):386-9.
54. Pongpanich B, Daengroongroj P. Juvenile rheumatoid arthritis: clinical characteristics in 100 Thai patients. *Clin Rheumatol* 1988;7(2):257-61.
55. Rangel L, Garralda ME, Hall A, et al. Psychiatric adjustment in chronic fatigue syndrome of childhood and in juvenile idiopathic arthritis. *Psychol Med* 2003;33(2):289-97.
56. Rask CU, Olsen EM, Elberling H, et al. Functional somatic symptoms and associated impairment in 5-7-year-old children: the Copenhagen Child Cohort 2000. *Eur J Epidemiol* 2009;24(10):625-34.
57. Rood MJ, ten CR, Van Suijlekom-Smit LWA, et al. Childhood-onset systemic lupus erythematosus. *Scand J Rheumatol* 1999;28(4):222-6.
58. Schur PH, Sandson J. Immunologic factors and clinical activity in systemic lupus erythematosus. *N Engl J Med* 1968;278(10):533-8.
59. Shapiro C, Maenz L, Hossain A, et al. Onset to first visit intervals in childhood rheumatic diseases. *J Rheumatol* 2007;34(9):1913-7.
60. Steffen C, Sanger L, Menzel J. Demonstration of antibodies to denatured type I and type II collagen in juvenile rheumatoid arthritis, Still's syndrome and controls by [14C] collagen radioimmunoassay. *Scand J Rheumatol* 1980;9(2):69-76.
61. Takei S, Maeno N, Shigemori M, et al. Clinical features of Japanese children and adolescents with systemic lupus erythematosus: results of 1980-1994 survey. *Acta Paediatr Jpn* 1997;39(2):250-6.

62. Walker SM, McCurdy DK, Shaham B, Brik R, Wietting H, Arora Y et al. High prevalence of IgA rheumatoid factor in severe polyarticular-onset juvenile rheumatoid arthritis, but not in systemic-onset or pauciarticular-onset disease. *Arthritis Rheum* 1990; 33(2):199-204.
63. Wu JF, Yang YH, Wang LC, et al. Antinucleosome antibodies correlate with the disease severity in children with systemic lupus erythematosus. *J Autoimmun* 2006;27(2):119-24.
64. Yanni G, Bresnihan B. Experience with a juvenile rheumatology clinic. *Ir J Med Sci* 1991;160(7):197-8.

Reference standard (N = 7)

The following studies did not have JIA or SLE as the reference standard.

1. Adib N, Davies K, Grahame R, et al. Joint hypermobility syndrome in childhood. A not so benign multisystem disorder? *Rheumatology* 2005;44(6):744-50.
2. Cabral DA, Petty RE, Fung M, et al. Persistent antinuclear antibodies in children without identifiable inflammatory rheumatic or autoimmune disease. *Pediatrics* 1992;89(3):441-4.
3. Freed GL, Jee S, Stein L, et al. Comparing the self-reported referral and management preferences of pediatricians and family physicians for children with juvenile rheumatoid arthritis. *J Rheumatol* 2003;30(12):2700-4.
4. Goel KM, Logan RW, Barnard WP, et al. Serum immunoglobulin and beta 1C-beta 1A globulin concentrations in juvenile rheumatoid arthritis. *Ann Rheum Dis* 1974;33(1):35-8.
5. Gulez N, Karaca NE, Aksu G, et al. Increased percentages of autoantibodies in immunoglobulin A-deficient children do not correlate with clinical manifestations. *Autoimmunity* 2009;42(1):74-9.
6. Rosenberg AM. Longitudinal analysis of a pediatric rheumatology clinic population. *J Rheumatol* 2005;32(10):1992-2001.
7. Sadi-Pooya AA, Bordbar MR. Are laboratory tests necessary in making the diagnosis of limb pains typical for growing pains in children? *Pediatr Int* 2007;49(6):833-5.

Outcome data (N = 155)

The following studies were excluded because they did not report sufficient numeric data on test result.

1. Abdwani R, Rizvi SG, El-Nour I. Childhood systemic lupus erythematosus in Sultanate of Oman: demographics and clinical analysis. *Lupus* 2008;17(7):683-6.
2. Agarwal I, Kumar TS, Ranjini K, et al. Clinical features and outcome of systemic lupus erythematosus. *Indian Pediatrics* 2009;46(8):711-5.
3. Agarwal V, Misra R, Aggarwal A. Immune complexes contain immunoglobulin A rheumatoid factor in serum and synovial fluid of patients with polyarticular juvenile rheumatoid arthritis. *Rheumatology* 2002;41(4):466-7.
4. Aggarwal A, Misra R. Juvenile chronic arthritis in India: is it different from that seen in Western countries? *Rheumatology International* 1994;14(2):53-6.
5. Aggarwal A, Misra RN. Juvenile rheumatoid arthritis in India--rarity of antinuclear antibody and uveitis. *INDIAN J PEDIATR* 1996;63(3):301-4.
6. Aggarwal A, Dabadghao S, Naik S, et al. Serum IgM rheumatoid factor by enzyme linked immunosorbent assay (ELISA) delineates a subset of patients with deforming joint disease in seronegative juvenile rheumatoid arthritis. *Rheumatology International* 1994;14(4):135-8.
7. Akinci A, Cakar N, Uncu N, et al. Keratoconjunctivitis sicca in juvenile rheumatoid arthritis. *Cornea* 2007;26(8):941-4.
8. Al-Matar MJ, Petty RE, Tucker LB, et al. The early pattern of joint involvement predicts disease progression in children with oligoarticular (pauciarticular) juvenile rheumatoid arthritis. *ARTHRITIS RHEUM* 2002;46(10):2708-15.
9. Al-Mosawi Z, Al-Hermi BE, Al-Saad KK, et al. Juvenile systemic lupus erythematosus in Bahrain. A tertiary referral center experience. *Saudi Medical Journal* 2009;30(5):667-72.
10. Al-Wahadneh AM, bu-Zeid AF, Khreisat WH. Juvenile idiopathic oligoarthritis: analysis of 42 cases in Jordan. *EAST MEDITERRANEAN HEALTH J* 2007;13(2):461-4.
11. Alsaeid K, Kamal H, Haider MZ, et al. Systemic lupus erythematosus in Kuwaiti children: organ system involvement and serological findings. *Lupus* 2004;13(8):613-7.
12. Alsaeid K, Haider MZ, Sharma PN, et al. The prevalence of human leukocyte antigen (HLA) DR/DQ/DP alleles in Kuwaiti children with oligoarticular juvenile idiopathic arthritis. *Rheumatology International* 2006;26(3):224-8.
13. Alspaugh MA, Miller JJ. A study of specificities of antinuclear antibodies in juvenile rheumatoid arthritis. *J PEDIATR* 1977;90(3):391-5.
14. Alvarez MC, Gonzalez FA, Lisbona MM, et al. Thyroid disorders and childhood rheumatic diseases [Spanish]. *Anales de Pediatría* 2009;70(1):53-6.
15. Andersson GB, Fasth A, Andersson J, et al. Incidence and prevalence of juvenile chronic arthritis: a population survey. *ANN RHEUM DIS* 1987;46(4):277-81.
16. Arguedas O, Fasth A, Andersson-Gare B. A prospective population based study on outcome of juvenile chronic arthritis in Costa Rica. *J RHEUMATOL* 2002;29(1):174-83.
17. Arguedas O, Porras O, Fasth A. Juvenile chronic arthritis in Costa Rica. A pilot referral study. *Clinical & Experimental Rheumatology* 1995;13(1):119-23.
18. Arguedas O, Fasth A, Andersson-Gare B, et al. Juvenile chronic arthritis in urban San Jose, Costa Rica: a 2 year prospective study. *J RHEUMATOL* 1998;25(9):1844-50.

19. Avcin T, Ambrozic A, Bozic B, et al. Estimation of anticardiolipin antibodies, anti-beta2 glycoprotein I antibodies and lupus anticoagulant in a prospective longitudinal study of children with juvenile idiopathic arthritis. *Clinical & Experimental Rheumatology* 2002;20(1):101-8.
20. Bahabri S, Al-Sewairi W, Al-Mazyad A, et al. Juvenile rheumatoid arthritis: The Saudi experience. *Annals of Saudi Medicine* 1997;17(4):413-8.
21. Bakr A. Epidemiology treatment and outcome of childhood systemic lupus erythematosus in Egypt. *PEDIATR NEPHROL* 2005;20(8):1081-6.
22. Balkaran BN, Roberts LA, Ramcharan J. Systemic lupus erythematosus in Trinidadian children. *Annals of Tropical Paediatrics* 2004;24(3):241-4.
23. Bardare M, De VM, Giani M, et al. Systemic lupus erythematosus in childhood: review of the literature and personal observations on 32 cases [Italian]. *Pediatrica Medica e Chirurgica* 1990;12(6):577-86.
24. Berk AT, Kocak N, Unsal E. Uveitis in juvenile arthritis. *Ocular Immunology & Inflammation* 2001;9(4):243-51.
25. Bernatsky S, Duffy C, Malleson P, et al. Economic impact of juvenile idiopathic arthritis. *ARTHRITIS RHEUM* 2007;57(1):44-8.
26. Bharadwaj A, Aggarwal A, Misra R. Clinical relevance of IgA rheumatoid factor (RF) in children with juvenile rheumatoid arthritis. *Rheumatology International* 1999;19(1-2):47-9.
27. Blockey NJ, Gibson AA, Goel KM. Monarticular juvenile rheumatoid arthritis. *Journal of Bone & Joint Surgery - British Volume* 1980;62(3):368-71.
28. Bluestone RH, Goldberg LS, Katz R, et al. Juvenile rheumatoid arthritis: a serological survey of 200 consecutive patients. *ANN RHEUM DIS* 1970;29(3):337-8.
29. Bolt IB, Cannizzaro E, Seger R, et al. Risk factors and longterm outcome of juvenile idiopathic arthritis-associated uveitis in Switzerland. *J RHEUMATOL* 2008;35(4):703-6.
30. Buoncompagni A, Barbano GC, Pistoia V, et al. Childhood systemic lupus erythematosus: A review of 30 cases. *Clinical and experimental rheumatology* 1991;9(4):425-30.
31. Burlingame RW, Rubin RL, Rosenberg AM. Autoantibodies to chromatin components in juvenile rheumatoid arthritis. *ARTHRITIS RHEUM* 1993;36(6):836-41.
32. Butbul YA, Tyrrell PN, Schneider R, et al. Comparison of patients with juvenile psoriatic arthritis and nonpsoriatic juvenile idiopathic arthritis: how different are they? *J RHEUMATOL* 2009;36(9):2033-41.
33. Cerna M, Vavrincova P, Havelka S. HLA and juvenile rheumatoid arthritis. *Acta Universitatis Carolinae - Medica* 1994;40(1-4):69-73.
34. Chacon DP, Quintero V, Chavez MEF, et al. Pediatric systemic lupus erythematosus: Characterization of 30 cases [Spanish]. *Salus* 2009;13(2):11-20.
35. Chandrasekaran AN, Rajendran CP, Madhavan R. Juvenile rheumatoid arthritis--Madras experience. *INDIAN J PEDIATR* 1996;63(4):501-10.
36. Chen C-Y, Chen L-C, Yeh K-W, et al. Sequential changes to clinical parameters and adhesion molecules following intravenous pulse cyclophosphamide and methylprednisolone treatment of refractory juvenile idiopathic arthritis. *Clinical and experimental rheumatology* 2004;22(2):259-64.
37. Crowe WE, Hug G, Chuck G. A study of the relationship of alpha₁-antitrypsin phenotype to the occurrence and severity of juvenile rheumatoid arthritis. *Arthritis and rheumatism* 1982;25(8):1010-2.
38. Cuesta IA, Kerr K, Simpson P, et al. Subspecialty referrals for pauciarticular juvenile rheumatoid arthritis. *ARCH PEDIATR ADOLESC MED* 2000;154(2):122-5.

39. Deane PM, Liard G, Siegel DM, et al. The outcome of children referred to a pediatric rheumatology clinic with a positive antinuclear antibody test but without an autoimmune disease. *Pediatrics* 1995;95(6):892-5.
40. Dolman KM, Brouwer N, Frakking FN, et al. Mannose-binding lectin deficiency is associated with early onset of polyarticular juvenile rheumatoid arthritis: a cohort study. *Arthritis Research & Therapy* 2008;10(2):R32.
41. Dracou C, Constantinidou N, Constantopoulos A. Juvenile chronic arthritis profile in Greek children. *Acta Paediatrica Japonica* 1998;40(6):558-63.
42. Edelsten C, Lee V, Bentley CR, et al. An evaluation of baseline risk factors predicting severity in juvenile idiopathic arthritis associated uveitis and other chronic anterior uveitis in early childhood. *British Journal of Ophthalmology* 2002;86(1):51-6.
43. Egeskjold EM, Hoyeraal HM, Permin H, et al. Immunoglobulins, anti-IgG antibodies and antinuclear antibodies in paired serum and synovial fluid samples. A comparison between juvenile and adult rheumatoid arthritis. *Scandinavian Journal of Rheumatology* 1985;14(1):51-7.
44. Emad Y, Ragab Y, Shaarawy A, et al. Can magnetic resonance imaging differentiate undifferentiated arthritis based on knee imaging? *J RHEUMATOL* 2009;36(9):1963-70.
45. Fantini F, Gerloni V, Gattinara M, et al. Remission in juvenile chronic arthritis: A cohort study of 683 consecutive cases with a mean 10 year followup. *J RHEUMATOL* 2003;30(3):579-84.
46. Ferraz MB, Goldenberg J, Hilario MO, et al. Evaluation of the 1982 ARA lupus criteria data set in pediatric patients. Committees of Pediatric Rheumatology of the Brazilian Society of Pediatrics and the Brazilian Society of Rheumatology. *Clinical & Experimental Rheumatology* 1994;12(1):83-7.
47. Galea P, D'amato B, Goel KM. Ocular complications in juvenile chronic arthritis (JCA). *Scottish Medical Journal* 1985;30(3):164-7.
48. Gao JS, Wu H, Tian J. Treatment of patients with juvenile rheumatoid arthritis with combination of leflunomide and methotrexate [Chinese]. *Zhonghua Erke Zazhi* 2003;41(6):435-8.
49. Gedalia A, Molina JF, Molina J, et al. Childhood-onset systemic lupus erythematosus: a comparative study of African Americans and Latin Americans. *Journal of the National Medical Association* 1999;91(9):497-501.
50. Giannini EH, Ilowite NT, Lovell DJ, et al. Long-term safety and effectiveness of etanercept in children with selected categories of juvenile idiopathic arthritis. *ARTHRITIS RHEUM* 2009;60(9):2794-804.
51. Grassi A, Corona F, Casellato A, et al. Prevalence and outcome of juvenile idiopathic arthritis-associated uveitis and relation to articular disease. *J RHEUMATOL* 2007;34(5):1139-45.
52. Griffin TA, Barnes MG, Ilowite NT, et al. Gene expression signatures in polyarticular juvenile idiopathic arthritis demonstrate disease heterogeneity and offer a molecular classification of disease subsets. *ARTHRITIS RHEUM* 2009;60(7):2113-23.
53. Gutowska-Grzegorzczak G, Baum J. Serum immunoglobulin and complement interrelationships in juvenile rheumatoid arthritis. *J RHEUMATOL* 1977;4(2):179-85.
54. Haber PL, Osborn TG, Moore TL. Antinuclear antibody in juvenile rheumatoid arthritis sera reacts with 50-40 kDa antigen(s) found in HeLa nuclear extracts. *J RHEUMATOL* 1989;16(7):949-54.
55. Han T-X. Combined examination of antikeratin antibody, antiperinuclear factor and anti-cyclic citrullinated peptide antibody in the diagnosis and evaluation of juvenile rheumatoid arthritis. [Chinese]. *Journal of Clinical Rehabilitative Tissue Engineering Research* 2007;11(27):5285-90.

56. Heiligenhaus A, Niewerth M, Ganser G, et al. Prevalence and complications of uveitis in juvenile idiopathic arthritis in a population-based nation-wide study in Germany: suggested modification of the current screening guidelines. *Rheumatology* 2007;46(6):1015-9.
57. Heinz C, Mingels A, Goebel C, et al. Chronic uveitis in children with and without juvenile idiopathic arthritis: differences in patient characteristics and clinical course. *J RHEUMATOL* 2008;35(7):1403-7.
58. Heshmat NM, El-Kerdany TH. Serum levels of vascular endothelial growth factor in children and adolescents with systemic lupus erythematosus. *Pediatric Allergy & Immunology* 2007;18(4):346-53.
59. Ilowite NT, Wedgwood JF, Moore TL, et al. Hidden rheumatoid factor and Wa idiotypic expression in juvenile rheumatoid arthritis. *Scandinavian Journal of Immunology* 1991;34(4):453-60.
60. Ince A, Akhter IM, Moore TL, et al. Hidden 19S IgM rheumatoid factors in Turkish patients with juvenile rheumatoid arthritis. *J RHEUMATOL* 1998;25(1):190-2.
61. Ingegnoli F, Del PN, Lupi E, et al. Anti-chromatin antibodies in juvenile rheumatoid arthritis [Italian]. *Reumatismo* 2003;55(4):240-4.
62. Iqbal S, Sher MR, Good RA, et al. Diversity in presenting manifestations of systemic lupus erythematosus in children. *J PEDIATR* 1999;135(4):500-5.
63. Itoh Y, Hamada H, Imai T, et al. Antinuclear antibodies in children with chronic nonspecific complaints. *Autoimmunity* 1997;25(4):243-50.
64. Jarvis JN, Pousak T, Krenz M. Detection of IgM rheumatoid factors by enzyme-linked immunosorbent assay in children with juvenile rheumatoid arthritis: correlation with articular disease and laboratory abnormalities. *Pediatrics* 1992;90(6):945-9.
65. Jurencak R, Fritzler M, Tyrrell P, et al. Autoantibodies in pediatric systemic lupus erythematosus: ethnic grouping, cluster analysis, and clinical correlations. *J RHEUMATOL* 2009;36(2):416-21.
66. Kanski JJ. Clinical and immunological study of anterior uveitis in juvenile chronic polyarthritis. *Transactions of the Ophthalmological Societies of the United Kingdom* 1976;96(1):123-30.
67. Karhulahti T, Ylijoki H, Ronning O. Mandibular condyle lesions related to age at onset and subtypes of juvenile rheumatoid arthritis in 15-year-old children. *Scandinavian Journal of Dental Research* 1993;101(5):332-8.
68. Kasapcopur O, Yologlu N, Ozyazgan Y, et al. Uveitis and anti nuclear antibody positivity in children with juvenile idiopathic arthritis. *Indian Pediatrics* 2004;41(10):1035-9.
69. Katargina LA, Khvatova AV, Krichevskaiia GI, et al. Clinical variants and immunologic features of rheumatoid uveitis in children of different age [Russian]. *Vestnik Oftalmologii* 2001;117(1):30-3.
70. Kump LI, Castaneda RA, Androudi SN, et al. Visual outcomes in children with juvenile idiopathic arthritis-associated uveitis. *Ophthalmology* 2006;113(10):1874-7.
71. Kuna AT, Lamot L, Miler M, et al. Antibodies to mutated citrullinated vimentin and antibodies to cyclic citrullinated peptides in juvenile idiopathic arthritis. *Clin Chem Lab Med* 2009;47(12):1525-30.
72. Kunnamo I, Kallio P, Pelkonen P, et al. Clinical signs and laboratory tests in the differential diagnosis of arthritis in children. *American Journal of Diseases of Children* 1987;141(1):34-40.
73. Leak AM, Ansell BM, Burman SJ. Antinuclear antibody studies in juvenile chronic arthritis. *ARCH DIS CHILD* 1986;61(2):168-72.
74. Lee DH, Daud U, Wipfl J, et al. The Decreasing Prevalence of Uveitis Associated with Juvenile Rheumatoid Arthritis: Do NSAIDs Play a Role? *Journal of Clinical Rheumatology* 2003;9(3):151-5.

75. Lehman TJ, Hanson V, Zvaifler N, et al. Antibodies to nonhistone nuclear antigens and antilymphocyte antibodies among children and adults with systemic lupus erythematosus and their relatives. *J RHEUMATOL* 1984;11(5):644-7.
76. Lepore L, Del SM, Malorgio C, et al. Treatment of juvenile idiopathic arthritis with intra-articular triamcinolone hexacetonide: evaluation of clinical effectiveness correlated with circulating ANA and T gamma/delta + and B CD5+ lymphocyte populations of synovial fluid. *Clinical & Experimental Rheumatology* 2002;20(5):719-22.
77. Lepvrier-Guibal N, Tiret A, Prieur AM, et al. Uveitis in juvenile chronic arthritis [French]. *Journal Francais d Ophthalmologie* 1994;17(8-9):489-95.
78. Lepvrier-Guibal N, Tiret A, Prieur AM, et al. Uveitis in juvenile chronic arthritis [French]. *Journal Francais d'Ophthalmologie* 1994;17(8-9):489-95.
79. Li CW, Hu J, Pi SH. Clinical characteristics of children with enthesitis related arthritis [Chinese]. *Zhonghua Erke Zazhi* 2003;41(11):835-8.
80. Lin SJ, Huang JL, Chao HC, et al. A follow-up study of systemic-onset juvenile rheumatoid arthritis in children. *Acta Paediatrica Taiwanica* 1999;40(3):176-81.
81. Machado SH, von Muhlen CA, Brenol JC, et al. The prevalence of anti-cyclic citrullinated peptide antibodies in juvenile idiopathic arthritis [Portuguese]. *Jornal de Pediatria* 2005;81(6):491-4.
82. Machado SH, von Muhlen CA, Brenol JCT, et al. The prevalence of anti-cyclic citrullinated peptide antibodies in juvenile idiopathic arthritis. *Jornal de Pediatria* 2005;81(6):491-4.
83. Magsaam J, Ferjencik P, Tempels M, et al. A new method for the detection of hidden IgM rheumatoid factor in patients with juvenile rheumatoid arthritis. *J RHEUMATOL* 1987;14(5):964-7.
84. Malleson PN, Fung MY, Petty RE, et al. Autoantibodies in chronic arthritis of childhood: relations with each other and with histocompatibility antigens. *ANN RHEUM DIS* 1992;51(12):1301-6.
85. Malleson PN, Sailer M, Mackinnon MJ. Usefulness of antinuclear antibody testing to screen for rheumatic diseases. *ARCH DIS CHILD* 1997;77(4):299-304.
86. Manzotti F, Orsoni JG, Zavota L, et al. Autoimmune uveitis in children: clinical correlation between antinuclear antibody positivity and ocular recurrences. *Rheumatology International* 2002;21(4):127-32.
87. Marks SD, Pilkington C, Woo P, et al. The use of the British Isles Lupus Assessment Group (BILAG) index as a valid tool in assessing disease activity in childhood-onset systemic lupus erythematosus. *Rheumatology* 2004;43(9):1186-9.
88. Martinez-Cairo CS, Antonio OD, Frati A. Juvenile rheumatoid arthritis. Study of 46 cases [Spanish]. *Boletin Medico del Hospital Infantil de Mexico* 1978;35(4):711-7.
89. Martinez-Cordero E, Martinez-Miranda E, Negrete-Garcia MC, et al. Autoantibodies in juvenile rheumatoid arthritis. Clinical and serologic study [Spanish]. *Boletin Medico del Hospital Infantil de Mexico* 1989;46(5):316-21.
90. Martinez-Rojano H, Juarez HE, Ladron de GG, et al. Rheumatologic manifestations of pediatric HIV infection. *AIDS PATIENT CARE STDS* 2001;15(10):519-26.
91. Matsaniotis N, Kattamis C, Paidousis M, et al. The direct coombs test in juvenile rheumatoid arthritis. *Helvetica Paediatrica Acta* 1974;29(6):609-13.
92. McCarthy PL, Wasserman D, Spiesel SZ, et al. Evaluation of arthritis and arthralgia in the pediatric patient. *CLIN PEDIATR* 1980;19(3):183-90.
93. McGhee JL, Kickingbird LM, Jarvis JN. Clinical utility of antinuclear antibody tests in children. *BMC Pediatrics* 2004;4:13.

94. McGhee JL, Burks FN, Sheckels JL, et al. Identifying children with chronic arthritis based on chief complaints: absence of predictive value for musculoskeletal pain as an indicator of rheumatic disease in children. *Pediatrics* 2002;110(2:Pt 1):9.
95. McMurray RW, Allen SH, Pepmueller PH, et al. Elevated serum prolactin levels in children with juvenile rheumatoid arthritis and antinuclear antibody seropositivity. *J RHEUMATOL* 1995;22(8):1577-80.
96. Mihailova D, Grigorova R, Vassileva B, et al. Autoimmune thyroid disorders in juvenile chronic arthritis and systemic lupus erythematosus. *Advances in Experimental Medicine & Biology* 1999;455:55-60.
97. Minden K, Niewerth M, Listing J, et al. Long-term outcome in patients with juvenile idiopathic arthritis. *ARTHRITIS RHEUM* 2002;46(9):2392-401.
98. Mo N, Amos N, Negi A, et al. The use of different ELISA kits to determine prevalence of anti-cyclic citrullinated peptide (anti-CCP) antibodies in children with juvenile idiopathic arthritis (JIA). *Rheumatology* 2008;47:II26.
99. Moe N, Rygg M. Epidemiology of juvenile chronic arthritis in northern Norway: a ten-year retrospective study. *Clinical & Experimental Rheumatology* 1998;16(1):99-101.
100. Moore TL, Osborn TG, Dorner RW. 19S IgM rheumatoid factor-7S IgG rheumatoid factor immune complexes isolated in sera of patients with juvenile rheumatoid arthritis. *Pediatric Research* 1986;20(10):977-81.
101. Moore TL, Osborn TG, Weiss TD, et al. Autoantibodies in juvenile arthritis. *SEMIN ARTHRITIS RHEUM* 1984;13(4):329-36.
102. Moore TL, Dorner RW, Alexander RL, et al. Enzyme linked (ELISA) immunoabsorbent assay for the detection of hidden 19S IgM rheumatoid factors in juvenile rheumatoid arthritis. *J RHEUMATOL* 1988;15(1):87-90.
103. Moore TL, Dorner RW, Sheridan PW, et al. Longitudinal study of the presence of hidden 19S IgM rheumatoid factor in juvenile rheumatoid arthritis. *J RHEUMATOL* 1982;9(4):599-602.
104. Moore TL, Dorner RW. Separation and characterization of complement-fixing immune complexes in juvenile rheumatoid arthritis patients. *Rheumatology International* 1986;6(2):49-52.
105. Moradinejad MH, Zamani GR, Kiani AR, et al. Clinical features of juvenile lupus erythematosus in Iranian children. *Acta Reumatologica Portuguesa* 2008;33(1):63-7.
106. Moradinejad MH. Lupus headaches in 55 childhood-onset SLE. *Iranian Journal of Pediatrics* 2007;17(2):135-9.
107. Munthe E. Anti-IgG and antinuclear antibodies in juvenile rheumatoid arthritis. *Scandinavian Journal of Rheumatology* 1972;1(4):161-70.
108. Nassberger L, Truedsson L, Svantesson H. Occurrence of autoantibodies against neutrophil granulocyte components in juvenile chronic arthritis. *Clinical & Experimental Rheumatology* 1991;9(1):79-83.
109. Oen K, Schroeder M, Jacobson K, et al. Juvenile rheumatoid arthritis in a Canadian First Nations (aboriginal) population: onset subtypes and HLA associations. *J RHEUMATOL* 1998;25(4):783-90.
110. Oen K, Tucker L, Huber AM, et al. Predictors of early inactive disease in a juvenile idiopathic arthritis cohort: results of a Canadian multicenter, prospective inception cohort study. *ARTHRITIS RHEUM* 2009;61(8):1077-86.
111. Ozdogan H, Kasapcopur O, Dede H, et al. Juvenile chronic arthritis in a Turkish population. *Clinical & Experimental Rheumatology* 1991;9(4):431-5.
112. Pagan TM, Arroyo IL. Juvenile rheumatoid arthritis in Caribbean children: a clinical characterization. *Boletin - Asociacion Medica de Puerto Rico* 1991;83(12):527-9.

113. Pedersen TK, Jensen JJ, Melsen B, et al. Resorption of the temporomandibular condylar bone according to subtypes of juvenile chronic arthritis. *J RHEUMATOL* 2001;28(9):2109-15.
114. Peralta JL, Prieur AM. Juvenile chronic arthritis with antinuclear antibodies in 136 patients [French]. *Archives Francaises de Pediatrie* 1990;47(7):497-502.
115. Pluchinotta FR, Schiavo B, Vittadello F, et al. Distinctive clinical features of pediatric systemic lupus erythematosus in three different age classes. *Lupus* 2007;16(8):550-5.
116. Pope RM, Yoshinoya S, McDuffy SJ. Detection of immune complexes and their relationship to rheumatoid factor in a variety of autoimmune disorders. *Clinical and Experimental Immunology* 1981;46(2):259-67.
117. Pratsidou-Gertsi P, Kanakoudi-Tsakalidou F, Spyropoulou M, et al. Nationwide collaborative study of HLA class II associations with distinct types of juvenile chronic arthritis (JCA) in Greece. *European Journal of Immunogenetics* 1999;26(4):299-310.
118. Ravelli A, Felici E, Magni-Manzoni S, et al. Patients with antinuclear antibody-positive juvenile idiopathic arthritis constitute a homogeneous subgroup irrespective of the course of joint disease. *ARTHRITIS RHEUM* 2005;52(3):826-32.
119. Riise OR, Handeland KS, Cvancarova M, et al. Incidence and characteristics of arthritis in Norwegian children: a population-based study. *Pediatrics* 2008;121(2):e299-e306.
120. Roberts-Thomson PJ, Shepherd K, Southwood TR, et al. Low molecular weight IgM in juvenile chronic arthritis. *ARCH DIS CHILD* 1988;63(12):1453-6.
121. Rondeel JM, van GW, van der LH, et al. Different strategies in the laboratory diagnosis of autoimmune disease: immunofluorescence, enzyme-linked immunosorbent assay or both? *Annals of Clinical Biochemistry* 1999;36(Pt 2):189-95.
122. Rosenberg AM, Romanchuk KG. Antinuclear antibodies in arthritic and nonarthritic children with uveitis. *J RHEUMATOL* 1990;17(1):60-1.
123. Rosenberg AM, Cordeiro DM. Relationship between sex and antibodies to high mobility group proteins 1 and 2 in juvenile idiopathic arthritis. *J RHEUMATOL* 2000;27(10):2489-93.
124. Rosenberg AM, Hauta SA, Prokopchuk PA, et al. Studies on associations of antinuclear antibodies with antibodies to an uveitogenic peptide of retinal S antigen in children with uveitis. *J RHEUMATOL* 1996;23(2):370-3.
125. Rossen RD, Brewer EJ, Person DA, et al. Circulating immune complexes and antinuclear antibodies in juvenile rheumatoid arthritis. *ARTHRITIS RHEUM* 1977;20e(8):1485-90.
126. Rudolf MCJ, Genel M, Tamborlane J, et al. Juvenile rheumatoid arthritis in children with diabetes mellitus. *J PEDIATR* 1981;99(4):519-24.
127. Ruperto N, Ravelli A, Levinson JE, et al. Longterm health outcomes and quality of life in American and Italian inception cohorts of patients with juvenile rheumatoid arthritis .2. Early predictors of outcome. *J RHEUMATOL* 1997;24(5):952-8.
128. Salah S, Hamshary A, Lotfy H, et al. Juvenile Idiopathic Arthritis, the Egyptian Experience. *Journal of Medical Sciences* 2009;9(2):98-102.
129. Saulsbury FT. Antibody to ribonucleoprotein in pauciarticular juvenile rheumatoid arthritis. *J RHEUMATOL* 1988;15(2):295-7.
130. Saurenmann RK, Rose JB, Tyrrell P, et al. Epidemiology of juvenile idiopathic arthritis in a multiethnic cohort: ethnicity as a risk factor. *ARTHRITIS RHEUM* 2007;56(6):1974-84.
131. Schaller J, Wedgwood RJ. Juvenile rheumatoid arthritis: a review. *Pediatrics* 1972;50(6):940-53.
132. Schaller JG, Ochs HD, Thomas ED, et al. Histocompatibility antigens in childhood-onset arthritis. *J PEDIATR* 1976;88(6):926-30.

- 133.Schwartz MM, Simpson P, Kerr KL, et al. Juvenile rheumatoid arthritis in African Americans. *J RHEUMATOL* 1997;24(9):1826-9.
- 134.Sendagorta E, Peralta J, Romero R, et al. Uveitis and idiopathic juvenile arthritis in Spain. Epidemiological and therapeutic aspects [Spanish]. *Archivos de la Sociedad Espanola de Oftalmologia* 2009;84(3):133-8.
- 135.Serra CR, Rodrigues SH, Silva NP, et al. Clinical significance of anticardiolipin antibodies in juvenile idiopathic arthritis. *Clinical & Experimental Rheumatology* 1999;17(3):375-80.
- 136.Seth V, Kabra SK, Semwal OP, et al. Clinico-immunological profile in juvenile rheumatoid arthritis--an Indian experience. *INDIAN J PEDIATR* 1996;63(3):293-300.
- 137.Sharma S, Sherry DD. Joint distribution at presentation in children with pauciarticular arthritis. *J PEDIATR* 1999;134(5):642-3.
- 138.Shin JI, Kim KH, Chun JK, et al. Prevalence and patterns of anti-nuclear antibodies in Korean children with juvenile idiopathic arthritis according to ILAR criteria. *Scandinavian Journal of Rheumatology* 2008;37(5):348-51.
- 139.Singh S, Salaria M, Kumar L, et al. Clinico-immunological profile of juvenile rheumatoid arthritis at Chandigarh. *Indian Pediatrics* 1999;36(5):449-54.
- 140.Smerdel A, Ploski R, Flato B, et al. Juvenile idiopathic arthritis (JIA) is primarily associated with HLA-DR8 but not DQ4 on the DR8-DQ4 haplotype. *ANN RHEUM DIS* 2002;61(4):354-7.
- 141.Southwood TR, Roberts-Thomson PJ, Ahern MJ, et al. Autoantibodies in patients with juvenile chronic arthritis and their immediate family relatives. *ANN RHEUM DIS* 1990;49(12):968-72.
- 142.Spadaro A, Ricciari V, Sili SA, et al. Interleukin-6 and soluble interleukin-2 receptor in juvenile chronic arthritis: correlations with clinical and laboratory parameters. *Revue du Rhumatisme (English Edition)* 1996;63(3):171-7.
- 143.Syed RH, Gilliam BE, Moore TL. Prevalence and significance of isotypes of anti-cyclic citrullinated peptide antibodies in juvenile idiopathic arthritis. *ANN RHEUM DIS* 2008;67(7):1049-51.
- 144.Thomas E, Barrett JH, Donn RP, et al. Subtyping of juvenile idiopathic arthritis using latent class analysis. *British Paediatric Rheumatology Group. ARTHRITIS RHEUM* 2000;43(7):1496-503.
- 145.Thomas E, Barrett JH, Donn RP, et al. Subtyping of juvenile idiopathic arthritis using latent class analysis. *Arthritis and rheumatism* 2000;43(7):1496-503.
- 146.Thorsteinsson L, Abrahamsen TG, Froland SS, et al. Monocyte cytotoxicity in connective tissue diseases. Correlation with disease groups. *Scandinavian Journal of Rheumatology* 1981;10(1):49-54.
- 147.Tsitsami E, Bozzola E, Magni-Manzoni S, et al. Positive family history of psoriasis does not affect the clinical expression and course of juvenile idiopathic arthritis patients with oligoarthritis. *Arthritis Care and Research* 2003;49(4):488-93.
- 148.Valverde GJ, Garrigosa PR, Pastor SF, et al. Chronic juvenile arthritis. A study of 46 patients [Spanish]. *Revista Espanola de Reumatologia* 1980;7(2):86-94.
- 149.van RM, Van SR, De KS, et al. Anti-cyclic citrullinated peptide (anti-CCP) antibodies in children with juvenile idiopathic arthritis. *J RHEUMATOL* 2003;30(4):825-8.
- 150.Walker SM, Shaham B, McCurdy DK, et al. Prevalence and concentration of IgM rheumatoid factor in polyarticular onset disease as compared to systemic or pauciarticular onset disease in active juvenile rheumatoid arthritis as measured by ELISA. *J RHEUMATOL* 1990;17(7):936-40.
- 151.Wu CJ, Huang JL, Yang MH, et al. Clinical characteristics of juvenile rheumatoid arthritis in Taiwan. *Journal of Microbiology, Immunology & Infection* 2001;34(3):211-4.
- 152.Xu M, Roberts BB, Busby BA, et al. Evaluation of multiplex antinuclear antibody assay in pediatric patients. *Laboratory Medicine* 2007;38(11):671-5.

153. Yilmaz M, Kendirli SG, Altintas DU, et al. Juvenile idiopathic arthritis profile in Turkish children. *PEDIATR INT* 2008;50(2):154-8.
154. Zak M, Fledelius H, Pedersen FK. Ocular complications and visual outcome in juvenile chronic arthritis: A 25 year follow-up study. *Acta Ophthalmologica Scandinavica* 2003;81(3):211-5.

155. Zerlin JM, Sullivan DB, Martel W. Distal interphalangeal joint abnormalities in children with polyarticular juvenile rheumatoid arthritis. *J RHEUMATOL* 1991;18(6):889-92.

Duplicates (N = 6)

The following studies were excluded because they were duplicates.

1. Agarwal I, Kumar TS, Ranjini K, et al. Clinical features and outcome of systemic lupus erythematosus. *Indian Pediatrics* 2009;46(8):711-5.
2. Al-Mendalawi MD. Juvenile systemic lupus erythematosus in Bahrain. A tertiary referral center experience. *Saudi Medical Journal* 2009;30(9):1240.
3. Brunner J, Sitzmann FC. The diagnostic value of anti-cyclic citrullinated peptide (CCP) antibodies in children with Juvenile Idiopathic Arthritis. *Clinical & Experimental Rheumatology* 2006;24(4):449-51.
4. Lipinska J, Smolewska E, Brozik H, et al. Diagnostic value of selected immunological markers in children with JIA. [Polish]. *Alergia Astma Immunologia* 10(2)(pp 75-82), 2005 Date of Publication: 2005 2005;(2):75-82.
5. Nandi M, Ganguli SK, Mondal R, et al. Clinico-serological profile of juvenile idiopathic arthritis. *Indian Pediatrics* 2009;46(7):640-1.
6. Varbanova B, Kolarov Z, Baleva M. IgA-RF as a marker of progressing of rheumatoid arthritis and juvenile idiopathic arthritis. [Bulgarian]. *Rheumatology* 10(2)(pp 49-53), 2002 Date of Publication: 2002 2002;(2):49-53.

Appendix E. Methodological Quality of Included Studies

Table E1. Methodological quality of studies evaluating an ANA test for pSLE in children with undiagnosed MSK pain

Author Year	Spectrum composition	Selection criteria	Reference standard	Test result definition	Disease progression bias	Partial verification	Differential verification	Incorporation bias	Index test execution	Test review bias	Reference standard bias	Uninterpretable results	Withdrawals
Fawcett ²⁹ 1999	No	No	Yes	Yes	U	Yes	Yes	No	Yes	Yes	Yes	Yes	Yes
Wananukul ³⁰ 2005	No	Yes	Yes	Yes	U	Yes	Yes	No	Yes	Yes	Yes	Yes	No

U = unclear

Table E2. Methodological quality of studies evaluating an ANA test for JIA in children with undiagnosed MSK pain

Author Year	Spectrum composition	Selection criteria	Reference standard	Test result definition	Disease progression bias	Partial verification	Differential verification	Incorporation bias	Index test execution	Test review bias	Reference standard bias	Uninterpretable results	Withdrawals
Fawcett ²⁹ 1999	No	No	Yes	Yes	U	Yes	Yes	No	Yes	Yes	Yes	Yes	Yes
Haynes ³¹ 1986	No	No	Yes	Yes	U	Yes	Yes	Yes	Yes	U	Yes	Yes	Yes
Jones ³² 2006	No	No	U	Yes	U	Yes	U	Yes	No	U	Yes	Yes	No
Lipinska ³³ 2008	No	No	Yes	Yes	U	Yes	Yes	Yes	U	Yes	Yes	Yes	Yes
Nordal ³⁴ 2009	No	Yes	Yes	Yes	U	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes
Osborn ³⁵ 1984	No	No	Yes	Yes	U	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes
Siamopoulou-Mavridou ³⁶ 1991	No	No	Yes	Yes	No	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes
Wakhlou ³⁷ 2003	No	No	Yes	Yes	U	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes

U = unclear

Table E3. Methodological quality of studies evaluating a RF test for pSLE in children with undiagnosed MSK pain

Author Year	Spectrum composition	Selection criteria	Reference standard	Test result definition	Disease progression bias	Partial verification	Differential verification	Incorporation bias	Index test execution	Test review bias	Reference standard bias	Uninterpretable results	Withdrawals
Hanson ³⁸ 1966	U	No	Yes	U	U	Yes	Yes	Yes	Yes	Yes	Yes	Yes	No

U = unclear

Table E4. Methodological quality of studies evaluating a RF (IgM) test for JIA in children with undiagnosed MSK pain

Author Year	Spectrum composition	Selection criteria	Reference standard	Test result definition	Disease progression bias	Partial verification	Differential verification	Incorporation bias	Index test execution	Test review bias	Reference standard bias	Uninterpretable results	Withdrawals
Andersson-Gare ³⁹ 1994	No	No	Yes	Yes	No	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes
Balogh ⁴⁰ 1980	No	No	Yes	No	No	Yes	Yes	Yes	Yes	U	Yes	U	Yes
Egeskjold ⁴¹ 1981	No	No	Yes	Yes	U	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes
Egeskjold ⁴⁹ 1982	No	No	Yes	U	No	Yes	Yes	Yes	Yes	U	U	Yes	Yes
Eichenfield ⁴⁸ 1986	Yes	No	Yes	Yes	U	Yes	Yes	Yes	U	Yes	U	Yes	U
Ferreira ⁴² 2007	No	No	Yes	Yes	No	Yes	Yes	Yes	Yes	Yes	Yes	U	Yes
Hanson ³⁸ 1966	No	No	Yes	U	U	Yes	Yes	Yes	Yes	Yes	Yes	Yes	No
Haynes ³¹ 1986	No	No	Yes	Yes	U	Yes	Yes	Yes	Yes	U	Yes	Yes	Yes
Lipinska ³³ 2008	No	No	Yes	Yes	U	Yes	Yes	Yes	U	Yes	Yes	Yes	Yes
Permin ⁴³ 1982	No	No	Yes	Yes	No	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes
Roizenblatt ⁴⁴ 1993	No	No	Yes	Yes	No	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes
Saulsbury ⁵⁰ 1990	No	No	Yes	Yes	U	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes
Siamopoulou-Mavridou ³⁶ 1991	No	No	Yes	Yes	No	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes
Taseski ⁴⁵ 1981	No	No	No	No	U	U	U	Yes	U	Yes	Yes	Yes	Yes
Varbanova ⁴⁶ 1999	No	No	Yes	Yes	U	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes
Wernick ⁴⁷ 1981	No	No	Yes	Yes	U	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes

U = unclear

Table E5. Methodological quality of studies evaluating a CCP test for JIA in children with undiagnosed MSK pain

Author Year	Spectrum composition	Selection criteria	Reference standard	Test result definition	Disease progression bias	Partial verification	Differential verification	Incorporation bias	Index test execution	Test review bias	Reference standard bias	Uninterpretable results	Withdrawals
Avcin 2002 ⁵¹	No	No	U	Yes	U	Yes	U	Yes	Yes	Yes	Yes	Yes	Yes
Brunner 2006 ⁵²	No	No	U	Yes	U	Yes	Yes	Yes	U	Yes	Yes	Yes	Yes
Ferucci 2005 ⁵³	No	Yes	Yes	Yes	No	Yes	Yes	Yes	U	Yes	Yes	Yes	Yes
Habib 2008 ⁵⁴	No	No	Yes	Yes	No	Yes	Yes	Yes	Yes	Yes	Yes	U	Yes
Kasapcopur 2004 ⁵⁵	No	U	Yes	Yes	No	Yes	Yes	Yes	No	U	Yes	Yes	Yes
Kwok 2005 ⁵⁶	No	Yes	Yes	Yes	No	Yes	Yes	Yes	Yes	U	Yes	Yes	Yes
Lipinska 2008 ³³	No	No	Yes	Yes	U	Yes	Yes	Yes	U	Yes	Yes	Yes	Yes

U = unclear

Appendix F. Evidence Tables

Study	Participants Characteristics	Index Test Characteristics
<p>Andersson-Gare, B., 1994 (39)</p> <p>Country (# centers): Sweden (NR)</p> <p>Funding: Government</p> <p>Study design: Case control (Prospective)</p> <p>Recruitment: Epidemiological survey</p>	<p>Inclusion criteria: NR</p> <p>Exclusion criteria: NR</p> <p>JIA patients $N_{\text{analyzed}}/N_{\text{enrolled}}$: 271/271</p> <p>Mean/median age (range): NR</p> <p>Female: NR</p> <p>Median time since diagnosis (range): 5.6yr (0.1-18.6yr)</p> <p>Subtype: Systemic (4.4%); Polyarticular (27.7%); Oligoarticular (36.9%); Monoarticular (24.7%); Juvenile ankylosing spondylitis (3.0%); Arthritis in connection with inflammatory bowel disease (0.4%); Juvenile psoriatic arthropathy (2.9%)</p> <p>Reference standard for JIA diagnosis: EULAR</p>	<p>IgM-RF test Assay method: Enzyme immunoassay</p> <p>Source of antigen: NR</p> <p>Manufacturer (kit type): Pharmacia Diagnostics, Uppsala, Sweden (RF IgM EIA)</p> <p>Positive cutoff: Mean of control group + 2 sd (= 7.34 AU/ml)</p> <p>IgA-RF test Assay method: Enzyme immunoassay</p> <p>Source of antigen: NR</p> <p>Manufacturer (kit type): Pharmacia Diagnostics, Uppsala, Sweden (RF IgA EIA)</p> <p>Positive cutoff: Mean of control group + 2 sd (= 3.58 AU/ml)</p>

Results (Overall)

		Reference	
		+	-
Index test	+	29	4
	-	242	126

Disease group = JIA
 Comparator = Healthy controls
 Index = IgM-RF
Sensitivity = 10.7%
Specificity = 96.9%

Overall

		Reference	
		+	-
Index test	+	26	5
	-	245	124

Disease group = JIA
 Comparator = Healthy controls
 Index = IgA-RF
Sensitivity = 9.6%
Specificity = 96.1%

By subtype

		Reference	
		+	-
Index test	+	6	4
	-	111	126

Disease group = JIA (mono- and oligoarticular)
 Comparator = Healthy controls
 Index = IgM-RF
Sensitivity = 5.1%
Specificity = 96.9%

By subtype

		Reference	
		+	-
Index test	+	21	4
	-	102	126

Disease group = JIA (polyarticular)
Comparator = Healthy controls
Index = IgM-RF
Sensitivity = 17.1%
Specificity = 96.9%

By subtype

		Reference	
		+	-
Index test	+	1	4
	-	6	126

Disease group = JIA (systemic)
Comparator = Healthy controls
Index = IgM-RF
Sensitivity = 14.3%
Specificity = 96.9%

By subtype

		Reference	
		+	-
Index test	+	10	5
	-	107	124

Disease group = JIA (monoarticular and oligoarticular)
Comparator = Healthy controls
Index = IgA-RF
Sensitivity = 8.5%
Specificity = 96.1%

By subtype

		Reference	
		+	-
Index test	+	13	5
	-	110	124

Disease group = JIA (polyarticular)
Comparator = Healthy controls
Index = IgA-RF
Sensitivity = 10.6%
Specificity = 96.1%

By subtype

		Reference	
		+	-
Index test	+	0	5
	-	7	124

Disease group = JIA (systemic)
Comparator = Healthy controls
Index = IgA-RF
Sensitivity = 0%
Specificity = 96.1%

Study	Participants Characteristics	Index Test Characteristics
<p>Avcin, T., 2002 (51)</p> <p>Country (# centers): Italy and Slovenia (4)</p> <p>Funding: Government</p> <p>Study design: Case control (Prospective)</p> <p>Recruitment: NR</p>	<p>Inclusion criteria: NR</p> <p>Exclusion criteria: NR</p> <p>JIA patients N_{analyzed}/N_{enrolled}: 109/109</p> <p>Mean age (range): 8.7yr (0.6-20.3yr)</p> <p>Female: 72.5%</p> <p>Mean time since diagnosis (range): 3.6yr (0.3-15.6yr)</p> <p>Subtype: Polyarticular (47.7%); Oligoarticular (46.8%); Systemic (5.5%)</p> <p>Reference standard for JIA diagnosis: NR</p> <p>SLE patients N_{analyzed}/N_{enrolled}: 25/25</p> <p>Mean age (range): NR</p> <p>Female: NR</p> <p>Mean/median time since diagnosis (range): NR</p> <p>Reference standard for SLE diagnosis: NR</p>	<p>CCP test Assay method: ELISA</p> <p>Source of antigen: NR</p> <p>Manufacturer (kit type): Euro-Diagnostica, Arnhem, The Netherlands (ImmunoScan RA anti-CCP test kit)</p> <p>Positive cutoff: 70 anti-CCP unit (AU)</p>

Results (Overall)

		Reference	
		+	-
Index test	+	2	0
	-	107	30

Disease group = JIA
 Comparator = Healthy controls
 Index = CCP
Sensitivity = 1.8%
Specificity = 100.0%

By subtype

		Reference	
		+	-
Index test	+	1	0
	-	51	30

Disease group = JIA (polyarticular)
 Comparator = Healthy controls
 Index = CCP
Sensitivity = 1.9%
Specificity = 100.0%

Overall

		Reference	
		+	-
Index test	+	0	0
	-	25	30

Disease group = SLE
 Comparator = Healthy controls
 Index = CCP
Sensitivity = 0.0%
Specificity = 100.0%

By subtype

		Reference	
		+	-
Index test	+	0	0
	-	6	30

Disease group = JIA (systemic)
 Comparator = Healthy controls
 Index = CCP
Sensitivity = 0%
Specificity = 100.0%

By subtype

		Reference	
		+	-
Index test	+	1	0
	-	50	30

Disease group = JIA (oligoarticular)
 Comparator = Healthy controls
 Index = CCP
Sensitivity = 2.0%
Specificity = 100.0%

Study	Participants Characteristics	Index Test Characteristics
<p>Balogh, Z., 1980 (40)</p> <p>Country (# centers): Hungary (1)</p> <p>Funding: NR</p> <p>Study design: Case control (Prospective)</p> <p>Recruitment: Consecutive patients</p>	<p>Inclusion criteria: Healthy controls: Age- and sex-matched with JIA patients</p> <p>Exclusion criteria: JIA group: Juvenile ankylosing spondylitis, psoriatic arthritis, and arthritis associated with inflammatory bowel disease patients</p> <p>JIA patients N_{analyzed}/N_{enrolled}: 46/46</p> <p>Mean age (range): 9.6yr (2.7-15.8yr)</p> <p>Female: 47.8%</p> <p>Mean/median time since diagnosis (range): NR</p> <p>Subtype: Oligoarticular (56.5%); Systemic (19.6%); Polyarticular (17.4%); Seropositive "adult-type" JCA (6.5%)</p> <p>Reference standard for JIA diagnosis: EULAR/WHO workshop, Oslo, 1978</p>	<p>ANA test Assay method: Immunofluorescence technique (Holborow and Johnson 1969)</p> <p>Source of antigen: Fluorescein-labeled anti-IgG (H+L) specific serum</p> <p>Manufacturer (kit type): Hyland (NR)</p> <p>Positive cutoff: Titer 1:10</p> <p>IgM-RF test Assay method: Latex fixation test</p> <p>Source of antigen: NR</p> <p>Manufacturer (kit type): NR</p> <p>Positive cutoff: NR</p> <p>RF test (unspecified isotype) Assay method: Waaler-Rose test</p> <p>Source of antigen: NR</p> <p>Manufacturer (kit type): NR</p> <p>Positive cutoff: NR</p>

Results (Overall)

		Reference	
		+	-
Index test	+	6	0
	-	40	10

Disease group = JIA
 Comparator = Healthy controls
 Index = ANA
Sensitivity = 13.0%
Specificity = 100.0%

Overall

		Reference	
		+	-
Index test	+	10	0
	-	36	10

Disease group = JIA
 Comparator = Healthy controls
 Index = IgM-RF (Latex)
Sensitivity = 21.7%
Specificity = 100.0%

By subtype

		Reference	
		+	-
Index test	+	7	0
	-	39	10

Disease group = JIA
 Comparator = Healthy controls
 Index = RF (Waalser-Rose)
Sensitivity = 15.2%
Specificity = 100.0%

By subtype

		Reference	
		+	-
Index test	+	2	0
	-	24	10

Disease group = JIA (oligoarticular)

Comparator = Healthy controls

Index = ANA

Sensitivity = 7.7%**Specificity** = 100.0%**By subtype**

		Reference	
		+	-
Index test	+	6	0
	-	20	10

Disease group = JIA (oligoarticular)

Comparator = Healthy controls

Index = IgM-RF (Latex)

Sensitivity = 23.1%**Specificity** = 100.0%**By subtype**

		Reference	
		+	-
Index test	+	3	0
	-	23	10

Disease group = JIA (oligoarticular)

Comparator = Healthy controls

Index = RF (Waalser-Rose)

Sensitivity = 11.5%**Specificity** = 100.0%**By subtype**

		Reference	
		+	-
Index test	+	2	0
	-	6	10

Disease group = JIA (polyarticular)

Comparator = Healthy controls

Index = ANA

Sensitivity = 25.0%**Specificity** = 100.0%**By subtype**

		Reference	
		+	-
Index test	+	0	0
	-	8	10

Disease group = JIA (polyarticular)

Comparator = Healthy controls

Index = IgM-RF (Latex)

Sensitivity = 0%**Specificity** = 100.0%**By subtype**

		Reference	
		+	-
Index test	+	0	0
	-	8	10

Disease group = JIA (polyarticular)

Comparator = Healthy controls

Index = RF (Waalser-Rose)

Sensitivity = 0%**Specificity** = 100.0%**By subtype**

		Reference	
		+	-
Index test	+	0	0
	-	9	10

Disease group = JIA (systemic)

Comparator = Healthy controls

Index = ANA

Sensitivity = 0%**Specificity** = 100.0%**By subtype**

		Reference	
		+	-
Index test	+	1	0
	-	8	10

Disease group = JIA (systemic)

Comparator = Healthy controls

Index = IgM-RF (Latex)

Sensitivity = 11.1%**Specificity** = 100.0%**By subtype**

		Reference	
		+	-
Index test	+	1	0
	-	8	10

Disease group = JIA (systemic)

Comparator = Healthy controls

Index = RF (Waalser-Rose)

Sensitivity = 11.1%**Specificity** = 100.0%

Study	Participants Characteristics	Index Test Characteristics
<p>Brunner, J., 2006 (52)</p> <p>Country (# centers): Germany (1)</p> <p>Funding: NR</p> <p>Study design: Case control (Prospective)</p> <p>Recruitment: NR</p>	<p>Inclusion criteria: ORG: With other autoimmune pathies such as Crohn's disease, reactive arthritis, diabetes mellitus type 1, uveitis, and myositis</p> <p>NRG: Undergoing interventional cardiac therapy. With non-autoimmunopathies such as infectious diseases, endocrinopathies, arthralgias, cystic fibrosis, galactosemia, hemophilia, ADD, epilepsy, Raynaud's phenomenon, osteochondroma, and fibromyalgia Healthy controls: Newborns with no known diseases</p> <p>Exclusion criteria: JIA: Written consent not obtained, bilateral anophthalmia, and insufficient blood volumes</p> <p>JIA patients N_{analyzed}/N_{enrolled}: 45/45 Mean age (range): 11.0yr (1.9-17.3yr) Female: 66.7% Mean time since diagnosis (range): 2.1yr (NR) Subtype: Polyarticular (15.6%); Systemic (6.7%); Oligoarticular (55.6%); Enthesitis-related arthritis (13.3%); Psoriatic (4.4%) Reference standard for JIA diagnosis: NR</p> <p>SLE patients N_{analyzed}/N_{enrolled}: 4/4 Mean age (range): NR Female: NR Mean time since diagnosis (range): NR Subtype: NR Reference standard for SLE diagnosis: NR</p>	<p>CCP test Assay method: ELISA</p> <p>Source of antigen: NR</p> <p>Manufacturer (kit type): Euroimmun lot 21122m, Germany (NR)</p> <p>Positive cutoff: 2.5 relative units (RU)</p>

Results (Overall)

		Reference	
		+	-
Index test	+	2	0
	-	43	42

Disease group = JIA
 Comparator = Healthy controls
 Index = CCP

Sensitivity = 4.4%
Specificity = 100%

Overall

		Reference	
		+	-
Index test	+	1	0
	-	3	42

Disease group = SLE
 Comparator = Healthy controls
 Index = CCP

Sensitivity = 25.0%
Specificity = 100%

Overall

		Reference	
		+	-
Index test	+	2	5
	-	43	29

Disease group = JIA
 Comparator = NRG
 Index = CCP

Sensitivity = 4.4%
Specificity = 85.3%

Overall

		Reference	
		+	-
Index test	+	1	5
	-	3	29

Disease group = SLE
 Comparator = NRG
 Index = CCP

Sensitivity = 25.0%
Specificity = 85.3%

Overall

		Reference	
		+	-
Index test	+	2	3
	-	43	34

Disease group = JIA
 Comparator = ORG
 Index = CCP

Sensitivity = 4.4%
Specificity = 91.9%

Overall

		Reference	
		+	-
Index test	+	1	3
	-	3	34

Disease group = SLE
 Comparator = ORG
 Index = CCP

Sensitivity = 25.0%
Specificity = 91.9%

Study	Participants Characteristics	Index Test Characteristics
Egeskjold, E.M., 1981 (41)	<p>Inclusion criteria: Healthy controls: Age- and sex-matched children without arthritis or chronic disease</p> <p>Exclusion criteria: NR</p> <p>JIA patients N_{analyzed}/N_{enrolled}: 48/48</p> <p>Mean/median age (range): NR (1.0-16.0yr)</p> <p>Female: 68.8%</p> <p>Mean/median time since diagnosis (range): NR</p> <p>Subtype: Oligoarticular (52.1%); Polyarticular (29.2%); Systemic (18.8%)</p> <p>Reference standard for JIA diagnosis: ARA</p>	<p>ANA test Assay method: Immunofluorescence technique</p> <p>Source of antigen: NR</p> <p>Manufacturer (kit type): NR</p> <p>Positive cutoff: Titer 1:18</p> <p>RF test (IgM, IgG, IgA) Assay method: IIF</p> <p>Source of antigen: Sheep red cell</p> <p>Manufacturer (kit type): DAKO (NR)</p> <p>Positive cutoff: Titer 1:9</p>

Results (Overall)

		Reference	
		+	-
Index test	+	32	2
	-	16	46

Disease group = JIA
 Comparator = Healthy controls
 Index = ANA
Sensitivity = 66.7%
Specificity = 95.8%

Overall

		Reference	
		+	-
Index test	+	42	1
	-	6	47

Disease group = JIA
 Comparator = Healthy controls
 Index = IgG-RF
Sensitivity = 87.5%
Specificity = 97.9%

Overall

		Reference	
		+	-
Index test	+	2	1
	-	46	47

Disease group = JIA
 Comparator = Healthy controls
 Index = IgM-RF
Sensitivity = 4.2%
Specificity = 97.9%

Overall

		Reference	
		+	-
Index test	+	1	0
	-	47	48

Disease group = JIA
 Comparator = Healthy controls
 Index = IgA-RF
Sensitivity = 2.1%
Specificity = 100%

By subtype

		Reference	
		+	-
Index test	+	9	1
	-	0	47

Disease group = JIA (systemic)
 Comparator = Healthy controls
 Index = IgG-RF
Sensitivity = 100%
Specificity = 97.9%

By subtype

		Reference	
		+	-
Index test	+	12	1
	-	4	47

Disease group = JIA (oligoarticular)
 Comparator = Healthy controls
 Index = IgG-RF
Sensitivity = 75.0%
Specificity = 97.9%

By subtype

		Reference	
		+	-
Index test	+	12	1
	-	2	47

Disease group = JIA (polyarticular)
 Comparator = Healthy controls
 Index = IgG-RF
Sensitivity = 85.7%
Specificity = 97.9%

Study	Participants Characteristics	Index Test Characteristics																																																																														
<p>Egeskjold, E.M., 1982 (49)</p> <p>Country (# centers): Denmark (NR)</p> <p>Funding: Government</p> <p>Study design: Case control (NR)</p> <p>Recruitment: NR</p>	<p>Inclusion criteria: NR</p> <p>Exclusion criteria: NR</p> <p>JIA patients$N_{analyzed}/N_{enrolled}$: 13/13</p> <p>Mean/median age (range): 7.1yr/ 7.0yr (2.0-15.0yr)</p> <p>Female: 69.2%</p> <p>Mean time since diagnosis (range): 3.5yr (0.6-6.4yr)</p> <p>Subtype: Oligoarticular (53.8%); Polyarticular (38.5%); Systemic (7.7%); Comorbid with chronic bilateral iridocyclitis (61.5%)</p> <p>Reference standard for JIA diagnosis: ARA</p>	<p>ANA (IgM, IgG, IgA) Assay method: IIF Source of antigen: Rat liver cryostat sections and leucocytes) Manufacturer (kit type): NR Positive cutoff: Maximum of peak 2 displacement beyond normal range</p> <p>RF test (IgM, IgG, IgA) Assay method: IIF Source of antigen: Sheep erythrocyte sensitized with rabbit IgG Manufacturer (kit type): NR Positive cutoff: Maximum of peak 2 displacement beyond normal range</p>																																																																														
<p>Results (Overall)</p> <table border="1"> <thead> <tr> <th colspan="2" rowspan="2"></th> <th colspan="2">Reference</th> </tr> <tr> <th>+</th> <th>-</th> </tr> </thead> <tbody> <tr> <th rowspan="2">Index test</th> <th>+</th> <td>8</td> <td>0</td> </tr> <tr> <th>-</th> <td>5</td> <td>3</td> </tr> </tbody> </table> <p>Disease group = JIA Comparator = Healthy controls Index = IgM-ANA Sensitivity = 61.5% Specificity = 100.0%</p> <p>Overall</p> <table border="1"> <thead> <tr> <th colspan="2" rowspan="2"></th> <th colspan="2">Reference</th> </tr> <tr> <th>+</th> <th>-</th> </tr> </thead> <tbody> <tr> <th rowspan="2">Index test</th> <th>+</th> <td>2</td> <td>0</td> </tr> <tr> <th>-</th> <td>11</td> <td>3</td> </tr> </tbody> </table> <p>Disease group = JIA Comparator = Healthy controls Index = IgM-RF Sensitivity = 15.4% Specificity = 100.0%</p>			Reference		+	-	Index test	+	8	0	-	5	3			Reference		+	-	Index test	+	2	0	-	11	3	<p>Overall</p> <table border="1"> <thead> <tr> <th colspan="2" rowspan="2"></th> <th colspan="2">Reference</th> </tr> <tr> <th>+</th> <th>-</th> </tr> </thead> <tbody> <tr> <th rowspan="2">Index test</th> <th>+</th> <td>11</td> <td>0</td> </tr> <tr> <th>-</th> <td>2</td> <td>3</td> </tr> </tbody> </table> <p>Disease group = JIA Comparator = Healthy controls Index = IgG-ANA Sensitivity = 84.676.9% Specificity = 100.0%</p> <p>Overall</p> <table border="1"> <thead> <tr> <th colspan="2" rowspan="2"></th> <th colspan="2">Reference</th> </tr> <tr> <th>+</th> <th>-</th> </tr> </thead> <tbody> <tr> <th rowspan="2">Index test</th> <th>+</th> <td>10</td> <td>0</td> </tr> <tr> <th>-</th> <td>3</td> <td>3</td> </tr> </tbody> </table> <p>Disease group = JIA Comparator = Healthy controls Index = IgG-RF Sensitivity = 76.9% Specificity = 100.0%</p>			Reference		+	-	Index test	+	11	0	-	2	3			Reference		+	-	Index test	+	10	0	-	3	3	<p>Overall</p> <table border="1"> <thead> <tr> <th colspan="2" rowspan="2"></th> <th colspan="2">Reference</th> </tr> <tr> <th>+</th> <th>-</th> </tr> </thead> <tbody> <tr> <th rowspan="2">Index test</th> <th>+</th> <td>0</td> <td>0</td> </tr> <tr> <th>-</th> <td>13</td> <td>3</td> </tr> </tbody> </table> <p>Disease group = JIA Comparator = Healthy controls Index = IgA-ANA Sensitivity = 0.0% Specificity = 100.0%</p> <p>Overall</p> <table border="1"> <thead> <tr> <th colspan="2" rowspan="2"></th> <th colspan="2">Reference</th> </tr> <tr> <th>+</th> <th>-</th> </tr> </thead> <tbody> <tr> <th rowspan="2">Index test</th> <th>+</th> <td>0</td> <td>0</td> </tr> <tr> <th>-</th> <td>13</td> <td>3</td> </tr> </tbody> </table> <p>Disease group = JIA Comparator = Healthy controls Index = IgA-RF Sensitivity = 0.0% Specificity = 100.0%</p>			Reference		+	-	Index test	+	0	0	-	13	3			Reference		+	-	Index test	+	0	0	-	13	3
			Reference																																																																													
		+	-																																																																													
Index test	+	8	0																																																																													
	-	5	3																																																																													
		Reference																																																																														
		+	-																																																																													
Index test	+	2	0																																																																													
	-	11	3																																																																													
		Reference																																																																														
		+	-																																																																													
Index test	+	11	0																																																																													
	-	2	3																																																																													
		Reference																																																																														
		+	-																																																																													
Index test	+	10	0																																																																													
	-	3	3																																																																													
		Reference																																																																														
		+	-																																																																													
Index test	+	0	0																																																																													
	-	13	3																																																																													
		Reference																																																																														
		+	-																																																																													
Index test	+	0	0																																																																													
	-	13	3																																																																													

Study	Participants Characteristics	Index Test Characteristics													
<p>Eichenfield, A.H., 1986 (48)</p> <p>Country (# centers): U.S. (1)</p> <p>Funding: NR</p> <p>Study design: Cohort (Retrospective)</p> <p>Recruitment: Consecutive patients</p>	<p>Inclusion criteria: Cohort: Children's complaints were referable to the musculoskeletal system or as being of an "autoimmune" nature. RF test results must be available and measured between Jan 1981 and Dec 1982</p> <p>Exclusion criteria: NR</p> <p>Historical patient cohort N_{analyzed}/N_{enrolled}: 437/437</p> <p>Mean/median age (range): NR</p> <p>Female: NR</p> <p>Mean/median time since diagnosis (range): NR</p> <p>Subtype: NR</p> <p>Reference standard for JIA diagnosis: ARA</p>	<p>IgM-RF test Assay method: Latex fixation test</p> <p>Source of antigen: NR</p> <p>Manufacturer (kit type): Hyland Laboratories (RA-test)</p> <p>Positive cutoff: Titer 1:80</p>													
<p>Results (Overall)</p> <table border="1"> <thead> <tr> <th colspan="2" rowspan="2"></th> <th colspan="2">Reference</th> </tr> <tr> <th>+</th> <th>-</th> </tr> </thead> <tbody> <tr> <th rowspan="2">Index test</th> <th>+</th> <td>5</td> <td>6</td> </tr> <tr> <th>-</th> <td>100</td> <td>326</td> </tr> </tbody> </table> <p>Disease group = JIA Comparator = Non-JIA children with MSK complaints Index = IgM-RF (Latex) Sensitivity = 5.0% Specificity = 98.2%</p>					Reference		+	-	Index test	+	5	6	-	100	326
		Reference													
		+	-												
Index test	+	5	6												
	-	100	326												

Study	Participants Characteristics	Index Test Characteristics
<p>Fawcett, P.T., 1999 (29)</p> <p>Country (# centers): U.S.</p> <p>Funding: Government</p> <p>Study design: Case control (Prospective)</p> <p>Recruitment: NR</p>	<p>Inclusion criteria: ORG: Underwent elective orthopedic surgical procedures</p> <p>Exclusion criteria: NR</p> <p>JIA patients N_{analyzed}/N_{enrolled}: 178/178</p> <p>Mean/median age (range): NR</p> <p>Female: NR</p> <p>Mean/median time since diagnosis (range): NR</p> <p>Subtype: Oligoarticular (49.4%); Polyarticular (38.2%); Systemic (12.4%)</p> <p>Reference standard for JIA diagnosis: ACR</p> <p>SLE patients N_{analyzed}/N_{enrolled}: 21/21</p> <p>Mean/median age (range): NR</p> <p>Female: NR</p> <p>Mean/median time since diagnosis (range): NR</p> <p>Reference standard for SLE diagnosis: ACR</p>	<p>ANA test Assay method: IIF and ELISA (Immuno Concepts, Helix, Zeus)</p> <p>Source of antigen: IIF: HEp-2 cell line . Immuno Concepts : Sm, RNP, SSA, SSB, Scl-70, Jo-1. Helix : Sm, SmRNP, SSA, SSB, Scl-70, Jo-1, ds-DNA, histones, and centromere antigens. Zeus: Sm, RNP, SSA, SSB, Scl-70, Jo-1, and ds-DNA</p> <p>Manufacturer (kit type): IIF: Antibodies Incorporated, Davis, CA. ELISA: Immuno Concepts, Sacramento/CA, Helix, Sacramento/CA, Zeus, Raritan/NJ; IIF: IFA kit; IIF: Zeiss Axioplan microscope equipped for epifluorescence with excitation filters</p> <p>Positive cutoff: IIF: Positive if a clearly discernible fluorescence pattern appears at 1:40 serum dilution.</p>

Results (Overall)

		Reference	
		+	-
Index test	+	110	4
	-	68	22

Disease group = JIA
 Comparator = ORG
 Index = ANA (IIF)
Sensitivity = 61.8%
Specificity = 84.6%

Overall

		Reference	
		+	-
Index test	+	119	7
	-	59	19

Disease group = JIA
 Comparator = ORG
 Index = ANA (ELISA – Zeus)
Sensitivity = 67.1%
Specificity = 73.1%

Overall

		Reference	
		+	-
Index test	+	20	5
	-	1	21

Disease group = SLE
 Comparator = ORG
 Index = ANA (ELISA – Helix)
Sensitivity = 95.2%
Specificity = 80.8%

By subtype

		Reference	
		+	-
Index test	+	5	0
	-	83	26

Disease group = JIA (oligoarticular)

Overall

		Reference	
		+	-
Index test	+	16	0
	-	162	26

Disease group = JIA
 Comparator = ORG
 Index = ANA (ELISA – Immuno Concepts)
Sensitivity = 9.0%
Specificity = 100.0%

Overall

		Reference	
		+	-
Index test	+	21	4
	-	0	22

Disease group = SLE
 Comparator = ORG
 Index = ANA (IIF)
Sensitivity = 100%
Specificity = 84.6%

Overall

		Reference	
		+	-
Index test	+	20	7
	-	1	19

Disease group = SLE
 Comparator = ORG
 Index = ANA (ELISA – Zeus)
Sensitivity = 95.2%
Specificity = 73%

By subtype

		Reference	
		+	-
Index test	+	33	5
	-	55	21

Disease group = JIA (oligoarticular)

Overall

		Reference	
		+	-
Index test	+	73	5
	-	105	21

Disease group = JIA
 Comparator = ORG
 Index = ANA (ELISA – Helix)
Sensitivity = 41.0%
Specificity = 80.8%

Overall

		Reference	
		+	-
Index test	+	16	0
	-	5	26

Disease group = SLE
 Comparator = ORG
 Index = ANA (ELISA – Immuno Concepts)
Sensitivity = 76.2%
Specificity = 100%

By subtype

		Reference	
		+	-
Index test	+	68	4
	-	20	22

Disease group = JIA (oligoarticular)
 Comparator = ORG
 Index = ANA (IIF)
Sensitivity = 77.3%
Specificity = 84.6%

By subtype

		Reference	
		+	-
Index test	+	56	7
	-	32	19

Disease group = JIA (oligoarticular)

Comparator = ORG
 Index = ANA (ELISA – Immuno Concepts)
Sensitivity = 5.7%
Specificity = 100%

By subtype

		Reference	
		+	-
Index test	+	14	7
	-	8	19

Disease group = JIA (systemic)
 Comparator = ORG
 Index = ANA (ELISA – Zeus)
Sensitivity = 63.6%
Specificity = 73.1%

By subtype

		Reference	
		+	-
Index test	+	49	7
	-	19	19

Disease group = JIA (polyarticular)
 Comparator = ORG
 Index = ANA (ELISA - Zeus)
Sensitivity = 72.1%
Specificity = 73.1%

By subtype

		Reference	
		+	-
Index test	+	7	5
	-	15	21

Disease group = JIA (systemic)
 Comparator = ORG
 Index = ANA (ELISA – Helix)
Sensitivity = 31.8%
Specificity = 80.8%

Comparator = ORG
 Index = ANA (ELISA – Helix)
Sensitivity = 37.5%
Specificity = 80.8%

By subtype

		Reference	
		+	-
Index test	+	10	0
	-	58	26

Disease group = JIA (polyarticular)
 Comparator = ORG
 Index = ANA (ELISA – Immuno Concepts)
Sensitivity = 14.7%
Specificity = 100%

By subtype

		Reference	
		+	-
Index test	+	4	4
	-	18	22

Disease group = JIA (systemic)
 Comparator = ORG
 Index = ANA (IIF)
Sensitivity = 18.2%
Specificity = 84.6%

By subtype

		Reference	
		+	-
Index test	+	14	7
	-	8	19

Disease group = JIA (systemic)
 Comparator = ORG
 Index = ANA (ELISA – Zeus)
Sensitivity = 63.6%
Specificity = 73.1%

Comparator = ORG
 Index = ANA (ELISA – Zeus)
Sensitivity = 63.6%
Specificity = 73.1%

By subtype

		Reference	
		+	-
Index test	+	33	5
	-	35	21

Disease group = JIA (polyarticular)
 Comparator = ORG
 Index = ANA (ELISA – Helix)
Sensitivity = 48.5%
Specificity = 80.8%

By subtype

		Reference	
		+	-
Index test	+	1	0
	-	21	26

Disease group = JIA (systemic)
 Comparator = ORG
 Index = ANA (ELISA – Immuno Concepts)
Sensitivity = 4.5%
Specificity = 100%

Study	Participants Characteristics	Index Test Characteristics
<p>Ferreira, R.A., 2007 (42)</p> <p>Country (# centers): Brazil (3)</p> <p>Funding: NR</p> <p>Study design: Case control (Prospective)</p> <p>Recruitment: Random selection</p>	<p>Inclusion criteria: NR</p> <p>Exclusion criteria: NR</p> <p>JIA patients N_{analyzed}/N_{enrolled}: 91/91</p> <p>Mean age (range): 10.5yr (2.1-22.7yr)</p> <p>Female: 64.8%</p> <p>Mean time since diagnosis (range): 5.2yr (0.2-17.3yr)</p> <p>Subtype: Polyarticular (27.5%); Oligoarticular (30.8%); Systemic (41.8%)</p> <p>Reference standard for JIA diagnosis: ACR</p>	<p>IgM-RF test Assay method: Latex fixation test</p> <p>Source of antigen: NR</p> <p>Manufacturer (kit type): RapiTex, Hoechst, Marburg, Germany (RF)</p> <p>Positive cutoff: 20 IU/ml</p> <p>IgM-RF test Assay method: ELISA</p> <p>Source of antigen: NR</p> <p>Manufacturer (kit type): Flow Laboratories, U.S. (Titertek Multiskan Plus)</p> <p>Positive cutoff: EI (absorbance of serum samples/cut off)> 1.0</p> <p>IgA-RF test Assay method: ELISA</p> <p>Source of antigen: Mouse IgG</p> <p>Manufacturer (kit type): Flow Laboratories, U.S. (Titertek Multiskan Plus)</p> <p>Positive cutoff: EI> 1.0</p>

Results (Overall)

		Reference	
		+	-
Index test	+	5	0
	-	86	45

Disease group = JIA
 Comparator = Healthy controls
 Index = IgM-RF (Latex)
Sensitivity = 5.5%

Overall

		Reference	
		+	-
Index test	+	30	3
	-	61	42

Disease group = JIA
 Comparator = Healthy controls
 Index = IgM-RF (ELISA)
Sensitivity = 33.0%

Overall

		Reference	
		+	-
Index test	+	40	7
	-	51	38

Disease group = JIA
 Comparator = Healthy controls
 Index = IgA-RF (ELISA)
Sensitivity = 44.0%

Specificity = 100.0%

By subtype

		Reference	
		+	-
Index test	+	0	0
	-	28	45

Disease group = JIA (oligoarticular)

Comparator = Healthy controls

Index = IgM-RF (Latex)

Sensitivity = 0%

Specificity = 100%

By subtype

		Reference	
		+	-
Index test	+	5	0
	-	20	45

Disease group = JIA (polyarticular)

Comparator = Healthy controls

Index = IgM-RF (Latex)

Sensitivity = 20.0%

Specificity = 100%

By subtype

		Reference	
		+	-
Index test	+	0	0
	-	38	45

Disease group = JIA (systemic)

Comparator = Healthy controls

Index = IgM-RF (Latex)

Sensitivity = 0%

Specificity = 100%

Specificity = 93.3%

By subtype

		Reference	
		+	-
Index test	+	9	3
	-	19	42

Disease group = JIA (oligoarticular)

Comparator = Healthy controls

Index = IgM-RF (ELISA)

Sensitivity = 32.1%

Specificity = 93.3%

By subtype

		Reference	
		+	-
Index test	+	13	3
	-	12	42

Disease group = (polyarticular)

Comparator = Healthy controls

Index = IgM-RF (ELISA)

Sensitivity = 52.0%

Specificity = 93.3%

By subtype

		Reference	
		+	-
Index test	+	8	3
	-	30	42

Disease group = JIA (systemic)

Comparator = Healthy controls

Index = IgM-RF (ELISA)

Sensitivity = 21.1%

Specificity = 93.3%

Specificity = 84.4%

By subtype

		Reference	
		+	-
Index test	+	13	7
	-	15	38

Disease group = JIA (oligoarticular)

Comparator = Healthy controls

Index = IgA-RF (ELISA)

Sensitivity = 46.4%

Specificity = 84.4%

By subtype

		Reference	
		+	-
Index test	+	10	7
	-	15	38

Disease group = (polyarticular)

Comparator = Healthy controls

Index = IgA-RF (ELISA)

Sensitivity = 40.0%

Specificity = 84.4%

By subtype

		Reference	
		+	-
Index test	+	17	7
	-	21	38

Disease group = JIA (systemic)

Comparator = Healthy controls

Index = IgA-RF (ELISA)

Sensitivity = 44.7%

Specificity = 84.4%

Study	Participants Characteristics	Index Test Characteristics
<p>Ferucci, E.D., 2005 (53)</p> <p>Country (# centers): U.S., throughout North America (NR)</p> <p>Funding: Government</p> <p>Study design: Case control (Prospective)</p> <p>Recruitment: Simplex patients were recruited among a local cohort in Cincinnati; multiplex patients were recruited from the National Institute of Arthritis and Musculoskeletal and Skin Diseases-supported JRA Affected Sibpair Registry. Siblings were selected randomly.</p>	<p>Inclusion criteria: Healthy controls: Children with increased risk of type I diabetes from the Diabetes and Autoimmunity Study in the Young (DAISY)</p> <p>Exclusion criteria: NR</p> <p>JIA patients N_{analyzed}/N_{enrolled}: 230/230</p> <p>Mean/median age (range): 14.7yr (NR)</p> <p>Female: 73.9%</p> <p>Mean/median time since diagnosis (range): 9.7yr (NR)</p> <p>Subtype: Polyarticular (33.5%); Oligoarticular (60.4%); Systemic (6.1%)</p> <p>Reference standard for JIA diagnosis: ACR</p>	<p>RF test (unspecified isotype) Assay method: Nephelometry</p> <p>Source of antigen: NR</p> <p>Manufacturer (kit type): Dade-Behring, Newark, DE (NR)</p> <p>Positive cutoff: 15 IU/ml</p> <p>CCP test Assay method: ELISA</p> <p>Source of antigen: NR</p> <p>Manufacturer (kit type): Diastat; Axis-Shield Diagnostics, Dundee, Scotland, UK (NR)</p> <p>Positive cutoff: 5 units/ml</p>

Results (Overall)

		Reference	
		+	-
Index test	+	25	23
	-	205	665

Disease group = JIA
 Comparator = Healthy controls
 Index = RF (unspecified isotype)
Sensitivity = 10.9%
Specificity = 96.7%

Overall

		Reference	
		+	-
Index test	+	13	4
	-	217	684

Disease group = JIA
 Comparator = Healthy controls
 Index = CCP
Sensitivity = 5.7%
Specificity = 99.4%

By subtype

		Reference	
		+	-
Index test	+	9	23
	-	130	665

Disease group = JIA (oligoarticular)
 Comparator = Healthy controls
 Index = RF (unspecified isotype)
Sensitivity = 6.5%
Specificity = 96.7%

By subtype

		Reference	
		+	-
Index test	+	3	4
	-	136	684

Disease group = JIA (oligoarticular)

Comparator = Healthy controls

Index = CCP

Sensitivity = 2.2%**Specificity** = 99.4%**By subtype**

		Reference	
		+	-
Index test	+	2	23
	-	12	665

Disease group = JIA (systemic)

Comparator = Healthy controls

Index = RF (unspecified isotype)

Sensitivity = 14.3%**Specificity** = 96.7%**By subtype**

		Reference	
		+	-
Index test	+	14	23
	-	63	665

Disease group = JIA (polyarticular)

Comparator = Healthy controls

Index = RF (unspecified isotype)

Sensitivity = 18.2%**Specificity** = 96.7%**By subtype**

		Reference	
		+	-
Index test	+	0	4
	-	14	684

Disease group = JIA (systemic)

Comparator = Healthy controls

Index = CCP

Sensitivity = 0%**Specificity** = 99.4%**By subtype**

		Reference	
		+	-
Index test	+	10	4
	-	67	684

Disease group = JIA (polyarticular)

Comparator = Healthy controls

Index = CCP

Sensitivity = 13.0%**Specificity** = 99.4%

Study	Participants Characteristics	Index Test Characteristics
<p>Habib, H.M., 2008 (54)</p> <p>Country (# centers): Egypt (1)</p> <p>Funding: NR</p> <p>Study design: Case control (Prospective)</p> <p>Recruitment: NR</p>	<p>Inclusion criteria: NR</p> <p>Exclusion criteria: NR</p> <p>JIA patients N_{analyzed}/N_{enrolled}: 68/68</p> <p>Mean age (range): 10.6yr (3.0-16.0yr)</p> <p>Female: 44.1%</p> <p>Mean time since diagnosis (range): 3.7yr (1.0-8.0yr)</p> <p>Subtype: Polyarticular (55.9%); Oligoarticular (29.4%); Systemic (14.7%)</p> <p>Reference standard for JIA diagnosis: ILAR</p> <p>SLE patients N_{analyzed}/N_{enrolled}: 14/14</p> <p>Mean age (range): 12.1yr (NR)</p> <p>Female: 92.9%</p> <p>Mean time since diagnosis (range): 3.7yr (NR)</p> <p>Reference standard for SLE diagnosis: NR</p>	<p>CCP test Assay method: ELISA</p> <p>Source of antigen: Synthetic circular peptide containing citrulline</p> <p>Manufacturer (kit type): INOVA, San Diego, U.S. (Quanta Lite™)</p> <p>Positive cutoff: 20 units/ml</p>

Results (Overall)

		Reference	
		+	-
Index test	+	14	0
	-	54	20

Disease group = JIA
 Comparator = Healthy controls
 Index = CCP
Sensitivity = 20.6%
Specificity = 100.0%

By subtype

		Reference	
		+	-
Index test	+	14	0
	-	24	20

Disease group = JIA (polyarticular)
 Comparator = Healthy controls
 Index = CCP
Sensitivity = 36.8%
Specificity = 100.0%

Overall

		Reference	
		+	-
Index test	+	0	0
	-	14	20

Disease group = SLE
 Comparator = Healthy controls
 Index = CCP
Sensitivity = 0.0%
Specificity = 100.0%

By subtype

		Reference	
		+	-
Index test	+	0	0
	-	10	20

Disease group = JIA (systemic)
 Comparator = Healthy controls
 Index = CCP
Sensitivity = 0%
Specificity = 100.0%

By subtype

		Reference	
		+	-
Index test	+	0	0
	-	20	20

Disease group = JIA (oligoarticular)
 Comparator = Healthy controls
 Index = CCP
Sensitivity = 0%
Specificity = 100.0%

Study	Participants Characteristics	Index Test Characteristics
Hanson, V., 1966 (38)	<p>Inclusion criteria: NR</p> <p>Exclusion criteria: NR</p> <p>JIA patients N_{analyzed}/N_{enrolled}: 45/58 Mean/median age (range): NR Female: NR Mean/median time since diagnosis (range): NR Subtype: NR Reference standard for JIA diagnosis: ARA</p> <p>SLE patients N_{analyzed}/N_{enrolled}: 14/14 Mean/median age (range): NR Female: 80% Mean/median time since diagnosis (range): NR Subtype: NR Reference standard for SLE diagnosis: Conformed to Cook et al, and Urbach, and also met the criteria by Bywaters (Weir et al.)</p>	<p>IgM-RF test Assay method: Latex titration test</p> <p>Source of antigen: NR</p> <p>Manufacturer (kit type): NR</p> <p>Positive cutoff: Titer 1:160</p>

Results (Overall)

		Reference	
		+	-
Index test	+	10	0
	-	35	32

Disease group = JIA
 Comparator = Healthy controls
 Index = IgM-RF (Latex)
Sensitivity = 22.2%
Specificity = 100.0%

Overall

		Reference	
		+	-
Index test	+	10	3
	-	35	30

Disease group = JIA
 Comparator = ORG (other collagen diseases)
 Index = IgM-RF (Latex)
Sensitivity = 22.2%
Specificity = 90.9%

Overall

		Reference	
		+	-
Index test	+	10	1
	-	35	22

Disease group = JIA
 Comparator = NRG (ulcerative colitis)
 Index = IgM-RF (Latex)
Sensitivity = 22.2%
Specificity = 95.7%

Overall

		Reference	
		+	-
Index test	+	4	0
	-	10	32

Disease group = SLE
 Comparator = Healthy controls
 Index = IgM-RF (Latex)
Sensitivity = 28.6%
Specificity = 100.0%

Overall

		Reference	
		+	-
Index test	+	4	3
	-	10	30

Disease group = SLE
 Comparator = ORG (other collagen diseases)
 Index = IgM-RF (Latex)
Sensitivity = 28.6%
Specificity = 90.9%

Overall

		Reference	
		+	-
Index test	+	4	1
	-	10	22

Disease group = SLE
 Comparator = NRG (ulcerative colitis)
 Index = IgM-RF (Latex)
Sensitivity = 28.6%
Specificity = 95.7%

Study	Participants Characteristics	Index Test Characteristics
<p>Haynes, D.C., 1986 (31)</p> <p>Country (# centers): U.S. (1)</p> <p>Funding: Government and non-commercial institution (NIH and House of St. Giles the Cripple)</p> <p>Study design: Case control (Prospective)</p> <p>Recruitment: NR</p>	<p>Inclusion criteria: NRG: Children with unspecified nonrheumatic disease. Age-matched with JIA patients</p> <p>Exclusion criteria: NR</p> <p>JIA patients N_{analyzed}/N_{enrolled}: 65/65</p> <p>Mean/median age (range): NR (3.0-19.0yr)</p> <p>Female: NR</p> <p>Mean/median time since diagnosis (range): NR</p> <p>Subtype: Oligoarticular (35.3%); polyarticular (36.9%); systemic (27.7%)</p> <p>Reference standard for JIA diagnosis: ARA</p>	<p>ANA test Assay method: IIF Source of antigen: HEp-2 cell line Manufacturer (kit type): Breit Laboratories, Inc, Sacramento, CA (NR) Positive cutoff: A reading of 1+ or greater in fluorescence at 1:20 dilution</p> <p>IgM-RF test Assay method: ELISA Source of antigen: Human IgG Manufacturer (kit type): Sigma Chemical Co. (NR); Tago, Inc. (NR); and Pharmacia Fine Chemicals (NR) Positive cutoff: Mean of control group + 2 sd</p> <p>IgG-RF test Assay method: ELISA Source of antigen: Rabbit IgG Manufacturer (kit type): Sigma Chemical Co. (NR); Cappel Labs., Cochranville, PA (NR) Positive cutoff: Mean of control group + 2 sd</p>

Results (Overall)

		Reference	
		+	-
Index test	+	33	1
	-	32	17

Disease group = JIA
 Comparator = NRG (Unspecified)
 Index = ANA
Sensitivity = 50.8%
Specificity = 94.4%

Overall

		Reference	
		+	-
Index test	+	23	0
	-	42	20

Disease group = JIA
 Comparator = NRG (Unspecified)
 Index = IgM-RF
Sensitivity = 33.8%
Specificity = 100.0%

Overall

		Reference	
		+	-
Index test	+	4	0
	-	61	20

Disease group = JIA
 Comparator = NRG (Unspecified)
 Index = IgG-RF
Sensitivity = 6.2%
Specificity = 94.4%

By subtype

		Reference	
		+	-
Index test	+	18	1
	-	6	17

Disease group = JIA (oligoarticular)

Comparator = NRG (Unspecified)

Index = ANA

Sensitivity = 75.0%**Specificity** = 94.4%**By subtype**

		Reference	
		+	-
Index test	+	5	0
	-	19	20

Disease group = JIA (oligoarticular)

Comparator = NRG (Unspecified)

Index = IgM-RF

Sensitivity = 20.8%**Specificity** = 100.0%**By subtype**

		Reference	
		+	-
Index test	+	1	0
	-	23	20

Disease group = JIA (oligoarticular)

Comparator = NRG (Unspecified)

Index = IgG-RF

Sensitivity = 4.2%**Specificity** = 100.0%**By subtype**

		Reference	
		+	-
Index test	+	12	1
	-	11	17

Disease group = JIA (polyarticular)

Comparator = NRG (Unspecified)

Index = ANA

Sensitivity = 52.2%**Specificity** = 94.4%**By subtype**

		Reference	
		+	-
Index test	+	10	0
	-	13	20

Disease group = JIA (polyarticular)

Comparator = NRG (Unspecified)

Index = IgM-RF

Sensitivity = 43.5%**Specificity** = 100.0%**By subtype**

		Reference	
		+	-
Index test	+	2	0
	-	21	20

Disease group = JIA (polyarticular)

Comparator = NRG (Unspecified)

Index = IgG-RF

Sensitivity = 8.7%**Specificity** = 100.0%**By subtype**

		Reference	
		+	-
Index test	+	3	1
	-	15	17

Disease group = JIA (systemic)

Comparator = NRG (Unspecified)

Index = ANA

Sensitivity = 16.7%**Specificity** = 94.4%**By subtype**

		Reference	
		+	-
Index test	+	8	0
	-	10	20

Disease group = JIA (systemic)

Comparator = NRG (Unspecified)

Index = IgM-RF

Sensitivity = 44.4%**Specificity** = 100.0%**By subtype**

		Reference	
		+	-
Index test	+	1	0
	-	17	20

Disease group = JIA (systemic)

Comparator = NRG (Unspecified)

Index = IgG-RF

Sensitivity = 5.6%**Specificity** = 100.0%

Study	Participants Characteristics	Index Test Characteristics
<p>Jones, O.Y., 2006 (32)</p> <p>Country (# centers): More than one countries from North America (7)</p> <p>Funding: NR</p> <p>Study design: Case control (Retrospective)</p> <p>Recruitment: Random selection</p>	<p>Inclusion criteria: NRG: Records available in the Pediatric Rheumatology Disease Registry (1992-1995)</p> <p>Exclusion criteria: NR</p> <p>JIA patients N_{analyzed}/N_{enrolled}: 194/206</p> <p>Mean age (range): 6.4yr (2.4-17.1yr)</p> <p>Female: 75.2%</p> <p>Mean/median time since diagnosis (range): NR</p> <p>Subtype: Oligoarticular (41.8%); Polyarticular (38.4%); Systemic (19.9%)</p> <p>Reference standard for JIA diagnosis: NR</p>	<p>ANA test</p> <p>Assay method: NR</p> <p>Source of antigen: NR</p> <p>Manufacturer (kit type): NR</p> <p>Positive cutoff: Titer 1:80</p>

Results (Overall)

		Reference	
		+	-
Index test	+	66	6
	-	128	28

Disease group = JIA
 Comparator = NRG (Acute lymphocytic leukemia)
 Index = ANA
Sensitivity = 34.0%
Specificity = 82.4%

Study	Participants Characteristics	Index Test Characteristics																										
<p>Kasapcopur, O., 2004 (55)</p> <p>Country (# centers): Turkey (1)</p> <p>Funding: Academic institution</p> <p>Study design: Case control (Prospective)</p> <p>Recruitment: Consecutive patients</p>	<p>Inclusion criteria: NR</p> <p>Exclusion criteria: NR</p> <p>JIA patients N_{analyzed}/N_{enrolled}: 122/122 Mean age (range): 8.8yr (1.2-19.3yr) Female: 59.0% Mean time since diagnosis (range): 4.3yr (NR) Subtype: Polyarticular (39.3%); Oligoarticular (29.5%); Systemic (23.0%); Entesitis-related arthritis (5.7%); Juvenile psoriatic arthritis (2.5%) Reference standard for JIA diagnosis: ILAR</p> <p>SLE patients N_{analyzed}/N_{enrolled}: 19/19 Mean age (range): 13.6yr (5.3-18.0yr) Female: 89.5% Mean time since diagnosis (range): 4.5yr (NR) Reference standard for SLE diagnosis: NR</p>	<p>CCP test Assay method: ELISA</p> <p>Source of antigen: NR</p> <p>Manufacturer (kit type): Euroimmun, Germany (NR)</p> <p>Positive cutoff: 5 relative units</p>																										
<hr/>																												
<p>Results (Overall)</p> <table border="1"> <thead> <tr> <th colspan="2" rowspan="2"></th> <th colspan="2">Reference</th> </tr> <tr> <th>+</th> <th>-</th> </tr> </thead> <tbody> <tr> <th rowspan="2">Index test</th> <th>+</th> <td>3</td> <td>0</td> </tr> <tr> <th>-</th> <td>119</td> <td>15</td> </tr> </tbody> </table> <p>Disease group = JIA Comparator = Healthy controls Index = CCP Sensitivity = 2.5% Specificity = 100.0%</p>			Reference		+	-	Index test	+	3	0	-	119	15	<p>Overall</p> <table border="1"> <thead> <tr> <th colspan="2" rowspan="2"></th> <th colspan="2">Reference</th> </tr> <tr> <th>+</th> <th>-</th> </tr> </thead> <tbody> <tr> <th rowspan="2">Index test</th> <th>+</th> <td>0</td> <td>0</td> </tr> <tr> <th>-</th> <td>19</td> <td>15</td> </tr> </tbody> </table> <p>Disease group = SLE Comparator = Healthy controls Index = CCP Sensitivity = 0.0% Specificity = 100.0%</p>			Reference		+	-	Index test	+	0	0	-	19	15	
			Reference																									
		+	-																									
Index test	+	3	0																									
	-	119	15																									
		Reference																										
		+	-																									
Index test	+	0	0																									
	-	19	15																									

Study	Participants Characteristics	Index Test Characteristics
<p>Kwok, J.S.Y., 2005 (56)</p> <p>Country (# centers): Hong Kong (1)</p> <p>Funding: Academic institution</p> <p>Study design: Case control (Retrospective)</p> <p>Recruitment: NR</p>	<p>Inclusion criteria: NR</p> <p>Exclusion criteria: NR</p> <p>JIA patients N_{analyzed}/N_{enrolled}: 59/59 Mean age (range): 15.0yr (2.4-24.0yr) Female: 40.7% Mean time since diagnosis (range): 4.3yr (0.4-16.0yr) Subtype: Oligoarticular (25.4%); Polyarticular (32.2%); Systemic (6.8%); Enthesitis-related (20.3%); Other arthritides (15.3%) Reference standard for JIA diagnosis: ILAR</p> <p>SLE patients N_{analyzed}/N_{enrolled}: 21/21 Mean age (range): NR Female: NR Mean time since diagnosis (range): NR Reference standard for SLE diagnosis: NR</p>	<p>CCP test Assay method: ELISA</p> <p>Source of antigen: NR</p> <p>Manufacturer (kit type): Inova Diagnostics Inc, San Diego, CA; QuantaLite CCP IgG ELISA kit; NR</p> <p>Positive cutoff: 20 AU</p>

Results (Overall)

		Reference	
		+	-
Index test	+	6	1
	-	53	25

Disease group = JIA
 Comparator = NRG (allergy, idiopathic thrombocytopenia, and hepatitis C)
 Index = CCP
Sensitivity = 10.2%
Specificity = 97.9%

Overall

		Reference	
		+	-
Index test	+	0	1
	-	21	25

Disease group = SLE
 Comparator = NRG (allergy, idiopathic thrombocytopenia, and hepatitis C)
 Index = CCP
Sensitivity = 0.0%
Specificity = 96.2%

By subtype

		Reference	
		+	-
Index test	+	1	1
	-	14	25

Disease group = JIA (oligoarticular)
 Comparator = NRG (allergy, idiopathic thrombocytopenia, and hepatitis C)
 Index = CCP
Sensitivity = 6.7%
Specificity = 97.9%

By subtype

		Reference	
		+	-
Index test	+	4	1
	-	15	25

Disease group = JIA (polyarticular)
 Comparator = NRG (allergy, idiopathic
 thrombocytopenia, and hepatitis C)
 Index = CCP

Sensitivity = 21.1%

Specificity = 97.9%

By subtype

		Reference	
		+	-
Index test	+	1	1
	-	3	25

Disease group = JIA (systemic)
 Comparator = NRG (allergy, idiopathic
 thrombocytopenia, and hepatitis C)
 Index = CCP

Sensitivity = 25.0%

Specificity = 97.9%

Study	Participants Characteristics	Index Test Characteristics
<p>Lipinska, J., 2008 (33)</p> <p>Country (# centers): Poland (NR)</p> <p>Funding: Academic institution</p> <p>Study design: Case control (Prospective)</p> <p>Recruitment: NR</p>	<p>Inclusion criteria: NRG: With functional cardio-vascular system dysfunction. Age- and sex-matched with JIA patients</p> <p>Exclusion criteria: NR</p> <p>JIA patients N_{analyzed}/N_{enrolled}: 96/96</p> <p>Mean/median age (range): 12.8yr (3.0-18.0yr)</p> <p>Female: 61.5%</p> <p>Mean/median time since diagnosis (range): NR</p> <p>Subtype: Oligoarticular (37.5%); Polyarticular (51.0%); Systemic (11.5%)</p> <p>Reference standard for JIA diagnosis: ILAR</p>	<p>ANA test Assay method: IIF</p> <p>Source of antigen: NR</p> <p>Manufacturer (kit type): Euroimmun Polska Sp. z o.o. (NR)</p> <p>Positive cutoff: Titer 1:320</p> <p>IgM-RF test Assay method: ELISA</p> <p>Source of antigen: NR</p> <p>Manufacturer (kit type): Biomedica Poland Sp. Z.o.o. (05-500 Piaseczno ELISA kit)</p> <p>Positive cutoff: 24 RU/ml</p> <p>CCP test Assay method: ELISA</p> <p>Source of antigen: NR</p> <p>Manufacturer (kit type): Euroimmun Polska Sp. z o.o. (52-219 Wroclaw)</p> <p>Positive cutoff: 5 RU/ml</p>

Results (Overall)

		Reference	
		+	-
Index test	+	8	0
	-	88	22

Disease group = JIA
 Comparator = NRG
 Index = ANA
Sensitivity = 8.3%
Specificity = 100.0%

Overall

		Reference	
		+	-
Index test	+	14	1
	-	82	21

Disease group = JIA
 Comparator = NRG
 Index = IgM-RF
Sensitivity = 14.6%
Specificity = 95.5%

Overall

		Reference	
		+	-
Index test	+	40	0
	-	56	22

Disease group = JIA
 Comparator = NRG
 Index = CCP
Sensitivity = 41.7%
Specificity = 100.0%

By subtype

		Reference	
		+	-
Index test	+	2	1
	-	34	21

Disease group = JIA (oligoarticular)

Comparator = NRG

Index = IgM-RF

Sensitivity = 5.6%**Specificity** = 95.5%**By subtype**

		Reference	
		+	-
Index test	+	13	0
	-	23	22

Disease group = JIA (oligoarticular)

Comparator = NRG

Index = CCP

Sensitivity = 36.1%**Specificity** = 100%**By subtype**

		Reference	
		+	-
Index test	+	11	1
	-	38	21

Disease group = JIA (polyarticular)

Comparator = NRG

Index = IgM-RF

Sensitivity = 22.4%**Specificity** = 95.5%**By subtype**

		Reference	
		+	-
Index test	+	21	0
	-	28	22

Disease group = JIA (polyarticular)

Comparator = NRG

Index = CCP

Sensitivity = 42.9%**Specificity** = 100%**By subtype**

		Reference	
		+	-
Index test	+	1	1
	-	10	21

Disease group = JIA (systemic)

Comparator = NRG

Index = IgM-RF

Sensitivity = 9.1%**Specificity** = 95.5%**By subtype**

		Reference	
		+	-
Index test	+	6	0
	-	5	22

Disease group = JIA (systemic)

Comparator = NRG

Index = CCP

Sensitivity = 54.5%**Specificity** = 100%

Study	Participants Characteristics	Index Test Characteristics																																																				
<p>Nordal, E.B., 2009 (34)</p> <p>Country (# centers): Norway (5)</p> <p>Funding: Non-commercial institution</p> <p>Study design: Case control (Prospective)</p> <p>Recruitment: Consecutive patients</p>	<p>Inclusion criteria: JIA patients: Newly diagnosed cases NRG: Children undergoing elective outpatient procedures with no diagnosis of inflammatory diseases</p> <p>Exclusion criteria: NR</p> <p>JIA patients $N_{\text{analyzed}}/N_{\text{enrolled}}$: 100/174 Mean/median age (range): NR Female: 71.0% Mean/median time since diagnosis (range): NR (max: 1.0yr) Subtype: Oligoarticular (50.0%) Reference standard for JIA diagnosis: ILAR</p>	<p>ANA test Assay method: IIF Source of antigen: HEp-2 cell line Manufacturer (kit type): Immunoconcepts, Sacramento, CA (NR) Positive cutoff: Titer 1:80</p> <p>ANA test Assay method: ELISA Source of antigen: Recombinant or purified native nuclear antigens Manufacturer (kit type): Pharmacia Diagnostics, Freiburg, Germany (Varelisa Recombi ANA screening test). Positive cutoff: Titer 1:101</p>																																																				
<p>Results (Overall)</p> <table border="1"> <thead> <tr> <th colspan="2" rowspan="2"></th> <th colspan="2">Reference</th> </tr> <tr> <th>+</th> <th>-</th> </tr> </thead> <tbody> <tr> <th rowspan="2">Index test</th> <th>+</th> <td>57</td> <td>3</td> </tr> <tr> <th>-</th> <td>43</td> <td>55</td> </tr> </tbody> </table> <p>Disease group = JIA Comparator = NRG (no inflammatory diseases; undergoing elective surgery) Index = ANA (IIF) Sensitivity = 57.0% Specificity = 94.8%</p> <p>By comorbidity</p> <table border="1"> <thead> <tr> <th colspan="2" rowspan="2"></th> <th colspan="2">Reference</th> </tr> <tr> <th>+</th> <th>-</th> </tr> </thead> <tbody> <tr> <th rowspan="2">Index test</th> <th>+</th> <td>44</td> <td>N/A</td> </tr> <tr> <th>-</th> <td>40</td> <td>N/A</td> </tr> </tbody> </table> <p>Disease group = JIA (without uveitis) Comparator = NRG (no inflammatory diseases; undergoing elective surgery) Index = ANA (IIF) Sensitivity = 52.4% Specificity = N/A</p>			Reference		+	-	Index test	+	57	3	-	43	55			Reference		+	-	Index test	+	44	N/A	-	40	N/A	<p>Overall</p> <table border="1"> <thead> <tr> <th colspan="2" rowspan="2"></th> <th colspan="2">Reference</th> </tr> <tr> <th>+</th> <th>-</th> </tr> </thead> <tbody> <tr> <th rowspan="2">Index test</th> <th>+</th> <td>4</td> <td>0</td> </tr> <tr> <th>-</th> <td>96</td> <td>58</td> </tr> </tbody> </table> <p>Disease group = JIA Comparator = NRG (no inflammatory diseases; undergoing elective surgery) Index = ANA (ELISA) Sensitivity = 4.0% Specificity = 100.0%</p>			Reference		+	-	Index test	+	4	0	-	96	58	<p>By comorbidity</p> <table border="1"> <thead> <tr> <th colspan="2" rowspan="2"></th> <th colspan="2">Reference</th> </tr> <tr> <th>+</th> <th>-</th> </tr> </thead> <tbody> <tr> <th rowspan="2">Index test</th> <th>+</th> <td>13</td> <td>N/A</td> </tr> <tr> <th>-</th> <td>3</td> <td>N/A</td> </tr> </tbody> </table> <p>Disease group = JIA (with uveitis) Comparator = NRG (no inflammatory diseases; undergoing elective surgery) Index = ANA (IIF) Sensitivity = 81.3% Specificity = N/A</p>			Reference		+	-	Index test	+	13	N/A	-	3	N/A
			Reference																																																			
		+	-																																																			
Index test	+	57	3																																																			
	-	43	55																																																			
		Reference																																																				
		+	-																																																			
Index test	+	44	N/A																																																			
	-	40	N/A																																																			
		Reference																																																				
		+	-																																																			
Index test	+	4	0																																																			
	-	96	58																																																			
		Reference																																																				
		+	-																																																			
Index test	+	13	N/A																																																			
	-	3	N/A																																																			

Study	Participants Characteristics	Index Test Characteristics
Osborn, T.G., 1984 (35)	Inclusion criteria: NR	ANA test Assay method: IIF
Country (# centers): U.S. (1)	Exclusion criteria: NR	Source of antigen: HEp-2 cell line
Funding: Government	JIA patients N_{analyzed}/N_{enrolled}: 217/217	Manufacturer (kit type): NR
Study design: Case control (Prospective)	Mean/median age (range): NR (0.2-16.0yr)	Positive cutoff: Titer 1:40
Recruitment: NR	Female: 66.4%	
	Mean/median time since diagnosis (range): NR	
	Subtype: Oligoarticular (46.5%); Systemic (12.9%); Polyarticular (40.6%)	
	Reference standard for JIA diagnosis: ARA	

Results (Overall)

		Reference	
		+	-
Index test	+	131	3
	-	86	32

Disease group = JIA
 Comparator = Healthy controls
 Index = ANA
Sensitivity = 60.4%
Specificity = 91.4%

Overall

		Reference	
		+	-
Index test	+	131	15
	-	86	16

Disease group = JIA
 Comparator = ORG (other connective tissue diseases)
 Index = ANA
Sensitivity = 60.4%
Specificity = 51.6%

By subtype

		Reference	
		+	-
Index test	+	63	3
	-	38	32

Disease group = JIA (oligoarticular)
 Comparator = Healthy controls
 Index = ANA
Sensitivity = 62.4%
Specificity = 91.4%

By subtype

		Reference	
		+	-
Index test	+	59	3
	-	29	32

Disease group = JIA (polyarticular)

Comparator = Healthy controls

Index = ANA

Sensitivity = 67.0%**Specificity** = 91.4%**By subtype**

		Reference	
		+	-
Index test	+	59	15
	-	29	16

Disease group = JIA (polyarticular)

Comparator = ORG (other connective tissue diseases)

Index = ANA

Sensitivity = 67.0%**Specificity** = 51.6%**By subtype**

		Reference	
		+	-
Index test	+	9	3
	-	19	32

Disease group = JIA (systemic)

Comparator = Healthy controls

Index = ANA

Sensitivity = 32.1%**Specificity** = 91.4%**By subtype**

		Reference	
		+	-
Index test	+	9	15
	-	19	16

Disease group = JIA (systemic)

Comparator = ORG (other connective tissue diseases)

Index = ANA

Sensitivity = 32.1%**Specificity** = 51.6%**By subtype**

		Reference	
		+	-
Index test	+	63	15
	-	38	16

Disease group = JIA (oligoarticular)

Comparator = ORG (other connective tissue diseases)

Index = ANA

Sensitivity = 62.4%**Specificity** = 51.6%

Study	Participants Characteristics	Index Test Characteristics
Permin, H., 1982 (43)	<p>Inclusion criteria: NR</p> <p>Exclusion criteria: NR</p> <p>JIA patients N_{analyzed}/N_{enrolled}: 26/26</p> <p>Mean age (range): 9.0yr (2.0-16.0yr)</p> <p>Female: 73.1%</p> <p>Mean time since diagnosis (range): 3.yr (0.5-15.0yr)</p> <p>Subtype: Polyarticular (42.3%); Oligoarticular (42.3%); Systemic (15.4%)</p> <p>Reference standard for JIA diagnosis: ARA</p>	<p>IgM-RF test Assay method: IIF Source of antigen: Formalin-fixed sheep red cell IgG Manufacturer (kit type): NR Positive cutoff: Titer 1:10</p> <p>IgG-RF test Assay method: IIF Source of antigen: Formalin-fixed sheep red cell IgG Manufacturer (kit type): NR Positive cutoff: Titer 1:10</p> <p>IgA-RF test Assay method: IIF Source of antigen: Formalin-fixed sheep red cell IgG Manufacturer (kit type): NR Positive cutoff: Titer 1:10</p> <p>IgE-RF test Assay method: IIF Source of antigen: Formalin-fixed sheep red cell IgG Manufacturer (kit type): NR Positive cutoff: Titer 1:16</p>

Results (Overall)

		Reference	
		+	-
Index test	+	1	0
	-	25	23

Disease group = JIA
 Comparator = Healthy controls
 Index = IgM-RF
Sensitivity = 3.9%
Specificity = 100.0%

Overall

		Reference	
		+	-
Index test	+	22	0
	-	4	23

Disease group = JIA
 Comparator = Healthy controls
 Index = IgG-RF
Sensitivity = 84.6%
Specificity = 100.0%

Overall

		Reference	
		+	-
Index test	+	0	0
	-	26	23

Disease group = JIA
 Comparator = Healthy controls
 Index = IgA-RF
Sensitivity = 0%
Specificity = 100.0%

Overall

		Reference	
		+	-
Index test	+	0	0
	-	26	23

Disease group = JIA

Comparator = Healthy controls

Index = IgE-RF

Sensitivity = 0%

Specificity = 100.%

Study	Participants Characteristics	Index Test Characteristics
Roizenblatt, S., 1993 (44) Country (# centers): Brazil (1) Funding: NR Study design: Case control (Prospective) Recruitment: NR	Inclusion criteria: ORG: Hypermobile children. Matched age and sex. Exclusion criteria: NR JIA patients $N_{analyzed}/N_{enrolled}$: 24/24 Mean age (range): 9.0yr (2.3-15.0yr) Female: 54.2% Mean/median time since diagnosis (range): NR (Min: 0.5yr) Subtype: Polyarticular (37.5%); Oligoarticular (50.0%); Systemic (12.5%) Reference standard for JIA diagnosis: ACR	IgM-RF test Assay method: ELISA Source of antigen: Sheep erythrocyte Manufacturer (kit type): Dako Ab (NR) Positive cutoff: Mean of control group + 2 sd IgG-RF test Assay method: ELISA Source of antigen: NR Manufacturer (kit type): NR Positive cutoff: Mean of control group + 2 sd

Results (Overall)

		Reference	
		+	-
Index test	+	1	0
	-	23	26

Disease group = JIA
 Comparator = ORG
 Index = IgM-RF
Sensitivity = 4.2%
Specificity = 100.0%

Overall

		Reference	
		+	-
Index test	+	19	0
	-	5	26

Disease group = JIA
 Comparator = ORG
 Index = IgG-RF
Sensitivity = 79.2%
Specificity = 100.0%

By subtype

		Reference	
		+	-
Index test	+	1	0
	-	11	26

Disease group = JIA (oligoarticular)
 Comparator = ORG
 Index = IgM-RF
Sensitivity = 8.3%
Specificity = 100.0%

By subtype

		Reference	
		+	-
Index test	+	0	0
	-	9	26

Disease group = JIA (polyarticular)

Comparator = ORG

Index = IgM-RF

Sensitivity = 0%**Specificity** = 100%**By subtype**

		Reference	
		+	-
Index test	+	7	0
	-	2	26

Disease group = JIA (polyarticular)

Comparator = ORG

Index = IgG-RF

Sensitivity = 77.8%**Specificity** = 100%**By subtype**

		Reference	
		+	-
Index test	+	0	0
	-	3	26

Disease group = JIA (systemic)

Comparator = ORG

Index = IgM-RF

Sensitivity = 0%**Specificity** = 100%**By subtype**

		Reference	
		+	-
Index test	+	2	0
	-	1	26

Disease group = JIA (systemic)

Comparator = ORG

Index = IgG-RF

Sensitivity = 66.7%**Specificity** = 100%**By subtype**

		Reference	
		+	-
Index test	+	10	0
	-	2	26

Disease group = JIA (oligoarticular)

Comparator = ORG

Index = IgG-RF

Sensitivity = 83.3%**Specificity** = 100%

Study	Participants Characteristics	Index Test Characteristics
<p>Saulsbury F.T., 1990 (50)</p> <p>Country (# centers): U.S. (NR)</p> <p>Funding: Non-commercial institution</p> <p>Study design: Case control (NR)</p> <p>Recruitment: NR</p>	<p>Inclusion criteria: NR</p> <p>Exclusion criteria: NR</p> <p>JIA patients N_{analyzed}/N_{enrolled}: 50/50</p> <p>Mean/median age (range): NR (1.0-18.0yr)</p> <p>Female: 78.0%</p> <p>Mean/median time since diagnosis (range): NR</p> <p>Subtype: Oligoarticular (56.0%); polyarticular (28.0%); systemic (16.0%)</p> <p>Reference standard for JIA diagnosis: Based on reference No. 7</p>	<p>IgM-RF test Assay method: ELISA</p> <p>Source of antigen: Latex beads coated with human IgG</p> <p>Manufacturer (kit type): Dynatech Laboratories, Alexandria, VA (NR)</p> <p>Positive cutoff: Titer 1:20</p> <p>IgG-RF test Assay method: ELISA</p> <p>Source of antigen: Rabbit IgG</p> <p>Manufacturer (kit type): Gamma Biologicals, Inc. Houston, TX (NR)</p> <p>Positive cutoff: Titer 1:20</p> <p>IgA-RF test Assay method: ELISA Source of antigen: Latex beads coated with human IgG Manufacturer (kit type): Dynatech Laboratories, Alexandria, VA (NR) Positive cutoff: Titer 1:20</p>

Results (Overall)

		Reference	
		+	-
Index test	+	11	1
	-	39	38

Disease group = JIA
 Comparator = Healthy controls
 Index = IgM-RF
Sensitivity = 22.0%
Specificity = 97.4%

Overall

		Reference	
		+	-
Index test	+	2	0
	-	48	39

Disease group = JIA
 Comparator = Healthy controls
 Index = IgG-RF
Sensitivity = 58.0%
Specificity = 94.9%

Overall

		Reference	
		+	-
Index test	+	11	1
	-	39	38

Disease group = JIA
 Comparator = Healthy controls
 Index = IgA-RF
Sensitivity = 32.0%
Specificity = 97.4%

By subtype

		Reference	
		+	-
Index test	+	5	1
	-	23	38

Disease group = JIA (oligoarticular)

Comparator = Healthy controls

Index = IgM-RF

Sensitivity = 17.9%**Specificity** = 97.4%**By subtype**

		Reference	
		+	-
Index test	+	0	0
	-	28	39

Disease group = JIA (oligoarticular)

Comparator = Healthy controls

Index = IgG-RF

Sensitivity = 0%**Specificity** = 100%**By subtype**

		Reference	
		+	-
Index test	+	8	1
	-	20	38

Disease group = JIA (oligoarticular)

Comparator = Healthy controls

Index = IgA-RF

Sensitivity = 28.9%**Specificity** = 97.4%**By subtype**

		Reference	
		+	-
Index test	+	5	1
	-	9	38

Disease group = JIA (polyarticular)

Comparator = Healthy controls

Index = IgM-RF

Sensitivity = 35.7%**Specificity** = 97.4%**By subtype**

		Reference	
		+	-
Index test	+	1	0
	-	13	39

Disease group = JIA (polyarticular)

Comparator = Healthy controls

Index = IgG-RF

Sensitivity = 7.1%**Specificity** = 100%**By subtype**

		Reference	
		+	-
Index test	+	2	1
	-	12	38

Disease group = JIA (polyarticular)

Comparator = Healthy controls

Index = IgA-RF

Sensitivity = 14.3%**Specificity** = 97.4%**By subtype**

		Reference	
		+	-
Index test	+	1	1
	-	7	38

Disease group = JIA (systemic)

Comparator = Healthy controls

Index = IgM-RF

Sensitivity = 12.5%**Specificity** = 97.4%**By subtype**

		Reference	
		+	-
Index test	+	1	0
	-	7	39

Disease group = JIA (systemic)

Comparator = Healthy controls

Index = IgG-RF

Sensitivity = 12.5%**Specificity** = 100%**By subtype**

		Reference	
		+	-
Index test	+	1	1
	-	7	38

Disease group = JIA (systemic)

Comparator = Healthy controls

Index = IgA-RF

Sensitivity = 12.5%**Specificity** = 97.4%

Study	Participants Characteristics	Index Test Characteristics
<p>Siamopoulou-Mavridou, A., 1991 (36)</p> <p>Country (# centers): Greece (NR)</p> <p>Funding: NR</p> <p>Study design: Case control (Prospective)</p> <p>Recruitment: Consecutive patients</p>	<p>Inclusion criteria: Healthy controls: Age- and sex-matched children without rheumatic disease</p> <p>Exclusion criteria: NR</p> <p>JIA patients N_{analyzed}/N_{enrolled}: 69/69</p> <p>Mean age (range): 8.6yr (1.0-15.0yr)</p> <p>Female: 59.4%</p> <p>Mean/median time since diagnosis (range): 5.7yr (1.0-9.0yr)</p> <p>Subtype: Oligoarticular (43.5%); Polyarticular (29.0%); Systemic (27.5%)</p> <p>Reference standard for JIA diagnosis: EULAR</p>	<p>ANA test Assay method: IIF</p> <p>Source of antigen: HEp-2 cell line</p> <p>Manufacturer (kit type): NR</p> <p>Positive cutoff: Titer 1:40</p> <p>IgM-RF test Assay method: Latex fixation test</p> <p>Source of antigen: NR</p> <p>Manufacturer (kit type): Behringwerke AG (NR)</p> <p>Positive cutoff: Mean optical density (OD) of healthy controls + 3 sd</p>

Results (Overall)

		Reference	
		+	-
Index test	+	41	2
	-	28	64

Disease group = JIA
 Comparator = Healthy controls
 Index = ANA
Sensitivity = 59.4%
Specificity = 97.0%

Overall

		Reference	
		+	-
Index test	+	3	0
	-	66	66

Disease group = JIA
 Comparator = Healthy controls
 Index = IgM-RF
Sensitivity = 4.4%
Specificity = 100.0%

By subtype

		Reference	
		+	-
Index test	+	21	2
	-	9	64

Disease group = JIA (oligoarticular)
 Comparator = Healthy controls
 Index = ANA
Sensitivity = 70.0%
Specificity = 97.0%

By subtype

		Reference	
		+	-
Index test	+	13	2
	-	7	64

Disease group = JIA (polyarticular)

Comparator = Healthy controls

Index = ANA

Sensitivity = 65.0%**Specificity** = 97.0%**By subtype**

		Reference	
		+	-
Index test	+	3	0
	-	17	66

Disease group = JIA (polyarticular)

Comparator = Healthy controls

Index = IgM-RF

Sensitivity = 15.0%**Specificity** = 100%**By subtype**

		Reference	
		+	-
Index test	+	7	2
	-	12	64

Disease group = JIA (systemic)

Comparator = Healthy controls

Index = ANA

Sensitivity = 36.8%**Specificity** = 97.0%**By subtype**

		Reference	
		+	-
Index test	+	0	0
	-	19	66

Disease group = JIA (systemic)

Comparator = Healthy controls

Index = IgM-RF

Sensitivity = 0%**Specificity** = 100%**By subtype**

		Reference	
		+	-
Index test	+	0	0
	-	30	66

Disease group = JIA (oligoarticular)

Comparator = Healthy controls

Index = IgM-RF

Sensitivity = 0%**Specificity** = 100%

Study	Participants Characteristics	Index Test Characteristics
<p>Taseski, B., 1981 (45)</p> <p>Country (# centers): Yugoslavia (1)</p> <p>Funding: NR</p> <p>Study design: Case control (Prospective)</p> <p>Recruitment: NR</p>	<p>Inclusion criteria: NR</p> <p>Exclusion criteria: NR</p> <p>JIA patients N_{analyzed}/N_{enrolled}: 40/40</p> <p>Mean/median age (range): NR (Max: 17.0yr)</p> <p>Female: NR</p> <p>Mean/median time since diagnosis (range): NR</p> <p>Subtype: Monoarticular (25.0%); Systemic (15.0%); Polyarticular (60.0%)</p> <p>Reference standard for JIA diagnosis: NR</p> <p>Reference standard for SLE diagnosis: NR</p>	<p>RF test (unspecified isotype) Assay method: Photometrical latex test (PLT)</p> <p>Source of antigen: NR</p> <p>Manufacturer (kit type): NR</p> <p>Positive cutoff: Photometrically measured even at the lowest serum dilution</p> <p>RF test (unspecified isotype) Assay method: Standard sensitized sheep-cell test (SSC)</p> <p>Source of antigen: Sheep cell</p> <p>Manufacturer (kit type): NR</p> <p>Positive cutoff: Titer 1:64</p> <p>IgM-RF test Assay method: Latex slide test (LST)</p> <p>Source of antigen: NR</p> <p>Manufacturer (kit type): NR</p> <p>Positive cutoff: Agglutination visually detected</p>

Results (Overall)

		Reference	
		+	-
Index test	+	20	4
	-	20	20

Disease group = JIA
 Comparator = Healthy controls
 Index = RF (PLT)
Sensitivity = 50.0%

Overall

		Reference	
		+	-
Index test	+	0	0
	-	40	24

Disease group = JIA
 Comparator = Healthy controls
 Index = IgM-RF (LST)
Sensitivity = 0%

Overall

		Reference	
		+	-
Index test	+	1	0
	-	39	24

Disease group = JIA
 Comparator = Healthy controls
 Index = RF (SSC)
Sensitivity = 2.5%

Specificity = 83.3%

Overall

		Reference	
		+	-
Index test	+	9	4
	-	18	20

Disease group = JIA

Comparator = ORG (collagen diseases)

Index = RF (PLT)

Sensitivity = 33.3%

Specificity = 83.3%

Overall

		Reference	
		+	-
Index test	+	2	4
	-	0	20

Disease group = SLE

Comparator = Healthy controls

Index = RF (PLT)

Sensitivity = 100 %

Specificity = 100.0%

Specificity = 100.0%

Overall

		Reference	
		+	-
Index test	+	1	0
	-	26	24

Disease group = JIA

Comparator = ORG (collagen diseases)

Index = IgM-RF (LST)

Sensitivity = 3.7%

Specificity = 100.0%

Specificity = 100.0%

Overall

		Reference	
		+	-
Index test	+	3	0
	-	24	24

Disease group = JIA

Comparator = ORG (collagen diseases)

Index = RF (SSC)

Sensitivity = 11.1%

Specificity = 100.0%

Study	Participants Characteristics	Index Test Characteristics
Varbanova, B.B., 1999 (46) Country (# centers): Bulgaria (NR) Funding: NR Study design: Case control (Prospective) Recruitment: NR	Inclusion criteria: NR Exclusion criteria: NR JIA patients $N_{analyzed}/N_{enrolled}$: 53/53 Mean/median age (range): NR (1.5-18.0yr) Female: NR Mean/median time since diagnosis (range): NR (Min: 1yr) Subtype: Oligoarticular (62.3%); Seronegative Polyarticular (37.7%) Reference standard for JIA diagnosis: EULAR SLE patients $N_{analyzed}/N_{enrolled}$: 22/22 Mean/median age (range): NR Female: NR Mean/median time since diagnosis (range): NR Reference standard for SLE diagnosis: NR	IgM-RF test Assay method: ELISA Source of antigen: Human gamma globulin Manufacturer (kit type): CLB-Amsterdam (NR) Positive cutoff: 6.25 IU IgG-RF test Assay method: ELISA Source of antigen: Rabbit IgG Manufacturer (kit type): NR Positive cutoff: Mean international unit (IU) of healthy controls + 2 sd IgA-RF test Assay method: ELISA Source of antigen: Rabbit IgG Manufacturer (kit type): NR Positive cutoff: Mean IU of healthy controls + 2 sd

Results (Overall)

		Reference	
		+	-
Index test	+	17	0
	-	36	58

Disease group = JIA
 Comparator = Healthy controls
 Index = IgM-RF
Sensitivity = 32.1%
Specificity = 100.0%

Overall

		Reference	
		+	-
Index test	+	7	0
	-	46	58

Disease group = JIA
 Comparator = Healthy controls
 Index = IgG-RF
Sensitivity = 13.2%
Specificity = 100.0%

Overall

		Reference	
		+	-
Index test	+	15	0
	-	38	58

Disease group = JIA
 Comparator = Healthy controls
 Index = IgA-RF
Sensitivity = 28.3%
Specificity = 100.0%

Overall

		Reference	
		+	-
Index test	+	2	0
	-	20	58

Disease group = SLE
 Comparator = Healthy controls
 Index = IgM-RF
Sensitivity = 9.1%
Specificity = 100.0%

By subtype

		Reference	
		+	-
Index test	+	6	0
	-	27	58

Disease group = JIA (oligoarticular)
 Comparator = Healthy controls
 Index = IgM-RF
Sensitivity = 18.2%
Specificity = 100%

By subtype

		Reference	
		+	-
Index test	+	3	0
	-	17	58

Disease group = JIA (polyarticular)
 Comparator = Healthy controls
 Index = IgG-RF
Sensitivity = 15.0%
Specificity = 100%

Overall

		Reference	
		+	-
Index test	+	3	0
	-	19	58

Disease group = SLE
 Comparator = Healthy controls
 Index = IgG-RF
Sensitivity = 13.6%
Specificity = 100%

By subtype

		Reference	
		+	-
Index test	+	11	0
	-	9	58

Disease group = JIA (polyarticular)
 Comparator = Healthy controls
 Index = IgM-RF
Sensitivity = 55.0%
Specificity = 100%

By subtype

		Reference	
		+	-
Index test	+	7	0
	-	26	58

Disease group = JIA (oligoarticular)
 Comparator = Healthy controls
 Index = IgA-RF
Sensitivity = 21.2%
Specificity = 100%

Overall

		Reference	
		+	-
Index test	+	2	0
	-	20	58

Disease group = SLE
 Comparator = Healthy controls
 Index = IgA-RF
Sensitivity = 9.1%
Specificity = 100%

By subtype

		Reference	
		+	-
Index test	+	4	0
	-	29	58

Disease group = JIA (oligoarticular)
 Comparator = Healthy controls
 Index = IgG-RF
Sensitivity = 12.1%
Specificity = 100%

By subtype

		Reference	
		+	-
Index test	+	8	0
	-	12	58

Disease group = JIA (polyarticular)
 Comparator = Healthy controls
 Index = IgA-RF
Sensitivity = 40.0%
Specificity = 100%

Study		Participants Characteristics		Index Test Characteristics																																														
Wakhlu, A., 2003 (37)		Inclusion criteria: NR Exclusion criteria: NR JIA patients $N_{\text{analyzed}}/N_{\text{enrolled}}$: 148/148 Median age (range): 14.0yr (2.0-26.0yr) Female: 43.2% Mean/median time since diagnosis (range): NR Subtype: Oligoarticular (36.5%); Polyarticular (43.2%); Systemic (20.3%) Reference standard for JIA diagnosis: ARA		ANA test Assay method: IIF Source of antigen: HEp-2 cell line Manufacturer (kit type): NR Positive cutoff: Titer 1:40																																														
Country (# centers): India (1) Funding: Academic institution Study design: Case control (Prospective) Recruitment: NR																																																		
Overall		By subtype		By subtype																																														
<table border="1"> <thead> <tr> <th colspan="2"></th> <th colspan="2">Reference</th> </tr> <tr> <th colspan="2"></th> <th>+</th> <th>-</th> </tr> </thead> <tbody> <tr> <th rowspan="2">Index test</th> <th>+</th> <td>2</td> <td>0</td> </tr> <tr> <th>-</th> <td>146</td> <td>25</td> </tr> </tbody> </table>				Reference				+	-	Index test	+	2	0	-	146	25	<table border="1"> <thead> <tr> <th colspan="2"></th> <th colspan="2">Reference</th> </tr> <tr> <th colspan="2"></th> <th>+</th> <th>-</th> </tr> </thead> <tbody> <tr> <th rowspan="2">Index test</th> <th>+</th> <td>0</td> <td>0</td> </tr> <tr> <th>-</th> <td>54</td> <td>25</td> </tr> </tbody> </table>				Reference				+	-	Index test	+	0	0	-	54	25	<table border="1"> <thead> <tr> <th colspan="2"></th> <th colspan="2">Reference</th> </tr> <tr> <th colspan="2"></th> <th>+</th> <th>-</th> </tr> </thead> <tbody> <tr> <th rowspan="2">Index test</th> <th>+</th> <td>2</td> <td>0</td> </tr> <tr> <th>-</th> <td>62</td> <td>25</td> </tr> </tbody> </table>				Reference				+	-	Index test	+	2	0	-	62	25
		Reference																																																
		+	-																																															
Index test	+	2	0																																															
	-	146	25																																															
		Reference																																																
		+	-																																															
Index test	+	0	0																																															
	-	54	25																																															
		Reference																																																
		+	-																																															
Index test	+	2	0																																															
	-	62	25																																															
Disease group = JIA Comparator = Healthy controls Index = ANA Sensitivity = 1.4% Specificity = 100.0%		Disease group = JIA (oligoarticular) Comparator = Healthy controls Index = ANA Sensitivity = 0% Specificity = 100.0%		Disease group = JIA (polyarticular) Comparator = Healthy controls Index = ANA Sensitivity = 3.1% Specificity = 100.0%																																														
By subtype																																																		
<table border="1"> <thead> <tr> <th colspan="2"></th> <th colspan="2">Reference</th> </tr> <tr> <th colspan="2"></th> <th>+</th> <th>-</th> </tr> </thead> <tbody> <tr> <th rowspan="2">Index test</th> <th>+</th> <td>0</td> <td>0</td> </tr> <tr> <th>-</th> <td>30</td> <td>25</td> </tr> </tbody> </table>				Reference				+	-	Index test	+	0	0	-	30	25																																		
		Reference																																																
		+	-																																															
Index test	+	0	0																																															
	-	30	25																																															
Disease group = JIA (systemic) Comparator = Healthy controls Index = ANA Sensitivity = 0% Specificity = 100.0%																																																		

Study	Participants Characteristics	Index Test Characteristics																										
<p>Wananukul, S., 2005 (30)</p> <p>Country (# centers): Thailand (1)</p> <p>Funding: Non-commercial institution</p> <p>Study design: Case control (Prospective)</p> <p>Recruitment: NR</p>	<p>Inclusion criteria: NRG: Scheduled for elective surgery (adenotonsillectomy, herniorrhaphy or plastic surgery)</p> <p>Exclusion criteria: All participants: Exclude overt autoimmune disease, or conditions associated with abnormal ANA titers (infection, hepatitis and malignancy) or underwent treatment with certain drugs (procainamide, hydralazine, chlorpromazine, etc)</p> <p>All participants: Exclude children aged less than 6 months old</p> <p>SLE patients N_{analyzed}/N_{enrolled}: 46/52 Mean/median age (range): NR (5.0-15.0yr) Female: NR Mean/median time since diagnosis (range): NR Subtype: NR Reference standard for SLE diagnosis: 1997 revised criteria for the classification of SLE</p>	<p>ANA test Assay method: IIF</p> <p>Source of antigen: HEp-2 cell line</p> <p>Manufacturer (kit type): Diasarin, Stillwater, MN (ANAFast kits)</p> <p>Positive cutoff: Titer \geq 1:40</p>																										
<p>Results (Overall)</p> <table border="1"> <thead> <tr> <th colspan="2" rowspan="2"></th> <th colspan="2">Reference</th> </tr> <tr> <th>+</th> <th>-</th> </tr> </thead> <tbody> <tr> <th rowspan="2">Index test</th> <th>+</th> <td>42</td> <td>15</td> </tr> <tr> <th>-</th> <td>4</td> <td>84</td> </tr> </tbody> </table> <p>Disease group = SLE Comparator = Healthy controls (elective surgery) Index = ANA Sensitivity = 91.3% Specificity = 84.8%</p>			Reference		+	-	Index test	+	42	15	-	4	84	<p>Overall</p> <table border="1"> <thead> <tr> <th colspan="2" rowspan="2"></th> <th colspan="2">Reference</th> </tr> <tr> <th>+</th> <th>-</th> </tr> </thead> <tbody> <tr> <th rowspan="2">Index test</th> <th>+</th> <td>42</td> <td>17</td> </tr> <tr> <th>-</th> <td>4</td> <td>91</td> </tr> </tbody> </table> <p>Disease group = SLE Comparator = Healthy controls Index = ANA Sensitivity = 91.3% Specificity = 84.3%</p>			Reference		+	-	Index test	+	42	17	-	4	91	
			Reference																									
		+	-																									
Index test	+	42	15																									
	-	4	84																									
		Reference																										
		+	-																									
Index test	+	42	17																									
	-	4	91																									

Study	Participants Characteristics	Index Test Characteristics
Wernick, R., 1981 (47)	<p>Inclusion criteria: NR</p> <p>Exclusion criteria: NR</p> <p>JIA patients N_{analyzed}/N_{enrolled}: 49/49 Mean/median age (range): NR Female: NR Mean/median time since diagnosis (range): NR Subtype: Polyarticular (22.4%); Oligoarticular (59.2%); Systemic (18.4%) Reference standard for JIA diagnosis: ARA</p> <p>SLE patients N_{analyzed}/N_{enrolled}: 7/7 Mean/median age (range): NR Female: NR Mean/median time since diagnosis (range): NR Reference standard for SLE diagnosis: ARA</p>	<p>IgM-RF test Assay method: Solid phase radioimmunoassay</p> <p>Source of antigen: Rabbit and human IgG</p> <p>Manufacturer (kit type): Signma Chemical Co., St. Louis, MO (Cohn Fraction II)</p> <p>Positive cutoff: Mean of normal controls + 2 SD</p> <p>IgG-RF test Assay method: Solid phase radioimmunoassay</p> <p>Source of antigen: Rabbit and human IgG</p> <p>Manufacturer (kit type): Signma Chemical Co., St. Louis, MO (Cohn Fraction II)</p> <p>Positive cutoff: Mean of normal controls + 2 SD</p>

Results (Overall)

		Reference	
		+	-
Index test	+	2	2
	-	47	30

Disease group = JIA
 Comparator = NRG (Scoliosis and neurologic diseases)
 Index = IgM-RF
Sensitivity = 4.1%
Specificity = 93.8%

Overall

		Reference	
		+	-
Index test	+	4	1
	-	45	32

Disease group = JIA
 Comparator = NRG (Scoliosis and neurologic diseases)
 Index = IgG-RF
Sensitivity = 8.2%
Specificity = 90.6%

Overall

		Reference	
		+	-
Index test	+	0	1
	-	7	32

Disease group = SLE
 Comparator = NRG (scoliosis and neurologic diseases)
 Index = IgM-RF
Sensitivity = 0%
Specificity = 97.0%

Overall

		Reference	
		+	-
Index test	+	1	2
	-	6	30

Disease group = SLE
 Comparator = NRG (Scoliosis and neurologic diseases)
 Index = IgG-RF
Sensitivity = 14.3%
Specificity = 93.8%

By subtype

		Reference	
		+	-
Index test	+	0	2
	-	9	30

Disease group = JIA (systemic)
 Comparator = NRG (Scoliosis and neurologic diseases)
 Index = IgM-RF
Sensitivity = 0%
Specificity = 93.8%

By subtype

		Reference	
		+	-
Index test	+	1	1
	-	8	32

Disease group = JIA (systemic)
 Comparator = NRG (Scoliosis and neurologic diseases)
 Index = IgG-RF
Sensitivity = 11.1%
Specificity = 90.6%

By subtype

		Reference	
		+	-
Index test	+	1	2
	-	28	30

Disease group = JIA (oligoarticular)
 Comparator = NRG (Scoliosis and neurologic diseases)
 Index = IgM-RF
Sensitivity = 3.4%
Specificity = 93.8%

By subtype

		Reference	
		+	-
Index test	+	2	1
	-	27	32

Disease group = JIA (oligoarticular)
 Comparator = NRG (Scoliosis and neurologic diseases)
 Index = IgG-RF
Sensitivity = 6.9%
Specificity = 97.0%

By subtype

		Reference	
		+	-
Index test	+	1	2
	-	10	30

Disease group = JIA (polyarticular)
 Comparator = NRG (Scoliosis and neurologic diseases)
 Index = IgM-RF
Sensitivity = 9.1%
Specificity = 93.8%

By subtype

		Reference	
		+	-
Index test	+	1	1
	-	10	32

Disease group = JIA (polyarticular)
 Comparator = NRG (Scoliosis and neurologic diseases)
 Index = IgG-RF
Sensitivity = 9.1%
Specificity = 97.0%

Appendix G. Subgroup Analyses by Onset Type of Juvenile Idiopathic Arthritis

Subgroup analysis of studies examining antinuclear antibody test for juvenile idiopathic arthritis (JIA; Figures G1 – G3). None of the studies provided subgroup data based on RF positivity for patients with polyarticular JIA.

Figure G1. Forest plot of sensitivity and specificity of antinuclear antibody test for oligoarticular juvenile idiopathic arthritis

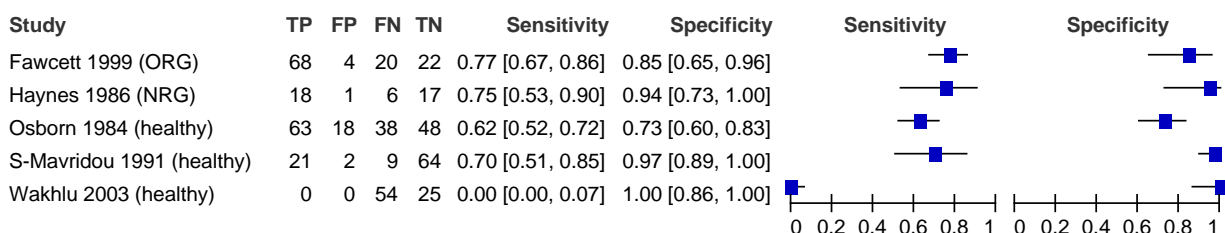


Figure G2. Forest plot of sensitivity and specificity of antinuclear antibody test for polyarticular juvenile idiopathic arthritis

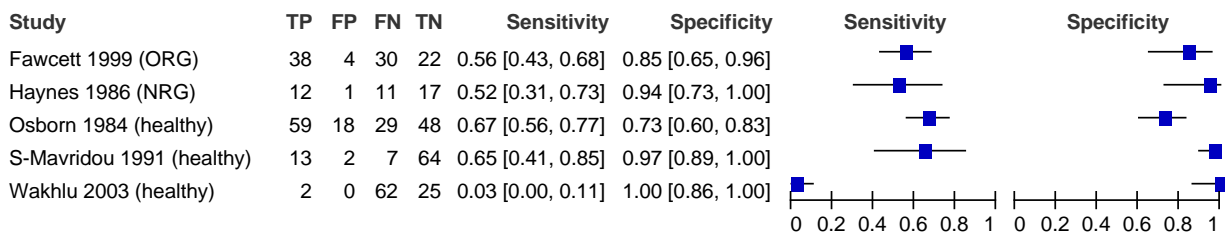
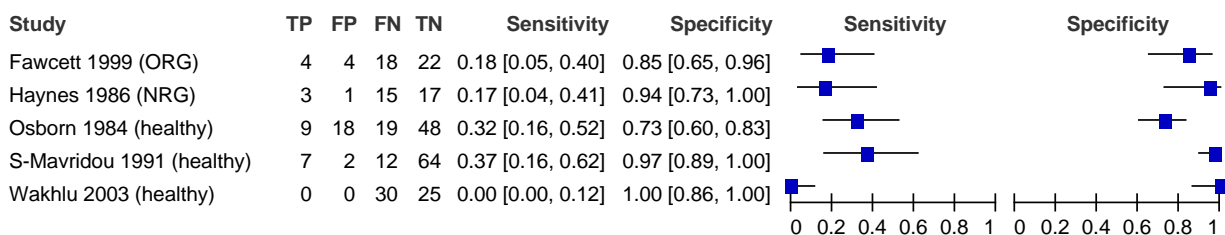


Figure G3. Forest plot of sensitivity and specificity of antinuclear antibody test for systemic juvenile idiopathic arthritis



Subgroup analysis of studies examining IgM-RF test for JIA (Figures G4–G6). None of the studies provided subgroup data based on RF positivity for patients with polyarticular JIA.

Figure G4. Forest plot of the sensitivity and specificity of IgM-rheumatoid factor test for oligoarticular juvenile idiopathic arthritis

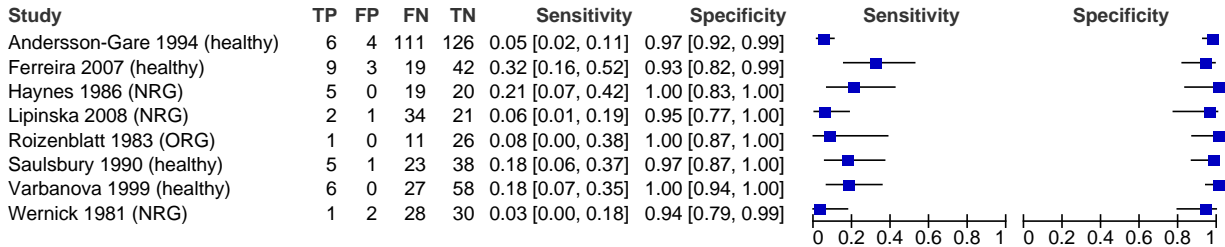


Figure G5. Forest plot of the sensitivity and specificity of IgM-rheumatoid factor test for polyarticular juvenile idiopathic arthritis

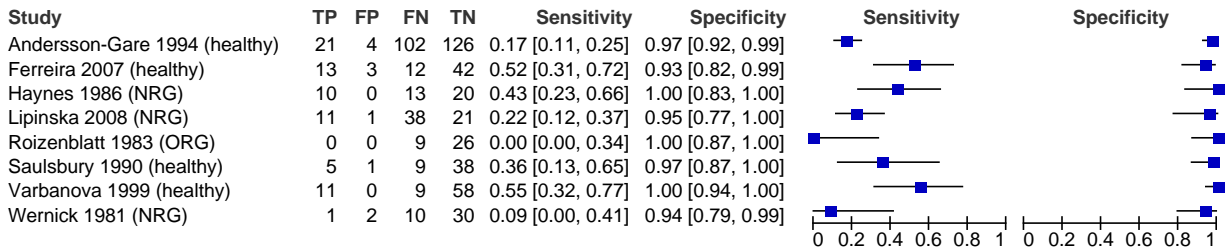
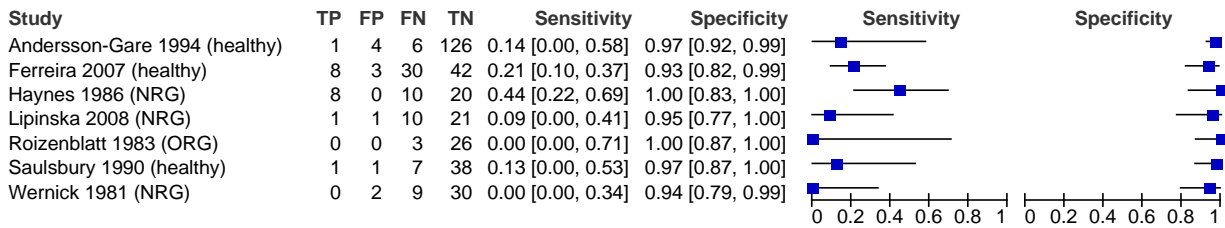


Figure G6. Forest plot of the sensitivity and specificity of IgM-rheumatoid factor test for systemic juvenile idiopathic arthritis



Subgroup analysis of studies examining CCP test for JIA (Figures G7–G9). None of the studies provided subgroup data based on RF positivity for patients with polyarticular JIA.

Figure G7. Forest plot of sensitivity and specificity of cyclic-citrullinated peptide test for oligoarticular juvenile idiopathic arthritis

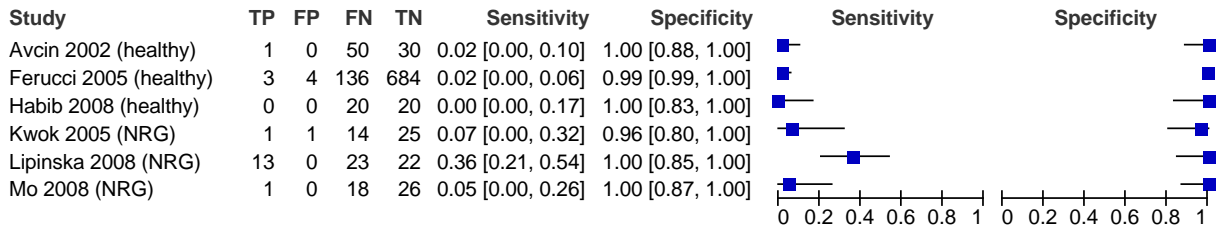


Figure G8. Forest plot of sensitivity and specificity of cyclic-citrullinated peptide test for polyarticular juvenile idiopathic arthritis

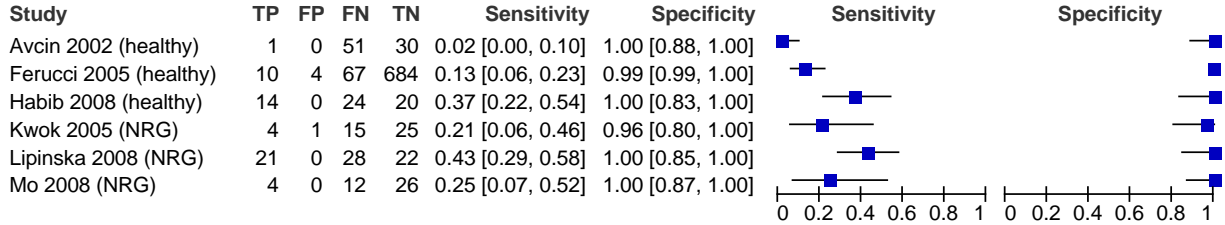


Figure G9. Forest plot of sensitivity and specificity of cyclic-citrullinated peptide test for systemic juvenile idiopathic arthritis

